

CONTRIBUTION TO THE STUDY ON CHEMICAL CONSTITUENTS OF *HEDYOTIS CRASSIFOLIA* L., (RUBIACEAE)

Received 30 May 2006

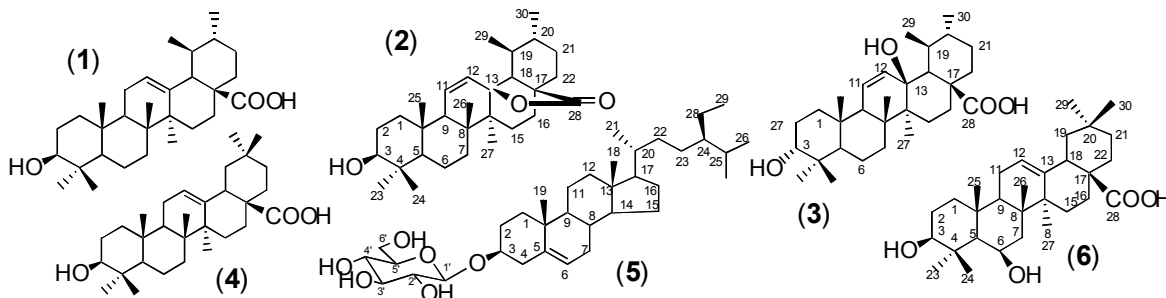
BUI CHI HUU¹, NGUYEN KIM PHI PHUNG²

¹Pedagogic College of Long An

²Department of Chemistry, UNS, National University - Ho Chi Minh City

SUMMARY

Six compounds as ursolic acid (1); 3 β -hydroxyurs-11-ene-28(13)-lactone (2); 3 α ,13 β -dihydroxyurs-11-ene-28-oic acid (3); oleanolic acid (4); 3 β -D-glucopyranosyl- β -sitosterol (5) and 3 β ,6 β -dihydroxyolean-12-ene-28-oic acid (6) were isolated from *Hedyotis crassifolia* L. (Rubiaceae). Among them, (3) was a new triterpen.



I - INTRODUCTION

Hedyotis corymbosa L., *Hedyotis diffusa* Willd, *Hedyotis heynei* R. Br. have widely used in the Asian traditional medicine to cure the inflammation of the liver [1], infected injury [2], snake-bite [3] and especially to cure cancer [4]. Their chemical constituents and pharmaceutical properties have already been reported.

Hedyotis crassifolia L. (Vietnamese name: An điền lá dầy), the plant that is the same genus with these above-mentioned ones but has not yet been studied. In this paper, we report a preliminary result about chemical constituents of this plants growing in Vietnam.



Figure 1: Part of *Hedyotis crassifolia*

Table 1: The comparison (δ ppm) of $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ spectra of compounds **2** and **3** with the authentic sample of $3\beta,13\beta$ -dihydroxyurs-11-ene-28-oic acid (**X**) [5]

C	(X) ($\text{C}_5\text{D}_5\text{N}$) (δ ppm)	2 (DMSO) (δ ppm)	3 (CDCl_3 + CD_3OD)	$^1\text{H-NMR}$ (δ ppm)		
				(X) ($\text{C}_5\text{D}_5\text{N}$)	2 (DMSO)	3 (CDCl_3 + CD_3OD)
1	38.7	38.42	38.20			
2	23.2	22.12	22.67			
3	78.0	76.58	78.53	3.44 (dd; J = 9.0; 7.3 Hz)	3.01 (dd; J = 10; 5.5 Hz)	3.61 (s)
4	39.6	38.94	38.78			
5	55.1	53.84	54.66			
6	18.0	17.30	17.66			
7	31.0	30.90	29.57			
8	42.1	41.06	41.59			
9	53.4	52.25	52.94			
10	36.7	37.03	36.23			
11	133.7	133.02	133.64	5.65 (dd; J = 10.4; 3.0 Hz)	5.49 (dd; J = 10.0; 2.5 Hz)	5.49 (dd; J = 10.0; 3.0 Hz)
12	129.4	128.49	128.50	6.01 (d; J = 10.4 Hz)	5.98 (d; J = 10.0 Hz)	5.93 (d; J = 10.0 Hz)
13	89.4	88.90	90.20			
14	42.3	41.33	41.83			
15	27.9	26.69	26.57			
16	25.9	24.95	25.41			
17	45.2	44.28	45.15			
18	60.6	59.35	60.47			
19	38.7	37.55	38.01			
20	40.4	39.95	40.12			
21	31.6	30.08	30.64			
22	32.0	30.90	31.12			
23	28.4	27.68	27.56	0.84 (s)	0.83 (s)	0.87 (s)
24	16.0	15.24	14.78	1.01 (s)	0.95 (s)	1.00 (s)
25	16.2	15.43	15.97	1.19 (s)	1.12 (s)	1.13 (s)
26	19.2	18.57	18.68	1.22 (s)	1.23 (s)	1.22 (s)
27	18.3	17.60	17.75	1.24 (s)	1.25 (s)	1.22 (s)
28	179.3	178.64	180.64			
29	18.2	17.21	17.57	0.97 (d, J = 6 Hz)	0.93 (d, J = 6 Hz)	0.95 (d., J = 6 Hz)
30	19.4	18.72	18.96	0.83 (d, J = 6 Hz)	0.89 (d, J = 6 Hz)	0.90 (d. J = 6 Hz)

Table 2: The ^1H -, ^{13}C -, DEPT-NMR, HMQC and HMBC (DMSO, 500 MHz) of **6**

C	^{13}C -NMR (δppm)	DEPT	^1H -NMR	HMBC ($^2\text{J}_{\text{CH}}$ and $^3\text{J}_{\text{CH}}$)
1	40.10	-CH ₂ -		
2	27.07	-CH ₂ -		
3	77.14	>CH-OH	2.92 (dd, J = 11.5; 6.5 Hz)	
4	40.05	>C<		
5	55.09	>CH-		C ₆ -OH/C-5 ($^3\text{J}_{\text{CH}}$)
6	66.19	>CH-OH	4.34 (br.s)	C ₆ -OH/C-6 ($^2\text{J}_{\text{CH}}$)
7	38.96	-CH ₂ -		
8	37.96	>C<		
9	47.34	>CH-		
10	36.02	>C<		
11	22.76	-CH ₂ -		
12	121.67	-CH=	5.18 (t, J = 0.2 Hz)	H-12/C-9; H-12/C-14; H-12/C-18
13	143.12	=C<		
14	41.65	>C<		
15	27.07	-CH ₂ -		
16	22.54	-CH ₂ -		
17	45.61	>C<		
18	40.75	>CH-	2.75 (dd, J = 14; 4 Hz)	H-18/C-19 ($^2\text{J}_{\text{CH}}$)
19	45.37	>CH ₂		
20	30.30	>C<		
21	33.25	-CH ₂ -		
22	32.05	-CH ₂ -		
23	27.77	-CH ₃	0.94 (s)	
24	17.13	-CH ₃	1.05 (s)	
25	16.29	-CH ₃	1.21 (s)	
26	17.67	-CH ₃	0.97 (s)	
27	25.55	-CH ₃	1.05 (s)	
28	178.46	-COOH		
29	32.73	-CH ₃	0.87 (s)	
30	23.26	-CH ₃	0.87 (s)	

II - EXPERIMENTAL

1. Plant material

Plants were collected in Long An province in October 2002 and was identified by Dr. Tran Hop, Department of Biology, University of Natural Sciences, National University - Ho Chi

Minh City.

2. Extraction and isolation

Plants were washed, dried, ground into powder and exhaustively extracted by 95° ethanol at room temperature. After evaporating, the ethanolic solution gave crude extract. The crude extract was subjected to silica gel solid

phase extraction [7], and then successively eluted with petroleum ether, chloroform A, chloroform B and methanol. Each fraction of chloroform A, chloroform B and methanol were rechromatographed to afford six compounds.

Compound **1** was isolated from chloroform A. Compounds **2**, **3** and **4** were isolated from chloroform B and compounds **5** and **6** from methanol fraction. Their yield (%) comparing to the dried powder was 0.27, 0.008, 0.001, 0.17, 0.034 and 0.035, respectively.

III - RESULTS AND DISCUSSION

Using modern methods (IR, NMR, MS) and comparing with references, the chemical structures of these compounds were elucidated.

1. Compound 2

White powder. M.p.: 161 - 163°C (methanol). Silica gel TLC with eluent of benzene : chloroform : methanol (1 : 9 : 0.1) revealed by concentrated sulfuric acid gave one lotus-red spot with Rf = 0.70. LC-MS, ESI spectrum showed the [M]⁺ ion peak at *m/z* = 454 corresponded to C₃₀H₄₆O₃. IR(KBr) ν_{\max} cm⁻¹: 3447 (O-H); 1767 (strong, C=O lactone); 1638 (C=C); 1090 (C-O). ¹H-, ¹³C-NMR (DMSO, 500 MHz) were presented in table 1.

2. Compound 3

White powder. M.p.: 231 - 232°C (recrystallized in methanol). Silica gel TLC with eluent of benzene : chloroform : methanol (1 : 9 : 0.1) revealed by concentrated sulfuric acid gave one lotus-red spot with Rf = 0.56. IR(KBr) ν_{\max} cm⁻¹: 3443 (O-H); 1695 (medium, C=O acid); 1639 (C=C); 1217 (C-O). ¹H-, ¹³C-NMR (CDCl₃+CD₃OD, 500 MHz) were presented in table 1. There was a little difference from the standard compound (3 β ,13 β -dihydroxyurs-11-ene-28-oic acid [5]) that compound **3** had a pointed resonant peak of H3 at δ ppm = 3.61, showed that this proton occupied β -position so the hydroxyl group occupied the α -position. These findings substantiated that **3** is 3 α ,13 β -dihydroxyurs-11-ene-28-oic acid and is a new triterpen.

3. Compound 6

White powder. M.p.: 211 - 212°C (methanol). Silica gel TLC with eluent of hexane: ethyl acetate (1:1) revealed by concentrated sulfuric acid gave one dark violet spot with Rf = 0.50. LC-MS, ESI spectrum showed the [M]⁺ peak at *m/z* = 472, suited to the formula C₃₀H₄₈O₄ or C₃₀H₄₇O₃(OH). The spectra also had peaks at *m/z* = 248 (100), 203 (90) that were characteristic peaks of oleanolic acid, so **6** perhaps was an oleanolic acid that contained one more hydroxyl group. ¹H-, ¹³C-, DEPT-NMR, HMQC and HMBC (DMSO, 500 MHz) were presented in table 2 and figure 2. The ¹H-NMR and HMQC showed that the second hydroxyl group is at C-6 (-CH₆-OH at δ ppm = 4.34). The signal was a broad singlet that meant it had the little J, so the hydrogen -C₆H-OH was at the position α and the hydroxyl group -C₆H-OH was at the position β . In conclusion, **6** was 3 β ,6 β -dihydroxyolean-12-ene-28-oic acid (acid sumaresinolic) [6].

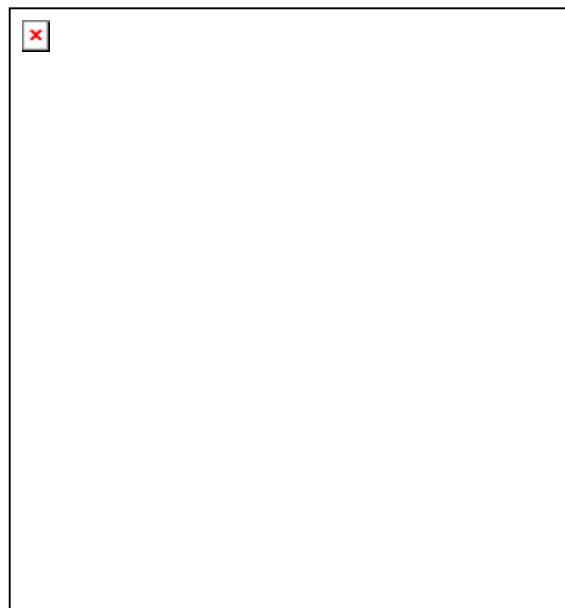


Figure 2: HMBC spectrum of **6**

REFERENCES

1. Y. Yoshida, M. Wang, B. E. Shan, U. Yamashita. International Journal of Immu-

- nopharmacology. 19(7). 359 - 370 (1997).
2. Lai Quang Long, Pham Thanh Ky, Vu Van cien, Chu Dinh Kinh. Magazine of Pharmaceutics, 9, 9 - 11 (2001).
 3. Le Tran Duc. Medicinal plant of Viet Nam, cultivated, processed, initially treatment. Agriculture Publishing House. 1259 (1965).
 4. Huang Xiao Jian. Zhongcaoyao, 27 (7), 408 (1996) (C.A., 125, 257321w).
 5. Huang Hao, Sun Han Dong, Zhao Shou Xun. Phytochemistry, 42(6), 1665 - 1666 (1996)
 6. Shashi B. Mahato, Asish P. Kundu. Phytochemistry, 37(6), 1517 - 1575 (1994)
 7. Richard J.P. Cannell. Natural products isolation. Humana Press, 68 - 74 (1998).