

Structure elucidation of seven steroids from *Sinularia conferta*

Ninh Thi Ngoc^{1,2}, Pham Thi Mai Huong¹, Nguyen Thi Phuong Chi¹, Do Cong Thung³,
Nguyen Xuan Cuong¹, Nguyen Van Thanh¹, Nguyen Hoai Nam^{1*}, Chau Van Minh¹

¹Advanced Center for Bioorganic Chemistry, Institute of Marine Biochemistry,
Vietnam Academy of Science and Technology (VAST)

²Graduate University of Science and Technology, VAST

³Institute of Marine Environment and Resources, VAST

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Abstract

Seven steroids were isolated from the methanol extract of the soft corals *Sinularia conferta*. These steroids were elucidated as 7 α -methoxyergosta-5,24(28)-diene-3 β -ol (1), ergosta-5-ene-3 β ,7 α -diol (2), 3 β ,7 α -dihydroxyergosta-5,24(28)-diene (3), 3 β -hydroxyergosta-5,24(28)-diene-7-one (4), ergosta-24(28)-ene-3 β ,5 α ,6 β -triol-6-acetate (5), ergosta-24(28)-ene-3 β ,5 α ,6 β -triol (6), and ergosta-3 β ,5 α ,6 β -triol (7) by 1D and 2D-NMR experiments and comparison with reported data.

Keywords. *Sinularia conferta*, Alcyoniidae, soft coral, steroid.

1. INTRODUCTION

The phylum Cnidaria are traditionally divided into three classes: Anthozoa, Hydrozoa, and Scyphozoa. The best-known representatives of the class Anthozoa are the gorgonians and soft corals. Soft corals attracted considerable attention because of the wide range of their bioactive secondary metabolites. Among these marine invertebrates, genus *Sinularia* is one of the most widely distributed soft corals and constitutes a dominant portion of the biomass in the tropical reef environment [1, 2]. Prior investigations demonstrated that *Sinularia* species are a rich source of steroids and terpenoids [2-5].

As a part of our recent investigations on chemical constituents and biological activities of Vietnamese *Sinularia* soft corals, we have recently reported twelve steroids from the *Sinularia conferta* species and their *in vitro* cytotoxicity [6]. The current paper deals with detailed structure elucidation of seven steroids from this species.

2. EXPERIMENTAL

2.1. General experimental procedures

The ¹H-NMR (500 MHz) and ¹³C-NMR (125 MHz) spectra were recorded on a Bruker AM500 and AVANCE III HD 500 FT-NMR spectrometers with

TMS used as an internal standard. Medium pressure liquid chromatography (MPLC) was carried out on a Biotage - Isolera One system (SE-751 03 Uppsala, Sweden). Column chromatography (CC) was performed on silica gel (Kieselgel 60, 70–230 mesh and 230-400 mesh, Merck, Darmstadt, Germany), YMC*GEL (ODS-A, 12 nm S-150 mm, YMC Co., Ltd., Japan), Sephadex LH-20 (Sigma-Aldrich, USA), and Diaion HP-20 (Supelco, USA) resins. Thin layer chromatography (TLC) used pre-coated silica gel 60 F₂₅₄ (1.05554.0001, Merck) and RP-18 F_{254S} plates (1.15685.0001, Merck). Compounds were visualized by spraying with aqueous 10 % H₂SO₄ and heating for 3-5 minutes.

2.2. Marine materials

The samples of *S. conferta* (Dana, 1846) were collected near the Con Co island, Quangtri, Vietnam, in May 2015, and identified by Prof. Do Cong Thung, Institute of Marine Environment and Resources, VAST. A voucher specimen (SCO-052015) was deposited at the Institute of Marine Environment and Resources and Institute of Marine Biochemistry, VAST, Vietnam.

2.3. Isolation

Dried bodies of the soft coral *S. conferta* (2.5 kg) were extracted three times with methanol (5 L each)

under ultrasonic condition. The resulting solutions were filtered, combined, and concentrated under reduced pressure to give the methanol residue (SCO.M, 150.0 g), which was suspended in water

and extracted in turn with *n*-hexane and CH₂Cl₂ resulting in extracts of *n*-hexane (SCO.H, 100.0 g), CH₂Cl₂ (SCO.D, 6.0 g), and water layer.

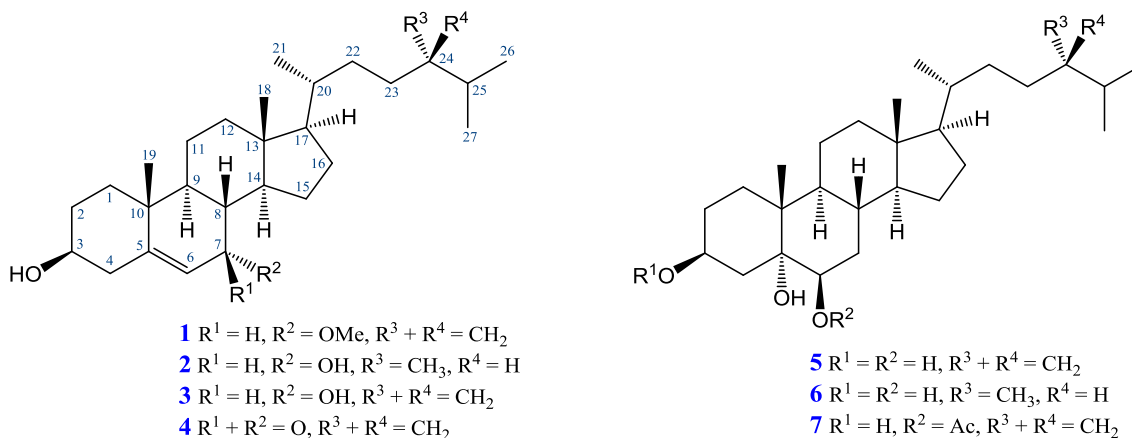


Figure 1: Chemical structures of compounds 1-7

Table 1: ¹H-NMR (500 MHz, CDCl₃) and ¹³C-NMR (125 MHz, CDCl₃) data of 1 and 2

| C | ^a δ _C | 1 | | ^b δ _C | 2 | |
|-----|-----------------------------|----------------|-------------------------------|-----------------------------|----------------|-------------------------------|
| | | δ _C | δ _H mult. (J = Hz) | | δ _C | δ _H mult. (J = Hz) |
| 1 | 37.1 | 36.77 | 1.16 m/1.83 m | 37.6 | 37.04 | 1.12 m/1.86 m |
| 2 | 31.4 | 31.49 | 1.52 m/1.85 m | 32.4 | 30.44 | 0.95 m/1.38 m |
| 3 | 71.2 | 71.46 | 3.62 m | 71.0 | 71.38 | 3.59 m |
| 4 | 42.1 | 42.33 | 2.30 m/2.33 m | 43.4 | 42.04 | 2.28 m/2.33 m |
| 5 | 146.2 | 146.15 | - | 144.9 | 146.26 | - |
| 6 | 121.5 | 120.78 | 5.73 dd (1.5, 4.0) | 125.5 | 123.90 | 5.61 dd (1.5, 5.0) |
| 7 | 73.7 | 73.92 | 3.29 br s | 64.8 | 65.38 | 3.85 br s |
| 8 | 37.2 | 37.24 | 1.49 m | 38.4 | 37.56 | 1.48 m |
| 9 | 42.4 | 42.76 | 1.32 m | 42.7 | 42.30 | 1.23 m |
| 10 | 37.1 | 37.47 | - | 37.7 | 37.42 | - |
| 11 | 20.6 | 20.84 | 1.45 m/1.50 m | 21.2 | 20.73 | 1.50 m/1.54 m |
| 12 | 38.9 | 39.09 | 1.18 m/2.17 m | 39.8 | 39.10 | 1.17 m/2.00 m |
| 13 | 42.0 | 42.16 | - | 42.3 | 42.16 | - |
| 14 | 48.8 | 49.09 | 1.50 m | 50.2 | 49.44 | 1.44 m |
| 15 | 24.1 | 24.27 | 0.98 m/1.63 m | 24.7 | 24.32 | 1.05 m/1.73 m |
| 16 | 28.1 | 28.23 | 1.28 m/1.90 m | 28.7 | 28.24 | 1.31 m/1.90 m |
| 17 | 55.5 | 55.67 | 1.21 m | 56.0 | 55.70 | 1.02 m |
| 18 | 11.4 | 11.49 | 0.67 s | 12.0 | 11.65 | 0.68 s |
| 19 | 18.1 | 18.27 | 0.99 s | 18.5 | 18.26 | 1.00 s |
| 20 | 35.6 | 35.77 | 1.43 m | 36.5 | 36.16 | 1.38 m |
| 21 | 18.6 | 18.76 | 0.95 d (6.5) | 19.2 | 18.93 | 0.93 d (6.5) |
| 22 | 34.7 | 34.72 | 1.16 m/1.55 m | 34.0 | 33.73 | 0.98 m/1.40 m |
| 23 | 30.9 | 30.91 | 1.88 m/2.10 m | 30.8 | 31.41 | 1.52 m/1.86 m |
| 24 | 156.7 | 156.97 | - | 39.3 | 39.20 | 1.21 m |
| 25 | 33.7 | 33.86 | 2.23 m | 31.7 | 31.52 | 1.58 m |
| 26 | 21.8 | 21.89 | 1.02 d (6.5) | 17.7 | 17.66 | 0.79 d (6.5) |
| 27 | 21.9 | 22.03 | 1.03 d (6.5) | 20.7 | 20.50 | 0.86 d (6.5) |
| 28 | 105.8 | 105.92 | 4.44 s/4.71 s | 15.6 | 15.49 | 0.78 d (6.5) |
| OMe | 56.6 | 56.78 | 3.35 s | | | |

^aδ_C of 7α-methoxyergosta-5,24(28)-diene-3β-ol in CDCl₃ [7], ^bδ of ergost-5-en-3β,7α-diol in pyridine-*d*₅ [8].

Table 2: ¹H-NMR (500 MHz, CD₃OD) and ¹³C-NMR (125 MHz, CD₃OD) data of **5** and **6**

| C | ^a δ _C | 5 | | ^b δ _C | 6 | |
|----|-----------------------------|----------------|-------------------------------|-----------------------------|----------------|-------------------------------|
| | | δ _C | δ _H mult. (J = Hz) | | δ _C | δ _H mult. (J = Hz) |
| 1 | 33.3 | 33.49 | 1.36 m/1.61 m | 33.3 | 33.48 | 1.35 m/1.60 m |
| 2 | 32.5 | 31.69 | 1.52 m/1.78 m | 32.5 | 31.62 | 1.00 m/1.50 m |
| 3 | 67.4 | 68.33 | 4.03 m | 67.4 | 68.33 | 4.02 m |
| 4 | 42.9 | 41.44 | 1.20 m/2.08 m | 42.9 | 41.44 | 1.54 m/2.08 m |
| 5 | 75.9 | 76.81 | - | 75.9 | 76.82 | - |
| 6 | 76.3 | 76.53 | 3.47 br s | 76.3 | 76.53 | 3.47 br s |
| 7 | 35.7 | 35.28 | 1.55 m/1.72 m | 35.7 | 35.29 | 1.52 m/1.71 m |
| 8 | 31.4 | 31.62 | 1.74 m | 31.2 | 31.69 | 1.71 m |
| 9 | 45.9 | 46.57 | 1.39 m | 45.9 | 46.58 | 1.39 m |
| 10 | 39.1 | 39.31 | - | 39.2 | 39.31 | - |
| 11 | 21.8 | 22.31 | 1.39 m | 21.8 | 22.30 | 1.40 m |
| 12 | 40.7 | 41.50 | 1.56 m/2.03 m | 40.7 | 41.49 | 1.19 m/2.03 m |
| 13 | 43.1 | 43.96 | - | 43.1 | 43.91 | - |
| 14 | 56.5 | 57.54 | 1.16 m | 56.5 | 57.55 | 1.13 m |
| 15 | 24.6 | 25.22 | 1.13 m/1.63 m | 24.6 | 25.22 | 1.12 m/1.60 m |
| 16 | 28.6 | 29.33 | 1.30 m/1.88 m | 28.6 | 29.31 | 1.31 m/1.87 m |
| 17 | 56.6 | 57.45 | 1.13 m | 56.6 | 57.46 | 1.13 m |
| 18 | 12.4 | 12.62 | 0.74 s | 12.4 | 12.50 | 0.73 s |
| 19 | 17.2 | 17.31 | 1.18 s | 17.2 | 17.20 | 1.17 s |
| 20 | 36.1 | 36.99 | 1.45 m | 36.6 | 37.53 | 1.40 m |
| 21 | 18.9 | 19.21 | 0.99 d (6.5) | 19.1 | 19.20 | 0.96 d (6.5) |
| 22 | 35.1 | 36.02 | 1.17 m/1.58 m | 34.0 | 34.93 | 1.00 m/1.46 m |
| 23 | 31.2 | 32.11 | 1.93 m/2.14 m | 31.0 | 30.80 | 1.31 m |
| 24 | 156.7 | 157.78 | - | 39.4 | 40.44 | 1.24 m |
| 25 | 34.1 | 34.91 | 2.25 m | 31.7 | 32.71 | 1.60 m |
| 26 | 22.1 | 22.30 | 1.04 d (6.5) | 17.8 | 18.00 | 0.83 d (6.5) |
| 27 | 22.0 | 22.47 | 1.05 d (6.5) | 20.7 | 20.86 | 0.90 d (6.5) |
| 28 | 106.6 | 106.88 | 4.67 br s/4.74 br s | 15.7 | 16.00 | 0.83 d (6.5) |

^aδ_C of ergosta-24(28)-ene-3β,5α,6β-triol in pyridine-*d*₅ [8]; ^bδ_C of ergosta-3β,5α,6β-triol in pyridine-*d*₅ [9].

Extract SCO.H (100.0 g) was fractionated on silica gel MPLC using the mobile phase of *n*-hexane-acetone (gradient 50:1→1:1, v/v) to yield eight fractions, SCO.H1–SCO.H8. Fraction SCO.H5 (10 g) was further subjected to RP-18 MPLC using the mobile phase of MeOH–H₂O (gradient 2:1→10:1, v/v) to give seventeen subfractions, SCO.H5A1–SCO.H5A17. Subfraction SCO.H5A14 (800 mg) was fractionated into ten smaller fractions, SCO.H5A14A–SCO.H5A14K, by silica gel CC using CH₂Cl₂-acetone (10:1, v/v) as eluent. Fraction SCO.H5A14C (80 mg) was purified by YMC CC eluting with acetone–H₂O (5:1, v/v), followed by silica gel CC with *n*-hexane-ethyl acetate (4.5:1, v/v) to afford compound **1** (3.0 mg). Fraction SCO.H5A14F (100 mg) was further fractionated by silica gel CC with *n*-hexane-acetone (2:1, v/v), followed by YMC CC eluting with acetone–H₂O (4:1, v/v), to furnish compound **4** (5.0 mg). Compounds **2** (1.0 mg) and **3** (2.0 mg) were purified

from fraction SCO.H5A14H (60 mg) after subjecting it on YMC CC eluting with acetone–H₂O (5:1, v/v). Fraction SCO.H5A14K (50 mg) was purified by silica gel CC eluting with *n*-hexane-acetone (5:1, v/v) to give compound **7** (9.5 mg). Subfraction SCO.H5A12 (700 mg) was fractionated into five smaller fractions, SCO.H5A12A–SCO.H5A12E, by silica gel CC with CH₂Cl₂-acetone (10:1, v/v). Fraction SCO.H5A12K (34 mg) was purified by silica gel CC eluting with *n*-hexane-acetone (3:1, v/v), followed by YMC CC with methanol–H₂O (10:1, v/v), to give compounds **5** (2.4 mg) and **6** (1.0 mg).

7α-Methoxyergosta-5,24(28)-diene-3β-ol (1): White powder; ¹H-NMR (500 MHz, CDCl₃) and ¹³C-NMR (125 MHz, CDCl₃) see table 1; ESI-MS *m/z* 429 [M+H]⁺ (C₂₉H₄₈O₂, M = 428).

Ergosta-5-ene-3β,7α-diol (2): White powder; ¹H-NMR (500 MHz, CDCl₃) and ¹³C-NMR (125

MHz, CDCl₃) see table 1; ESI-MS m/z 399 [M-H₂O+H]⁺ (C₂₈H₄₈O₂, M = 416).

Ergosta-24(28)-ene-3 β ,5 α ,6 β -triol (5): White powder; ¹H-NMR (500 MHz, CDCl₃) and ¹³C-NMR (125 MHz, CDCl₃) see table 2; ESI-MS m/z 433 [M+H]⁺ (C₂₈H₄₈O₃, M = 432).

Ergosta-3 β ,5 α ,6 β -triol (6): White powder; ¹H-NMR (500 MHz, CDCl₃) and ¹³C-NMR (125 MHz, CDCl₃) see table 2; ESI-MS m/z 435 [M+H]⁺ (C₂₈H₅₀O₃, M = 434).

3. RESULTS AND DISCUSSION

Compound **1** was isolated as a white powder. Its NMR features are indicative for an ergosterol derivative, one main constituent of soft corals [5]. The ¹H- and ¹³C-NMR spectra revealed typical signals of two oxymethines [δ_C 71.46 (C-3) and 73.92 (C-7)]/ δ_H 3.62 (1H, m, H-3) and 3.29 (1H, br s, H-7)], one trisubstituted double bond [δ_C 146.15 (C, C-4) and 120.78 (CH, C-5)]/ δ_H 5.73 (1H, dd, J = 1.0, 4.0 Hz, H-6)], one terminal disubstituted double bond [δ_C 156.97 (C, C-24) and 105.92 (CH₂, C-28)]/ δ_H 4.44 and 4.71, each 1H, s, H-28], two *tert*-methyls [δ_C 11.49 (C-18) and 18.27 (C-19)]/ δ_H 0.67 (H-18) and 0.99 (H-19), each 3H, s], three *sec*-methyls [δ_C 18.67 (C-21), 21.89 (C-26), and 22.03 (C-27)]/ δ_H 0.95 (H-21), 1.02 (H-26), and 1.03 (H-27), each 3H, d, J = 6.5 Hz], and one methoxy group [δ_C 56.78/ δ_H 3.35 (3H, s)].

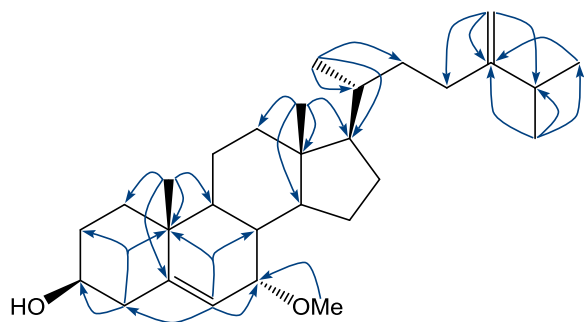


Figure 2: Key HMBC (H \rightarrow C) correlations of **1**

The HMBC cross-peaks of H-19 with C-5 and H-6 with C-8 and C-10 confirmed the position of the trisubstituted double bond at C-5/C-6. The disubstituted double bond was identified at C-24/C-28 by HMBC cross-peaks of H-28 with C-23 and C-25 and H-26/H-27 with C-24 and C-25. The HMBC correlation of the methoxy proton with C-7 confirming attachment of the methoxy group at this carbon. Comparison of the ¹³C-NMR data of **1** with those reported (table 1) and detailed analysis of the other HMBC cross-peaks (Fig. 2) led to the

identification of **1** as 7 α -methoxyergosta-5,24(28)-diene-3 β -ol [7, 10].

The NMR data of **2** were similar to those of **1**, except for absence of the methoxy group and presence of a methine and a *sec*-methyl in **2** instead of a terminal double bond in **1**. Comparison of the ¹³C-NMR data of **2** with those reported (table 1) and detailed analysis of the HSQC and HMBC correlations led to the identification of **2** as ergost-5-en-3 β ,7 α -diol [8].

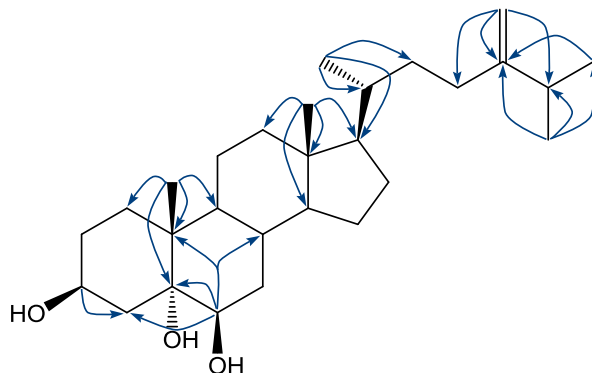


Figure 3: Key HMBC (H \rightarrow C) correlations of **5**

The NMR features of **5** are also indicative for an ergosterol derivative. The ¹H- and ¹³C-NMR spectra revealed typical signals of two oxymethines [δ_C 68.33 (C-3) and 76.53 (C-6)]/ δ_H 4.03 (1H, m, H-3) and 3.47 (1H, br s, H-6)], one oxygenated quaternary carbon [δ_C 76.81 (C-5)], one terminal disubstituted double bond [δ_C 157.78 (C, C-24) and 106.88 (CH₂, C-28)]/ δ_H 4.67 and 4.74, each 1H, s, H-28], two *tert*-methyls [δ_C 12.62 (C-18) and 17.31 (C-19)]/ δ_H 0.74 (H-18) and 1.18 (H-19), each 3H, s] and three *sec*-methyls [δ_C 19.21 (C-21), 22.30 (C-26), and 22.47 (C-27)]/ δ_H 0.99 (H-21), 1.04 (H-26), and 1.05 (H-27), each 3H, d, J = 6.5 Hz]. Comparison of the ¹³C-NMR data of **5** with those reported (Table 2) and detailed analysis of the HSQC and HMBC correlations (Fig. 3) led to identification of **5** as ergosta-24(28)-ene-3 β ,5 α ,6 β -triol [8].

Compound **6** was also isolated as a white powder. Its NMR data were similar to those of **5** (table 2), except for the presence of a methine and a *sec*-methyl in **6** instead of a terminal double bond in **5**. Comparison of the ¹³C-NMR data of **6** with those reported (table 2) and detailed analysis of the HSQC and HMBC correlations led to the identification of **6** as ergosta-3 β ,5 α ,6 β -triol [9].

The other compounds were elucidated as 3 β ,7 α -dihydroxyergosta-5,24(28)-diene (**3**) [11, 12], 3 β -hydroxyergosta-5,24(28)-diene-7-one (**4**) [13, 14], and ergosta-24(28)-ene-3 β ,5 α ,6 β -triol-6-acetate (**7**) [12, 15] by comparison of their ¹H- and ¹³C-NMR

values with those reported and detailed analysis of HSQC and HMBC correlations.

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Corresponding author: **Nguyen Hoai Nam**

Institute of Marine Biochemistry
Vietnam Academy of Science and Technology
No. 18, Hoang Quoc Viet, Cau Giay, Hanoi, Vietnam
E-mail: namnguyenhoai@imbc.vast.vn.