# INVESTIGATING THE PHYTOCHEMICAL PRODUCTION AND ANTIOXIDANT ACTIVITY IN FOUR PLANT SPECIES BELONGING TO THE ASTERACEAE FAMILY

Thi Van Anh Le<sup>®1</sup>, Ngoc Trung Anh Tran<sup>®1</sup>, Thi Hien Diu Dinh<sup>®1</sup>, Phuong Anh Duong <sup>®1</sup>, Tran Bao Chau Ha<sup>1</sup> and Nga Thi Phuong Mai<sup>®1,⊠</sup>

<sup>1</sup>University of Science and Technology of Hanoi- Vietnam Academy of Science and Technology, 18 Hoang Quoc Viet, Cau Giay, Ha Noi, 10000, Vietnam.

<sup>\infty</sup>To whom correspondence should be addressed. E-mail: mai-thi-phuong.nga@usth.edu.vn

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### ABSTRACT

Vietnam has a rich and profound traditional medicine system that is widely used today. Medicinal plants are used to treat colds, coughs, bone and joint diseases, digestion, respiratory diseases, etc. In this study, leaves of Artemisia vulgaris, Taraxacum officinale, Blumea balsamifera, and flowers of Xerochrysum bracteatum from the Asteraceae family were selected to determine the antioxidant capacity and relationship with the active ingredients in the plants. The methanolic extracts were screened for chemical compositions via the total phenolic content (TPC) assay, the total flavonoid content (TFC) assay, and DPPH radical scavenging activity. The highest radical scavenging activity was observed in the flowers of X. bracteatum (IC<sub>50</sub> = 0.061 mg/mL), followed by the leaves of Taraxacum  $(IC_{50} = 0.313 \text{ mg/mL})$ , A. vulgaris  $(IC_{50} = 1.367 \text{ mg/mL})$ , and B. balsamifera  $(IC_{50} = 1.4502 \text{ mg/mL})$ mg/mL). The TPC of the studied plants ranged from  $19.98 \pm 1.355$  to  $195.78 \pm 42.518$ mgGAE/g extract, while the TFC ranged from  $60.31 \pm 1.725$  to  $339.14 \pm 26.299$  mgQE/g extract. The highest TPC and TFC were recorded in the methanol extract of X. bracteatum. The strongly negative correlation between the TPC and TFC and the  $IC_{50}$  values ( $R^2 = -0.78$ ,  $R^2 = -0.76$ ) suggests that TFC and TPC could strongly contribute to the antioxidant activity of these plants. These results not only highlight the relevance of these plants in traditional medicine but also scientifically validate their use, particularly in the context of their antioxidant properties. The study underscores the close relationship between the traditional use of these plants and their scientifically observed effects, reinforcing the value of folk remedies.

**Keywords:** Antioxidant, *Asteraceae*, flavonoid, free radical scavenging activity, phenolic, secondary metabolites

### **INTRODUCTION**

The *Asteraceae* family, commonly called the sunflower family, is a remarkably diverse

and widespread group of flowering plants comprising over 1.600 genera and approximately 25.000 species distributed across the globe, except Antarctica. It includes many well-known plants, such as chicory, sunflower, daisies, and dandelion, which are renowned for their potential medicinal functions, including antioxidant, antimicrobial, anticancer, and antidiabetic (Rolnik & Beata, 2021).

Furthermore, the Asteraceae family is characterized by a rich diversity of phytochemical compounds, including polyphenols, phenolic acids, flavonoids, sesquiterpene lactones, essential oils. saponins, and lignans (Rolnik et al., 2021). This complex chemical composition contributes to the biological activities exhibited by many species within the family, valuable making them resources in traditional medicine practices across various cultures.

Artemisia vulgaris, or mugwort, is a perennial herb with a fragrant and bitter taste. It has unique pharmacological properties that are used in traditional medicinal systems of China, Europe, India, and Vietnam. According to traditional medicine, mugwort is used as a medicine and food with the therapeutic effect of warming the meridians, relieving pain, dispelling dampness, and dispersing cold, so it is used to treat diseases such as convulsions. fever. malaria. regulating menstruation, and the digestive tract (Thangjam et al., 2020) thanks to its important phytochemicals such as oil, phenol, flavonoids, terpenoids, and saponin, as well as tannin (Thangjam et al., 2020; Ashok & Upadhyaya, 2013).

*Blumea balsamifera* (sembung plant) is a traditional medicine widely used across Asian cultures for its diverse health benefits. In traditional systems, every part of the *B. balsamifera* plant has been employed to treat numerous ailments, such as remedies for coughs, colds, fever, and joint pain, even

relieving skin infections and inflammation (Pang *et al.*, 2014). Its various pharmacological actions are supported by studies demonstrating high antioxidant activity with powerful bioactive compounds such as flavonoids and phenolic acids (Dai *et al.*, 2023). Additional investigations have shown it possesses anti-inflammatory, antibacterial, and hepatoprotective effects (Widhiantara & Jawi, 2021).

*Xerochrysum bracteatum*, commonly known as golden everlasting and *Helichrysum bracteatum* for many years before, is an aromatic herbaceous plant native to eastern Australia. *Helichrysum* species have been recognized for their therapeutic aspects as diuretic, anti-inflammatory, hepatoprotective, anti-psoriasis, antimicrobial, and antioxidant activities (Najar *et al.*, 2019; Akinyede *et al.*, 2021).

Taraxacum officinale, known as dandelion, is a recognizable worldwide herb in temperate grasslands and pastures. Dandelion originated from Europe and has a long history of use in folk medicine across different traditional systems, treating liver and digestive problems (Fan et al., 2023; Schütz et al., 2006). Many authors reported the antioxidant, anti-inflammatory, anticancer, and other effects of dandelion (Jalili et al., 2020) thanks to its phenolics, flavonoids, terpenes, and coumarins (Yan et al., 2024).

This study aimed to determine the antioxidant potential and free radical scavenging activity of wild medicinal plant species belonging to the Asteraceae family. A range of assays would be used to evaluate each plant species' antioxidant capacity and ability scavenge free radicals. to Additionally, the total phenolic and flavonoid content would be quantified and correlated with the observed antioxidant activity to analyze their relationship. By providing scientific evidence of their antioxidant profiles and phytochemical contents, the study intended to validate the traditional uses of the plant species.

### MATERIALS AND METHODS

### **Plants materials**

Mature leaves of *Artemisia vulgaris*, *Taraxacum*, *Blumea balsamifera*, and flowers of *Xerochrysum bracteatum* of the 2-year-old plants from the *Asteraceae* family were collected in Ninh Binh province and dried for further studies.

### Methods

### Preparation of methanolic extracts

The samples were cleaned and dried at 60°C until they reached a constant weight. Once dried, they were ground into a fine powder and placed in separate vials. Our study used methanol as the solvent to extract the bioactive compound from the samples. One gram of every sample added with 99.5% methanol (Xilong) at the 1:11 (v/v) ratio was sonicated at 38°C for 30 minutes, followed by being centrifuged at 13000 rpm 4°C for 10 minutes. The supernatant, containing the crude extract, was then evaporated in an oven set at 60°C for 2 days to remove the methanol and concentrate the bioactive compounds. The extract mass was determined and the concentration was standardized by adding a certain amount of methanol for the final 40 mg/mL concentration.

## 2,2-Diphenyl-1-picrylhydrazyl radical scavenging activity (DPPH)

In vitro antioxidant activities of the extracts were determined using the DPPH free radical scavenging assay, described by Mai *et al.* (2024). The ascorbic acid was used as a positive control. In each reaction,  $40 \,\mu$ L of the plant extract was mixed with 760  $\mu$ L of DPPH 0.25 mM (Sigma-Aldrich) and incubated at room temperature for 20 minutes in the dark. The absorbance of the mixture was measured at 517 nm by UV-Vis spectrophotometer (SpectraMax® iD5) against the blank solution.

### Total phenolic content (TPC)

The TPC in the methanolic extracts was estimated using the Folin-Ciocalteu colorimetric method (Singleton et al., 1999) with some modifications. Gallic acid (GAE) (Acros Organics) was prepared for the standard calibration curve. In each sample, 40 µL of the plant extract was mixed with 480 µL of Folin-Ciocalteu reagent 10% (Merck) and then incubated at 40°C for a minute. Thus, 480 µL of sodium carbonate 6% was added and continuously incubated at 40°C for 15 minutes. The absorbance of the blue-colored mixtures was measured at 765 nm by using the UV-Vis spectrophotometer (SpectraMax® iD5) against the blank solution. The TPC value was expressed as mg GAE/g extract.

### Total flavonoid content (TFC)

The TFC in the methanolic plant extracts was estimated using the aluminum chloride colorimetric method (Phuyal *et al.*, 2020) with slight modification. Quercetin (QE) (Aldrich Sigma) was prepared for the standard calibration curve. In each reaction, 240  $\mu$ L of plant extract was mixed with 40  $\mu$ L of 5% sodium nitrite solution and incubated at 27°C for 6 minutes. After incubation, 40  $\mu$ L of 10% aluminum chloride solution was added to the mixture and continuously incubated for 6 minutes. Thus, 400  $\mu$ L of NaOH 1M and 280  $\mu$ L of ethanol 30% were added simultaneously and left for 30 minutes. Finally, the OD of mixtures was measured at 510 nm by UV-Vis spectrophotometer (SpectraMax® iD5) against the blank solution. The TPC value was expressed as mg QE/g extract.

# All the experiments were repeated three times. The data was presented by means of three replicates $\pm$ standard deviation. The difference between parameters was analyzed using one-way ANOVA and Turkey's posthoc test to assess the statistical significance of the means, with p < 0.05 considered significant.

### RESULTS

### Statistical Analysis

Total phenolic contents in four studied plants

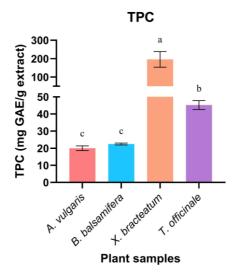


Figure 1. Total phenolic contents in four studied plants from the Asteraceae family

The analysis of TPC presented in Figure 1 highlighted significant variations among four plant species in the *Asteraceae* family - *A. vulgaris, B. balsamifera, X. bracteatum,* and *T. officinale.* An exceptionally high TPC of 195.78  $\pm$  42.52 mg GAE/g was demonstrated by *X. bracteatum.* A moderate TPC of 45.22  $\pm$  2.58 mg GAE/g (p < 0.05) was shown by *T. officinale.* In contrast, a lower TPC of 22.45  $\pm$  0.56 mg GAE/g was recorded for *B. balsamifera*, which was not significantly different (p > 0.05) from *A.* 

*vulgaris*, the species with the lowest TPC at 19.98  $\pm$  1.35 mg GAE/g. These findings suggested that *X. bracteatum* was likely the most potent source of phenolic compounds among the species studied, making it a valuable candidate for applications requiring high antioxidant capacity.



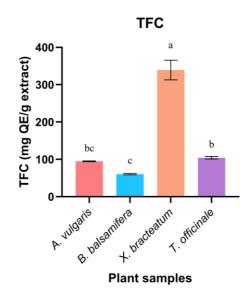


Figure 2. Total flavonoid content in four studied plants from the Asteraceae family

Based on the TFC data shown in Figure 2 for the four plant species, significant differences were revealed among them. The highest TFC value of 339.145  $\pm$  26.299 mg QE/g was exhibited by *X. bracteatum*, which was found to be substantially higher than that of the other species (p<0.05), indicating a strong presence of flavonoids. Conversely, the lowest TFC was observed in *B. balsamifera*, with a mean of 60.309  $\pm$  1.725 mg QE/g (p<0.05). Intermediate TFC values were recorded for *A. vulgaris* and *T*. officinale, with means of  $94.813 \pm 0.993$  mg QE/g and  $104.321 \pm 3.443$  mg QE/g extract, respectively. The TFC of *A. vulgaris* was found to be significantly different from that of *B. balsamifera* but not from *Taraxacum*. While the TFC of *T. officinale* was determined to be statistically higher than that of *B. balsamifera* but lower than *X. bracteatum*.

### 2,2-Diphenyl-1-picrylhydrazyl radical scavenging activity (DPPH)

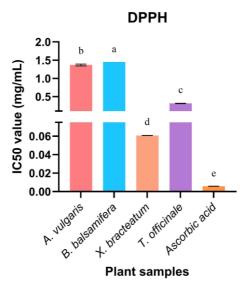
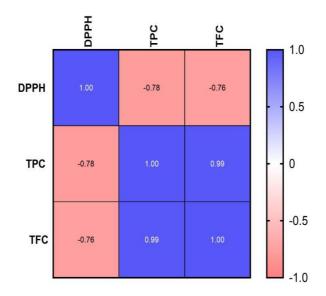


Figure 3. IC<sub>50</sub> values of four samples for free radical scavenging activity by DPPH radical.

DPPH Figure 3 shows the radical scavenging activity, measured as IC<sub>50</sub> values, for four plant species within the Asteraceae family, along with ascorbic acid as a reference standard. Ascorbic acid exhibited the lowest IC<sub>50</sub> value of approximately 0.006 mg/mL. Among the plant species tested, although it is significantly ten times less potent than ascorbic acid, X. bracteatum demonstrated the strongest antioxidant activity with an IC<sub>50</sub> value of approximately

0.061 mg/mL. *T. officinale* also exhibited moderate antioxidant activity, with an IC<sub>50</sub> of approximately 0.313 mg/mL. In contrast, *A. vulgaris* and *B. balsamifera* showed weaker antioxidant activities, with IC<sub>50</sub> values of approximately 1.367 mg/mL and 1.45 mg/mL, respectively.

The correlation between antioxidant parameters in four studied plants from the Asteraceae family



**Figure 4.** Pearson correlation matrix between DPPH, TPC, and TFC in four extracts from studied plants in the *Asteraceae* family.

The Pearson correlation matrix presented in Figure 4 highlighted the relationships between DPPH radical scavenging activity, TPC, and TFC for the four plant species belonging to the Asteraceae family that were studied. A strong negative correlation was observed between DPPH and TPC, with a correlation coefficient of -0.78. This suggested that higher total phenolic content was associated with stronger antioxidant activity, as indicated by lower DPPH IC<sub>50</sub> negative values. Similarly, a strong correlation of -0.76 was found between DPPH and TFC, indicating that higher flavonoid content was also linked to increased antioxidant activity. An almost perfect positive correlation of 0.99 was observed between TPC and TFC, suggesting that in these plants, phenolic and flavonoid contents were closely related, likely because flavonoids are a subclass of phenolic compounds. These findings implied that the plants with higher TPC and TFC, such as X.

*bracteatum*, were likely to exhibit stronger antioxidant properties.

### DISCUSSION

This study investigated the antioxidant properties and phytochemical composition of some medicinal plants of the Asteraceae family commonly used in traditional medicine in various cultures, especially in Specifically, we Vietnam. evaluated Artemisia vulgaris, Blumea balsamifera, Xerochrysum bracteatum, and T. officinale species to determine their total phenolic, flavonoid content. and free radical scavenging capacity.

Our study on *A. vulgaris* showed lower TPC, TFC, and weaker antioxidant activity compared to other *Asteraceae* species, with TPC of 19.98  $\pm$  1.35 mg GAE/g, TFC of 94.81  $\pm$  0.99 mg QE/g and IC<sub>50</sub> value of 1.367 mg/mL in the DPPH assay. Differently, the study by Sharma *et al.* 

(2023), which also used methanol as solvent to extract metabolites, found higher TPC and antioxidant activity but lower TFC in A. vulgaris grown in Nepal, with TPC of 26.04  $\pm$  1.66 mg GAE/g, TFC of 33.41  $\pm$  0.27 mg OE/g and  $IC_{50}$  of 149.62  $\pm$  2.40 µg/mL for leaf extract from Chitwan, Nepal. These differences highlight the significant impact of environmental factors, such as geographical location (Ninh Binh, Vietnam vs. Chitwan, Nepal), as well as differences in extraction methods and experimental conditions on the phytochemical composition and biological activities of A. vulgaris. Although our study observed lower TPC and antioxidant activity but higher TFC than Nepalese samples, A. vulgaris still exhibited valuable bioactive properties. This suggests that with further optimization of extraction techniques and exploration of different growth conditions, the antioxidant potential and therapeutic applications of A. vulgaris could be significantly enhanced.

Although in our report, B. balsamifera exhibited lower antioxidant activity than other species in the Asteraceae family, they are still regarded as having the potential for exploitation due to their numerous pharmacological benefits, including pain relief and fever reduction, etc. Zulkiflee. (2022) conducted a study on the TPC and TFC of B. balsamifera using a variety of extract solvents, including ethanol, methanol, and hexane, which the TPC and TFC contents of B. balsamifera extract with methanol were approximately four times lower than the results stated in our report, even with other extract solvents. On the other hand, in the report of Rawati et al. (2023), the hexane-extracted B. balsamifera sample demonstrated a lower  $IC_{50}$  value than the methanol-extracted sample in our report. This indicates that the hexane-extracted

sample had a superior DPPH radical scavenging ability. The discrepancy may be attributed to the different extraction methods, sample extraction solvents, or growth media. Besides, the scavenging capacity of a sample in the DPPH radical scavenging experiment is determined by its electron or hydrogen atom transfer potential, which is controlled the redox characteristics of by the phytochemicals present in the sample. While flavonoids and phenolic compounds are efficient DPPH frequently radical scavengers, their efficacy can differ depending on their particular chemical compositions and the existence of other substances that may influence their action.

Out of the species investigated from the Asteraceae family, X. bracteatum was identified as the most effective antioxidant plant. Compared to research by Kandylis, 2022, the overall finding indicated that X. bracteatum exhibited a superior capacity to scavenge free radicals compared to the other plant species. In addition, our findings indicated a notably greater phytochemical content in Vietnamese X. bracteatum compared to Kandylis' research. Particularly, this was proven through the phenolic and flavonoid content analysis, revealing that the Vietnamese X. bracteatum flowers have approximately triple the number of phenolic compounds. The differences can be explained differences by the in environmental conditions such as soil, and climate. Although X. bracteatum is wellknown for its antibacterial and antiinflammatory properties, this discovery has highlighted its important potential in combating free radicals.

Compared to other plant extracts in the *Asteraceae* family, the methanol extract of Vietnamese *T. officinale* exhibited moderate antioxidant properties, with an  $IC_{50}$  of

approximately 0.313 mg/mL, which was comparable to that reported for *T. officinale* grown in Turkey, which showed an IC<sub>50</sub> of 0.482 mg/mL (Ergün, 2021). Furthermore, the high DPPH scavenging ability exhibited by the Vietnamese extract corroborates the traditional medicinal use of this species to treat diseases associated with oxidative stress and free radical-induced tissue damage. Besides. the quantitative phytochemical analysis revealed the extract contained significant amounts of phenolic compounds (45.225 mg GAE/g) and flavonoids (104.321 mg QE/g) in our study, demonstrating the rich polyphenolic profile of the Vietnamese T. officinale leaf extract. Ergün (2021) evaluated 36.53 mg GAE/g and 43.31 mg QE/g in T. officinale leaf extract, markedly lower than the present findings. However, Liu et al. (2020) extracted the T. officinale by fermentation pre-treatment and measured elevated flavonoid levels ranging from 109.49 to 183.72 mg/g, suggesting fermentation optimization of bioactive compounds' yields. This suggested that fermentation processing can improve phenolic and flavonoid biosynthesis and extraction. Notably, the Vietnamese methanol extract of Т. officinale's leaf contained flavonoid levels on par with the samples reported by Liu *et al.* (2020) implying the specific regional factors modulate the production of these secondary metabolites.

### CONCLUSION

Our study reveals the high potential antioxidant activity of four different plant species belonging to the *Asteraceae* family. Among them, *Xerochrysum bracteatum* appears to be the best candidate for therapeutic application in terms of antioxidant activity, where we obtained the highest TPC, TFC, and strongest DPPH scavenging activity. Further studies need to be conducted to optimize the metabolite extraction process in order to obtain the higher antioxidant compounds or to study the effect of environmental conditions in the production of metabolites.

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### **CONFLICT OF INTEREST**

The authors declare that there is no conflict of interest.

### REFERENCES

Akinyede KA, Cupido CN, Hughes GD, Oguntibeju OO, Ekpo OE (2021) Medicinal properties and in vitro biological activities of selected *Helichrysum* species from South Africa: A Review. *Plants* 10(8): 1566.

Ashok PK, Upadhyaya K (2013) Evaluation of analgesic and anti-inflammatory activities of aerial parts of *Artemisia vulgaris* L. in experimental animal models. *J. of Bio. Active Prod. from Nature* 3(1): 101–105. https://doi.org/10.1080/22311866.2013.782761.

Chaachouay N, Zidane L (2024) Plant-Derived Natural Products: a source for drug discovery and development. *Drugs and Drug Candidates* 3(1): 184–207.

https://doi.org/10.3390/ddc3010011.

Dai L, Cai S, Chu D, Pang R, Deng J, Zheng X, Dai W (2023) Identification of chemical constituents in *Blumea balsamifera* using UPLC–Q–Orbitrap HRMS and evaluation of their antioxidant activities. *Molecules* 28(11): 4504.

https://doi.org/10.3390/molecules28114504.

Ergün F (2021) Determination of phytochemicals and antioxidant capacity of

edible dandelion plant (*Taraxacum officinale*) collected from Kirşehir region. *Üniversitesi Akademik Veri Yönetim Sistemi*. https://www.cabidigitallibrary.org/doi/pdf/10.5 555/20220174035.

Fan M, Zhang X, Song H, Zhang Y (2023) Dandelion (Taraxacum Genus): A review of chemical constituents and pharmacological effects. *Molecules* 28(13): 5022. https://doi.org/10.3390/molecules28135022.

Garcia-Oliveira P, Barral M, Carpena M, Gullón P, Fraga-Corral M, Otero P, Prieto MA, Simal-Gandara J (2021) Traditional plants from Asteraceae family as potential candidates for functional food industry. *Food & Function 12*(7): 2850–2873. https://doi.org/10.1039/d0fo03433a.

Jalili C, Taghadosi M, Pazhouhi M, Bahrehmand F, Miraghaee SS, Pourmand D, Rashidi I (2020) An overview of therapeutic potentials of *Taraxacum officinale* (dandelion): a traditionally valuable herb with a rich historical background. *World Cancer Research Journal* 7: e1679. https://www.wcrj.net/article/1679.

Liu N, Song M, Wang N, Wang Y, Wang R, An X, Qi J (2020) The effects of solid-state fermentation on the content, composition, and in vitro antioxidant activity of flavonoids from dandelion. *PLoS ONE* 15(9): e0239076. https://doi.org/10.1371/journal.pone.0239076.

Najar B, Cervelli C, Ferri B, Cioni P, Pistelli L (2019) Essential oils and volatile emission of eight South African species of *Helichrysum* grown in uniform environmental conditions. *South African Journal of Botany* 124: 178–187. https://doi.org/10.1016/j.sajb.2019.05.015.

Nga MTP, Linh TK, Tra NPC, Linh HTP (2024) Strategies to enhance the production of biomass and natural antioxidants of *Cichorium intybus* L. hairy roots by using nanoparticle elicitors. *Vietnam Journal of Biotechnology* 22(2): 305– 317. https://doi.org/10.15625/vjbt-19545.

Pandey KB, Rizvi SI (2009) Plant polyphenols as dietary antioxidants in human health and

disease. Oxidative Medicine and Cellular Longevity 2(5): 270–278. https://doi.org/10.4161/oxim.2.5.9498.

Pang Y, Wang D, Fan Z, Chen X, Yu F, Hu X, Wang K, Yuan L (2014) *Blumea balsamifera*-A phytochemical and pharmacological review. *Molecules* 19(7): 9453–9477. https://doi.org/10.3390/molecules19079453.

Phuyal N, Jha PK, Raturi PP, Rajbhandary S (2020) Total phenolic, flavonoid contents, and antioxidant activities of fruit, seed, and bark extracts of Zanthoxylum Armatum DC. *The Scientific World Journal* 2020: 1–7. https://doi.org/10.1155/2020/8780704.

Rawati S, Ginting B, Maulana I, Yahya M, Saidi N, Murniana M, Hasballah K (2023) Phytochemical, antioxidant, and cytotoxicity screenings of n-hexane extract from *Blumea* balsamifera L. leaves. *Research Journal of Pharmacy and Technology* 16(4): 1664–1668. https://doi.org/10.52711/0974-360x.2023.00272.

Rolnik A, Olas B (2021) The plants of the *Asteraceae* family as agents in the protection of human health. *International Journal of Molecular Sciences* 22(6): 3009. https://doi.org/10.3390/ijms22063009.

Rolnik A, Soluch A, Kowalska I, Olas B (2021) Antioxidant and hemostatic properties of preparations from *Asteraceae* family and their chemical composition - Comparative studies. *Biomedicine & Pharmacotherapy* 142: 111982. https://doi.org/10.1016/j.biopha.2021.111982.

Schütz K, Carle R, Schieber A (2006) *Taraxacum*-A review on its phytochemical and pharmacological profile. *Journal of Ethnopharmacology* 107(3): 313–323. https://doi.org/10.1016/j.jep.2006.07.021.

Seca AML, Pinto DCGA (2019) Biological potential and medical use of secondary metabolites. *Medicines* 6(2): 66. https://doi.org/10.3390/medicines6020066.

Sharma KR, Adhikari S (2023) Phytochemical analysis and biological activities of *Artemisia* 

*vulgaris* grown in different altitudes of Nepal. *International Journal of Food Properties* 26(1): 414–427.

https://doi.org/10.1080/10942912.2023.216695 4.

Singleton VL, Orthofer R, Lamuela-Raventós RM (1999) Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. *Methods in enzymology* 152–178. https://doi.org/10.1016/s0076-6879(99)99017-1.

Thangjam NM, Taijong J, Kumar A (2020) Phytochemical and pharmacological activities of methanol extract of *Artemisia vulgaris* L. leaves. *Clinical Phytoscience* 6(1): 72. https://doi.org/10.1186/s40816-020-00214-8.

Widhiantara IG, Jawi IM (2021) Phytochemical composition and health properties of Sembung plant (*Blumea balsamifera*): A review. *Veterinary World* 14(5): 1185–1196. https://doi.org/10.14202/vetworld.2021.1185-1196.

Yan Q, Xing Q, Liu Z, Zou Y, Liu X, Xia H (2024) The phytochemical and pharmacological profile of dandelion. *Biomedicine & Pharmacotherapy* 179: 117334. https://doi.org/10.1016/j.biopha.2024.117334.