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# CHEMICAL STUDY OF THE LEAVES OF LAGERSTROEMIA SPECIOSA IN VIETNAM

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#### Abstract

The main constituents of the EtOH extract of leaves of *Lagerstroemia speciosa* supplied by Traphaco company have been quantitatively determined by HPLC to be corosolic acid (2.242 %), asiatic acid (1.219 %) and  $\beta$ -sitosterol. Repeated column chromatography of the *n*-hexane, ethyl acetate and dichloromethane extract of these leaves led to the isolation of corosolic acid, asiatic acid, quercetin,  $\beta$ -sitosterol and  $\beta$ -sitosterol-3-O- $\beta$ -D-glucopyranosyl glucoside. The structures of these compounds have been elucidated by the IR, MS and NMR spectral analysis.

Keywords. Lagerstroemia speciosa, leaves, quercetin, corosolic acid, asiatic acid.

#### 1. INTRODUCTION

The species Lagerstroemia speciosa (L.) Pers. (Vietnamese name: Bằng lăng nước) belonging to the family Lythraceae is a big tree of 10-15 meters high. This plant is growing wildly in India, Thailand, Laos, Cambodia, Philippine, Vietnam and South China. In the folk medicine of many countries Lagerstroemia speciosa is used for treatment of type II diabetes and other diseases like diarrhea, heart, stomach [1]. Studies on the chemical constituents of these plants have shown the presence of tannins, flavonoids [2] fatty acids [3] and terpenoids [4]. In a cooperation with Traphaco Company to develop a functional food product from Vietnamese L. speciosa for supporting the treatment of type II diabetes we report here the isolation of the main constituents from the leaves of this plant: corosolic acid, asiatic acid,  $\beta$ -sitosterol, daucosterol and quercetin.

### 2. EXPERIMENTAL

**Plant material**: the leaves of *L. speciosa* have been supplied by Traphaco Company, a herbal exemplar. No.BL02 is deposited in the Research Department of this company.

**Chemicals and methods**: Silica gel (Merck) with different sizes from 0.040-0.063 mm, reverse phase RP18 and Sephadex LH20 were used for column chromatography. TLC used the precoated

aluminum sheets with silica gel 60 GF 254, 0.2 mm (Merck). IR: Nicolet Impact 410 (KBr), MS: HP 5989 B MS Engine Agilent, NMR: Bruker Avance 500 MHz. For quantitative determination of the main constituents the HPLC Alliance series was used.

**Extraction**: Dried leaves of *L. speciosa* (1600 g) were extracted with ethanol-water mixture 70:30 (v/v) at 60 °C three times. Evaporation of solvents under reduced pressure afforded 240 g extract. Quantitative HPLC analysis of this extract gave corosolic acid (2.242 %) and asiatic acid (1.219 %) as the main constituents.

200 g EtOH extract were dissolved in H<sub>2</sub>O and extracted with *n*-hexane and ethyl acetate, successively to yield 12 g n-hexane and 25 g ethyl acetate extract. Chromatography of 12 g n-hexane extract on silica gel eluted by n-hexane/EtOAc mixture gradient from 5-20 % EtOAc gave 4 Fraction HE2 fractions (HE1-HE4). was rechromatographed on silica gel with the solvent mixture *n*-hexane/EtOAc 9:1 to furnish 700 mg  $\beta$ sitosterol (1). 25 g EtOAc extract were fractionated on a silica gel column with *n*-hexane/EtOAc mixture gradient from 10-70 % EtOAc to give 5 fractions (EA1-EA5). Repeated chromatography of fraction EA2 on silica gel with a solvent mixture hexane/EtOAc 85:15 (v/v) afforded 250 mg βsitosterol (1) and 30 mg quercetin (3). Fraction EA3 was rechromatographed on a RP 18 column with a mixture MeOH/H<sub>2</sub>O 8:1 as eluent to give 650 mg asiatic acid (5). Fraction EA4 was rechromatographed on silica gel (CH<sub>2</sub>Cl<sub>2</sub>/MeOH 9:1) to furnish 2500 mg corosolic acid (4) and 25 mg  $\beta$ -sitosterol-3-O- $\beta$ -D-glucopyranoside.

Compound **1** ( $\beta$ -sitosterol):

White crystals, mp 169-171 °C. <sup>1</sup>H-NMR ( $\delta$ , CDCl<sub>3</sub>): 1.02 ( 3H, s, H-18), 0.71 (3H, s, H-19), 0.86 (3H, d, J = 7.0 Hz, H-26), 0.85 (3H, d, J = 7.0 Hz, H-27), 0.93 (3H, d, J = 5.5 Hz, H-21), 0.85 (3H,t, J = 7.0 Hz, H-29), 5.38 (1H, dd, J = 3.0, 2.5 Hz, H-6),3.53 (1H, m, H-3).

<sup>13</sup>C-NMR spectrum is in full agreement with that of  $\beta$ -sitosterol in [5, 6].

Compound **2** ( $\beta$ -sitosterol-3-*O*- $\beta$ -D-glucopyranoside, Daucosterol), white powder, <sup>1</sup>H-NMR (DMSO-d<sub>6</sub>)  $\delta$  ppm: 0.65 (3H, s), 0.80 (3H, d, *J* = 6.9 Hz), 0.81 ( 3H, d, *J* = 6.9 Hz), 0.90 (3H, d, *J* = 6.5 Hz), 0.96 (3H, s), 1.00 (3H, d, *J* = 6.7 Hz), 1.23 (3H, s), 3.05-3.08 (1H, m), 3.10-3.14 (3H, m), 3.38-3.48 (2H, m), 3.64 (1H, dd, *J* = 5.5, 10.1 Hz), 4.22 (1H, d, *J* = 7.8 Hz), 4.39 (1H, t, *J* = 5.7 Hz), 4.83 (3H, m), 5.32 (1H, d, *J* = 7.0). <sup>13</sup>C-NMR spectrum is in full agreement with that for daucosterol in [7].

Compound **3** (quercetin): yellow needles. IR (KBr, cm<sup>-1</sup>): 3450 (OH), 1662 (C=O), 1614 (C=C), <sup>1</sup>H-NMR ( $\delta$ , CDCl<sub>3</sub>): 6.20 (1H, d, *J* = 1.9 Hz, H-6), 6.40 (1H, d, *J* = 1.9 Hz, H-8), 6.90 (1H, d, *J* = 8.5 HZ, H-5'), 7.65 (1H, dd, *J* = 2.0, 8.5 Hz, H-6'), 7.75 (1H, *J* = 2.0 Hz, H-1'). <sup>13</sup>C-NMR spectrum is in full agreement with that for quercetin in [8].

Compound 4 (corosolic acid): white powder, positive ions ESI- MS:  $m/z = 473 \text{ [M+H]}^+$ , <sup>1</sup>H and <sup>13</sup>C-NMR spectral data, see table 1.

Compound 5 (asiatic acid): white powder, IR

(KBr, cm<sup>-1</sup>): 3144 (OH), 2930, 2866 (CH<sub>2</sub>, CH<sub>3</sub>), 1690 (COOH), 1459, 1053, 694. ESI-MS (negative ions):  $m/z=487[M-H]^{-1}$ 

<sup>1</sup>H and <sup>13</sup>C- NMR spectral data, see table 1.

#### 3. RESULTS AND DISCUSSION

It is well known that the leaves extract of Lagerstroemia speciosa has a blood sugar content lowering effect and can be used for treatment of diabetes type II. The main active constituent in this extract has been evidenced as corosolic acid (4). Very important is to find out the Lagerstroemia speciosa or its varieties in Vietnam, which contain high enough amount of corosolic acid for developing a product used in diabetes type II treatment. In cooperation with Traphaco company we have had screenings of about 10 Lagerstroemia samples and found out that the sample BL02 was the best one. However, there were differencies between the amount of corosolic acid and asiatic acid determined by quantitative HPLC (2.242 % and 1.219 % of dry extract, respectively) and the isolated amount by repeated column chromatography (1.25 % and 0.3259 %, respect- tively). That means the contents of corosolic acid and asiatic acid in dry leaves of sample BL02 are 0.35 % and 0.19 %, respectively. So that the sample BL02 can be used as starting material for preparation of a functional food product used in treatment of diabetis type II.

The structures of isolated corosolic acid and asiatic acid have been determined by comparison of <sup>1</sup>H- and <sup>13</sup>C-NMR spectral data with the published data (table 1).



Table 1: NMR s	pectral data	of <b>4</b> and <b>5</b>
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Pos.	. <b>4</b> (DMSO- <i>d</i> <sub>6</sub> )		Corosolic acid (pyridine $-d_{\epsilon}$ ) [9]		<b>5</b> (CDCl <sub>3</sub> +CD <sub>3</sub> OD)		Asiatic acid [ 10 ]	
	δ <sub>C</sub>	$\delta_{\rm H} (J = {\rm Hz})$	δ	$\delta_{\rm H} (J = {\rm Hz})$	δ <sub>C</sub>	$\delta_{\rm H} (J = {\rm Hz})$	$\delta_{\rm C}$	$\delta_{\rm H} (J = {\rm Hz})$
1	47.0		48.1		48.0		48.0	
2	67.1	4.24 (dd)	68.1	4.11 (1H, ddd, J = 4.0, 9.5,	69.3	3.7 (dt,1H)	69.7	3.71 (dt,1H)
3	82.2	3.38 (d, <i>J</i> = 9.5 Hz)	84.3	$\frac{11.0 \text{ Hz}}{3.43 (1\text{H, d, }J)} = 9.5 \text{ Hz}$	78.2	3.38 (d, <i>J</i> = 9.5 Hz, 1H)	78.2	3.38 (d, <i>J</i> = 9.5 Hz, 1H)
4	39.1		40.3		44.1		44.1	
5	54.7		56.1		48.2		48.2	
6	18.0		19.3		19.1		19.1	
7	32.6		34.0		33.6		33.6	
8	39.2		40.5		40.7		40.7	
9	46.9		48.5		48.5		48.5	
10	37.5		38.9		39.0		38.9	
11	22.9		24.2		24.5		24.5	
12	124.5	5.26 (1H, t, J = 3.3 Hz)	126.0	5.48 (1H, br s)	126.4	5.28 (br,1H)	126.6	5.26 (br,1H)
13	138.2		139.8		140.0		139.8	
14	41.7		43.0		43.4		43.4	
15	27.5		29.1		29.2		29.1	
16	23.8		25.4		25.4		25.3	
17	47.0		48.6		48.8		48.7	
18	52.3		54.0		54.4		54.3	
19	38.4	2.12 (1H, d, <i>J</i> = 11.5 Hz)	39.9	2.65 (1H, d, J) = 11.0 Hz)	40.5		40.4	
20	38.5	2.74 (1H, dd, J = 4 and 9.5 Hz)	40.0		40.6		40.4	
21	30.1		31.5		31.7		31.7	
22	36.3		37.9		38.2		38.1	
23	28.8		29.8		66.5	3.29 (d, J = 11.0 Hz, 1H) 3.51(d, J = 11.0 Hz, 1H)	66.3	3.29 (d, J = 11.0 Hz, 1H) 3.53(d, J = 11.0 Hz, 1H)
24	17.1		18.2		13.9		13.9	
25	16.4		17.4		17.5		17.6	
26	16.9		17.9		17.5		17.7	
27	23.2		24.4		24.1		24.1	
28	178.2		180.4		181.6		181.6	
29	17.0		18.0		18.0		17.8	
30	21.0		21.9		21.6		21.5	

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