

Triterpenes from *Vitex limonifolia*

Nguyen Thi Kim Thoa², Ninh Khac Ban¹, Do Thi Trang¹, Tran My Linh¹, Vu Huong Giang¹,
Nguyen Xuan Nhiem¹, Phan Van Kiem^{2*}

¹*Institute of Marine Biochemistry, Vietnam Academy of Science and Technology (VAST)*

²*Faculty of Basic Science, Hanoi University of Mining and Geology*

Received 2 December 2017; Accepted for Publication 29 December 2017

Abstract

Using combined chromatographic methods, four triterpenoids, 2 α ,3 α -dihydroxyurs-12-en-28-oic acid (**1**), euscaphic acid (**2**), 2 α ,3 α -dihydroxy-19-oxo-18,19-*seco*-urs-11,13(18)-dien-28-oic acid (**3**), and maslinic acid (**4**) were isolated from the methanol extract of the leaves of *Vitex limonifolia*. Their structures were elucidated by 1D-, 2D-NMR spectra as well as by comparison with those reported in the literature. Compound **3** was reported from *Vitex* genus for the first time.

Keywords. *Vitex limonifolia*, triterpenoid, ursane, oleanane.

1. INTRODUCTION

Vitex is the genus of shrubs and trees and mainly distributed in tropics and subtropics [1]. Since ancient times, civilization used *Vitex* plants for treating many diseases such as malaria, herpes, itches, dermatitis or controlling menstruation [1]. Phytochemical study of the genus *Vitex* revealed the presence of flavonoids, terpenoids, ecdysteroids, iridoid glucosides, etc. [2]. However, phytochemistry study of this plant has not been studied yet. This paper reported the isolation and structure elucidation of four triterpenes from the methanol extract of the leaves of *V. limonifolia*.

2. MATERIAL AND METHODS

2.1. Plant materials

The leaves of *Vitex limonifolia* Wall. ex C.B. Clarke were collected in Bachma National Park, Thua Thien Hue, Vietnam in September 2015, and identified by one of the authors, Prof. Dr. Ninh Khac Ban. A voucher specimen was deposited at the Herbarium Institute of Marine Biochemistry, VAST.

2.2. General experimental procedures

Optical rotations were determined on a Jasco DIP-370 automatic polarimeter. The NMR spectra were recorded using a Bruker DRX 500 spectrometer (¹H-NMR, 500 MHz; ¹³C-NMR, 125 MHz). Column

chromatography was performed using silica-gel (Kieselgel 60, 230-400 mesh, Merck) or RP-18 resins (30-50 μ m, Fujisilisa Chemical Ltd.), and thin layer chromatography (TLC) was performed using a precoated silica gel 60 F₂₅₄ (0.25 mm, Merck) and RP-18 F_{254S} plates (0.25 mm, Merck).

2.3. Extraction and isolation

The dried leaves of *V. limonifolia* (4.2 kg) were extracted with hot MeOH three times (3 \times 5 L) under reflux for 12 h to yield 350 g extract after evaporation of the solvent. This extract was suspended in H₂O and successively partitioned with CH₂Cl₂ and EtOAc to obtain the CH₂Cl₂ (VIL1, 130.0 g), EtOAc (VIL2, 27.0 g), and H₂O (VIL3, 190.0 g) extracts after removal of the solvents *in vacuo*. The VIL1 fraction was chromatographed on a silica gel column eluting with a gradient of *n*-hexane:acetone (100:0 \rightarrow 0:1, v/v) to give six fractions, VIL1A – VIL1F.

The VIL1C fraction was chromatographed on an RP-18 column eluting with MeOH:water (5:1, v/v) to give three fractions, VIL1C1-VIL1C3. VIL1C3 was chromatographed on a silica gel column eluting with *n*-hexane:EtOAc (1.4:1, v/v) to yield **1** (16.0 mg).

The VIL1D fraction was chromatographed on an RP-18 column eluting with MeOH:water (4:1, v/v) to give five fractions, VIL1D1-VIL1D5. VIL1D2 was chromatographed on an RP-18 column eluting

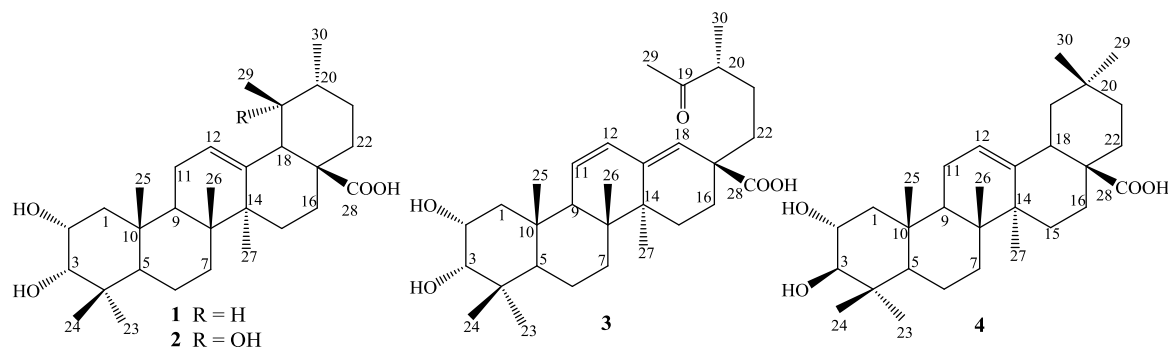


Figure 1: The chemical structures of compounds 1–4

with acetone:water (1.8:1, v/v) to give three smaller fractions, VIL1D2A-VIL1D2C. Compounds **2** (18.0 mg) and **3** (4.5 mg) were obtained from VIL1D2A fraction using silica gel column (*n*-hexane:EtOAc, 1.4:1, v/v as eluent solvent). Compound **4** (7.0 mg) was obtained from VIL1D2C using silica gel column with solvent of *n*-hexane:EtOAc (1.4:1, v/v).

2 α ,3 α -Dihydroxyurs-12-en-28-oic acid (1): white amorphous powder; $[\alpha]_D^{25}$: +30.0 (*c* 0.1, MeOH); ESIMS m/z 473 $[M+H]^+$, $C_{30}H_{48}O_4$; MW: 472; 1H - and ^{13}C -NMR (DMSO- d_6), see table 1.

Euscaphic acid (2): white amorphous powder; $[\alpha]_D^{25}$: +20.0 (*c* 0.1, MeOH); ESIMS m/z 489 $[M+H]^+$, $C_{30}H_{48}O_5$; MW: 488; 1H - and ^{13}C -NMR (DMSO- d_6), see table 1.

2 α ,3 α -Dihydroxy-19-oxo-18,19-*seco*-urs-11,13(18)-dien-28-oic acid (3): white amorphous powder; $[\alpha]_D^{25}$: -40.0 (*c* 0.1, MeOH); ESIMS m/z 487 $[M+H]^+$, $C_{30}H_{46}O_5$; MW: 486; 1H - and ^{13}C -NMR (CD_3OD), see table 1.

Maslinic acid (4): white amorphous powder; $[\alpha]_D^{25}$: -64.1 (*c* 0.1, MeOH); ESIMS m/z 473 $[M+H]^+$, $C_{30}H_{48}O_4$; MW: 472; 1H - and ^{13}C -NMR (CD_3OD), see table 1.

3. RESULTS AND DISCUSSION

Compound **1** was obtained as a white powder. The 1H -NMR spectrum of **1** showed the signals of one olefinic proton at δ_H 5.14 (1H, t, J = 3.0 Hz) and seven methyl groups at δ_H 0.70 (3H, s), 0.78 (3H, s), 0.82 (3H, d, J = 6.5 Hz), 0.88 (3H, s), 0.89 (3H, s), 0.91 (3H, d, J = 7.0 Hz), 1.04 (3H, s), two oximethine protons at δ_H 3.15 (br s) and 3.77 (br d, J = 11.0 Hz). The ^{13}C -NMR and DEPT spectra of **1** displayed the signals of thirty carbons, including seven methyls, eight methylenes, eight methines, and seven quaternary carbons in which two olefinic carbons at δ_C 124.5 and 138.2. Analysis the 1H - and

^{13}C -NMR spectra of **1** suggested the presence of an ursan-12-ene-*type* triterpene skeleton [3]. In addition, the NMR data of **1** was found to be similar to those of 2 α ,3 α -dihydroxyurs-12-en-28-oic acid methyl ester [4] except for the disappearance of methyl ester group. The HMBC correlations between H-23 (δ_H 0.89)/H-24 (δ_H 0.78) and C-3 (δ_C 77.8)/C-4 (δ_C 38.0)/C-5 (δ_C 47.6) indicated the position of the hydroxyl group at C-3. The HMBC correlations between H-1 (δ_H 1.13 and 1.41)/H-3 (δ_H 3.15) and C-2 (δ_C 64.7) indicated the hydroxyl group was located at C-2. Beside, the large coupling constant of H-1 and H-2, J = 11.0 Hz [H-2: δ_H 3.77 (br d, J = 11.0 Hz)] and small coupling constant of H-2 and H-3 (J ~ 0 Hz) [H-3: δ_H 3.15 (br s)] suggested the configurations of two hydroxyl groups at C-2/C-3 as *equatorial* and *axial*, respectively. The HMBC correlations between H-27 (δ_H 1.04) and C-8 (δ_C 39.1)/C-13 (δ_C 138.2)/C-14 (δ_C 41.7)/C-15 (δ_C 28.2); between H-12 (δ_H 5.14) and C-9 (δ_C 46.8)/C-14 (δ_C 41.7)/C-18 (δ_C 52.4) proved the position of the double bond at C-12/C-13. The ESIMS of **1** exhibited an ion peak at m/z 473 $[M+H]^+$, corresponding to the formula of $C_{30}H_{48}O_4$. Consequently, compound **1** was identified as 2 α ,3 α -dihydroxyurs-12-en-28-oic acid and previously reported from *V. altissima* [5] and *V. negundo* [6].

Analysis the NMR spectra of **2** exhibited the structure was similar to those of **1** except for the additional hydroxyl group at C-19. This was confirmed by the HMBC correlations between H-29 (δ_H 1.08) and C-18 (δ_C 53.2)/C-19 (δ_C 71.6)/C-20 (δ_C 41.4); between H-30 (δ_H 0.84) and C-19 (δ_C 71.6)/C-20 (δ_C 41.4)/C-21 (δ_C 25.9). The molecular formula of **2** was further confirmed by the exhibition of an ESIMS ion peak at m/z 489 $[M+H]^+$, corresponding to the formula of $C_{30}H_{48}O_5$. Thus, the structure of **2** was determined to be euscaphic acid [3]. This compound was already isolated from *V. altissima* [5], *V. negundo* [6].

Table 1: The ¹H- and ¹³C-NMR data for compounds 1-4

C	1			2			3			4		
	δ _C [#]	δ _C ^{a,c}	δ _H ^{a,d} (J in Hz)	δ _C [¥]	δ _C ^{a,c}	δ _H ^{a,d} (J in Hz)	δ _C [§]	δ _C ^{b,c}	δ _H ^{b,d} (J in Hz)	δ _C [£]	δ _C ^{b,c}	δ _H ^{b,d} (J in Hz)
1	42.2	41.7	1.13 (m) 1.41 (m)	42.6	41.6	1.15 (m) 1.39 (m)	43.0	42.3	1.32 (m) 1.85 (m)	46.8	48.2	0.92 (m) 1.95 (m)
2	66.7	64.7	3.77 (br d, 11.0)	66.2	64.7	3.77 (br d,11.0)	66.3	67.0	4.00 (dt, 4.0, 11.0)	68.8	69.5	3.64 (dt, 4.5, 10.0)
3	79.2	77.8	3.15 (br s)	79.3	77.9	3.15 (d, 1.0)	79.8	80.1	3.37 (br s)	83.8	84.4	2.93 (d, 10.0)
4	39.3	38.0	-	38.9	38.0	-	39.3	39.5	-	39.1	40.6	-
5	48.3	47.6	1.11 (m)	48.8	47.6	1.15 (dd, 4.0, 13.5)	48.7	49.0	1.36 (m)	55.3	56.7	0.86 (m)
6	18.2	17.6	1.27 (m) 1.35 (m)	18.8	17.7	1.28 (m) 1.35 (m)	18.6	19.0	1.45 (m) 1.54 (m)	18.3	19.6	1.45 (m) 1.58 (m)
7	33.0	32.6	1.25 (m) 1.42 (m)	33.7	32.6	1.21 (m) 1.43 (m)	32.9	33.2	1.30 (m) 1.42 (m)	32.6	33.8	1.52 (m) 1.57 (m)
8	40.0	39.1	-	40.8	39.5	-	41.6	41.9	-	39.1	40.5	-
9	47.6	46.8	1.54 (m)	47.8	46.5	1.66 (m)	55.0	55.3	2.19 (br s)	47.5	49.0	1.65 (m)
10	38.4	37.8	-	38.8	37.8	-	38.8	39.1	-	38.3	39.3	-
11	23.4	22.9	1.85 (m)	24.3	23.1	1.89 (m)	128.0	128.2	5.71 (dd, 1.5, 10.5)	23.5	24.6	1.92 (m) 1.96 (m)
12	125.8	124.5	5.14 (t, 3.0)	128.7	126.8	5.17 (br s)	130.9	131.3	6.01 (dd, 3.0, 10.5)	122.0	123.5	5.27 (t, 3.5)
13	138.7	138.2	-	139.6	138.6	-	142.9	144.0	-	143.6	145.3	-
14	42.4	41.7	-	42.3	41.2	-	41.9	42.4	-	41.7	42.9	-
15	28.2	27.4	0.98 (m) 1.80 (m)	29.1	28.0	0.89 (m) 1.69 (m)	26.9	27.1	1.16 1.82	27.6	28.8	1.10 (m) 1.79 (m)
16	24.4	23.8	1.52 (m) 1.93 (m)	26.3	25.2	1.39 (m) 2.49 (m)	27.8	27.9	1.48 (m) 2.20 (m)	23.1	24.0	1.62 (m) 2.03 (m)
17	48.3	46.9	-	48.3	46.9	-	47.9	48.5	-	46.6	47.6	-
18	53.2	52.4	2.11 (d, 11.5)	54.4	53.2	2.37 (s)	129.2	128.4	5.41 (s)	41.3	42.7	2.87 (dd, 4.0, 14.0)
19	39.1	38.5	1.31 (m)	73.2	71.6	-	211.8	215.1	-	45.8	47.2	1.16 (m) 1.51 (m)
20	38.5	38.4	0.94 (m)	42.5	41.4	1.13 (m)	47.8	48.7	2.56 (m)	30.7	31.6	-
21	30.8	30.2	1.28 (m) 1.43 (m)	27.0	25.9	1.12 (m) 1.61 (m)	28.6	28.7	1.34 (m) 1.70 (m)	33.8	34.9	1.22 (m) 1.42 (m)
22	36.8	36.3	1.51 (m) 1.59 (m)	38.4	37.3	1.50 (m) 1.59 (m)	39.4	39.4	1.42 (m) 1.69 (m)	32.3	33.9	1.36 (m) 1.76 (m)
23	28.6	28.9	0.89 (s)	29.5	28.9	0.88 (s)	29.7	29.1	1.01 (s)	28.6	29.3	1.04 (s)
24	22.0	21.9	0.78 (s)	22.3	21.8	0.78 (s)	22.1	21.8	0.88 (s)	16.8	17.7	0.84 (s)
25	16.5	16.2	0.88 (s)	16.7	16.1	0.88 (s)	19.6	20.3	0.99 (s)	16.8	17.5	0.83 (s)
26	17.1	17.0	0.70 (s)	17.4	16.6	0.68 (s)	17.2	17.1	0.75 (s)	16.8	17.1	1.03 (s)
27	23.9	23.2	1.04 (s)	24.7	24.1	1.29 (s)	20.4	19.4	1.00 (s)	26.0	26.4	1.18 (s)
28	178.4	178.3	-	179.4	179.0	-	178.3	178.5	-	178.0	181.8	-
29	17.1	16.9	0.82 (d, 6.5)	27.2	26.4	1.08 (s)	28.4	28.3	2.16 (s)	33.1	33.6	0.93 (s)
30	21.2	21.1	0.91 (d, 7.0)	16.6	16.3	0.84 (d, 6.5)	16.7	16.5	1.10 (d, 7.0)	23.5	24.0	0.96 (s)

^arecorded in DMSO-*d*₆, ^bCD₃OD, ^c125MHz, ^d500MHz, [#]δ_C of methyl 2α,3α-dihydroxy-urs-12-en-28-oate [4], [¥]δ_C of euscaphic acid [3], [§]δ_C of 2α,3α-dihydroxy-19-oxo-18,19-*seco*-urs-11,13(18)-diene-28-oic acid [7], [£]δ_C of maslinic acid [8].

Compound **3** was obtained as a white powder. The $^1\text{H-NMR}$ spectrum of **3** showed the signals of three olefinic protons at δ_{H} 5.41 (s), 5.71 (dd, $J = 1.5, 10.5$ Hz), and 6.01 (dd, $J = 3.0, 10.5$ Hz), two hydroxymethine protons at δ_{H} 3.37 (br s) and 4.00 (dt, $J = 4.0, 11.0$ Hz), and six methyl groups at δ_{H} 0.75 (s), 0.88 (s), 0.99 (d, $J = 6.5$ Hz), 1.00 (s), 1.01 (s), 1.10 (d, $J = 7.0$ Hz), and 2.16 (s). The $^{13}\text{C-NMR}$ and DEPT spectra of **3** displayed the signals of 30 carbons, including seven methyls at δ_{C} 16.5, 17.1, 19.4, 20.3, 21.8, 28.3, 29.1 and four olefinics at δ_{C} 128.2, 128.4, 131.3, 144.0 and one carbonyl carbon at δ_{C} 178.5. Analysis the $^1\text{H-}$ and $^{13}\text{C-NMR}$ spectra of **3** suggested **3** was very similar to those of $2\alpha,3\alpha$ -dihydroxy-19-oxo-18,19-*seco*-urs-11,13(18)-dien-28-oic acid [7]. The HMBC correlations between H-23 (δ_{H} 1.01)/H-24 (δ_{H} 0.88) and C-3 (δ_{C} 80.1); between H-3 (δ_{H} 3.37) and C-2 (δ_{C} 67.0) indicated the position of the hydroxyl group at C-3. The HMBC correlations indicated the hydroxyl group was located at C-2. The HMBC correlations from H-27 (δ_{H} 1.00) to C-8 (δ_{C} 41.9)/C-13 (δ_{C} 144.0)/C-14 (δ_{C} 42.4)/C-15 (δ_{C} 27.1); from H-12 (δ_{H} 6.01) to C-9 (δ_{C} 55.3)/C-11 (δ_{C} 128.2)/C-13 (δ_{C} 144.0)/C-18 (δ_{C} 128.4); and from H-18 (δ_{H} 5.41) to C-12 (δ_{C} 131.3)/C-14 (δ_{C} 42.4)/C-16 (δ_{C} 27.9)/C-17 (δ_{C} 48.5)/C-22 (δ_{C} 39.4) proved the location of two the double bond at C-11/C-12 and C-13/C-18. The HMBC correlations between H-29 (δ_{H} 2.16) and C-19 (δ_{C} 215.1)/C-20 (δ_{C} 48.7); between H-30 (δ_{H} 1.10) and C-19 (δ_{C} 215.1)/C-20 (δ_{C} 47.8)/C-21 (δ_{C} 28.7) proved the oxo group at C-19. All NMR assignments

of **3** were confirmed by detailed analyses of HSQC and HMBC spectra (table 1), which are in good agreement with those reported for $2\alpha,3\alpha$ -dihydroxy-19-oxo-18,19-*seco*-urs-11,13(18)-dien-28-oic acid in the literature [7], and further confirmed by the exhibition of an ESIMS ion peak at m/z 487 $[\text{M}+\text{H}]^+$, corresponding to the formula of $\text{C}_{30}\text{H}_{46}\text{O}_5$. Thus, compound **3** was identified as $2\alpha,3\alpha$ -dihydroxy-19-oxo-18,19-*seco*-urs-11,13(18)-dien-28-oic acid.

Analysis the NMR and mass spectra of **4** indicated that the structure of **4** was similar to those of maslinic acid [8]. The HMBC correlations between two methyl groups H-23 (δ_{H} 1.04)/H-24 (δ_{H} 0.84) and C-3 (δ_{C} 84.4); between H-3 (δ_{H} 2.93) and C-2 (δ_{C} 69.5) indicated two hydroxyl groups located at C-3 and C-2. The HMBC correlations between two methyl groups H-29 (δ_{H} 0.93)/H-30 (δ_{H} 0.96) and C-19 (δ_{C} 47.2)/C-20 (δ_{C} 31.6)/C-21 (δ_{C} 34.9) confirmed the location of two methyl groups at C-20. The HMBC correlations between H-27 (δ_{H} 1.18) and C-8 (δ_{C} 40.5)/C-14 (δ_{C} 42.9)/C-15 (δ_{C}

28.8) proved the double bond at C-12/C-13. Consequently, **4** was elucidated as maslinic acid [8] and previously reported from *V. altissima* [5], and *V. negundo* [9].

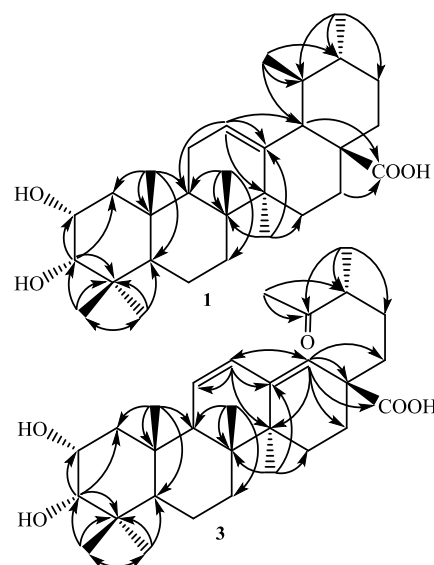


Figure 2: The key HMBC correlations of **1** and **3**

Acknowledgment. This research is funded by Vietnam National Foundation for Science and Technology Development (NAFOSTED) under grant number 104.01-2014.02.

REFERENCES

1. A. Rani, A. Sharma. *The genus Vitex: a review*. Pharmacognosy Reviews, **7**, 188-198 (2013).
2. S. Ganapaty, K. N. Vidyadhar. *Phytoconstituents and biological activities of Vitex-a review*. Journal of Natural Remedies, **5**, 75-95 (2005).
3. J. -J. Cheng, L. -J. Zhang, H. -L. Cheng, C. -T. Chiou, I. J. Lee, Y.-H. Kuo. *Cytotoxic hexacyclic triterpene acids from Euscaphis japonica*, Journal of Natural Products, **73**, 1655-1658 (2010).
4. Hisashi Kojima, H. Ogura. *Triterpenenoids from Prunella vulgaris*, Phytochemistry, **25**(3), 729-733 (1986).
5. C. Sridhar, K. V. Rao, G. V. Subbaraju. *Flavonoids, triterpenoids and a lignan from Vitex altissima*, Phytochemistry, **66**, 1707-1712 (2005).
6. M. M. Li, X. Q. Su, J. Sun, Y. F. Gu, Z. Huang, K. W. Zeng, Q. Zhang, Y. F. Zhao, D. Ferreira, J. K. Zjawiony, J. Li, P. F. Tu. *Anti-inflammatory ursane- and oleanane-type triterpenoids from Vitex negundo var. cannabifolia*, Journal of Natural Products, **77**, 2248-2254 (2014).
7. H. Ping, K. Gloria, W. Shanxin, W. Peter. *Triterpene acids from Toddalia asiatica*, Natural Product Research and Development, **17**, 404-408 (2005).

8. S. Taniguchi, Y. Imayoshi, E. Kobayashi, Y. Takamatsu, H. Ito, T. Hatano, H. Sakagami, H. Tokuda, H. Nishino, D. Sugita, S. Shimura, T. Yoshida. *Production of bioactive triterpenes by Eriobotrya japonica calli*, *Phytochemistry*, **59**, 315-323 (2002).
9. C. -J. Zheng, B. -K. Huang, Y. -B. Wu, T. Han, Q. -Y. Zhang, H. Zhang, L. -P. Qin. *Terpenoids from Vitex negundo seeds*, *Biochemical Systematics and Ecology*, **38**, 247-249 (2010).

Corresponding author: **Phan Van Kiem**

Institute of Marine Biochemistry
Vietnam Academy of Science and Technology
18, Hoang Quoc Viet road, Cau Giay district, Hanoi, Viet Nam
E-mail: phankiem@yahoo.com.