VARIANTS *APOA5* P.G185C AND P.S19W ASSOCIATED WITH EARLY-ONSET SEVERE HYPERTRIGLYCERIDEMIA-INDUCED PANCREATITIS AND DIABETES COMPLICATIONS IN A VIETNAMESE GIRL

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SUMMARY

An imbalance of glucose and severely high triglycerides are the key characteristics driving hypertriglyceridemia and hyperlipidemic pancreatitis. Hereditary factors considerably promote the increase of triglyceride levels and secondary complications. This study explored the clinical characteristics and genetic causes of a young girl with recurrent severe hypertriglyceridemia-developed pancreatitis at 9 years old and diabetes mellitus appearance at 14 years old. At 20 years old, she was admitted to the hospital in critical condition and had pancreatitis with a triglyceride level of 26.22 mmol/L, HbA1c level of 15%, and a glucose level of 21.3 mmol/L. Whole exome sequencing analysis showed that she had compound heterozygous variants, p.G185C and p.S19W, in the *APOA5* gene, which were reported as risk factors for hypertriglyceridemia, and other variants of uncertainty in the *APOE* and *CFRT* genes. The results and analysis propose that these variations were the underlying causes of elevated plasma triglycerides and led to the onset of metabolic complications at a juvenile age. These findings shed light on the molecular mechanism of elevated plasma triglyceride and the early onset of diabetes following hyperlipidemic pancreatitis in the patient.

Keywords: APOA5, APOE, hyperlipidemic pancreatitis, hypertriglyceridemia, WES

INTRODUCTION

Hypertriglyceridemia (HTG) occurs when the human body exhibits an increased concentration of serum triglycerides (TG). The TG level reaches from 1.7 to 5.6 mmol/L to be considered moderate and severe HTG above 5.6 mmol/L (Grundy *et al.*, 2019). HTG is elicited by a variety of factors, including genetic disorders, obesity, diabetes, alcohol

consumption, smoking, and the intake of certain medications. Elevated levels of lipids in the bloodstream can result in complications of heart disease, pancreatitis, and diabetes (Shemesh, Zafrir, 2019).

Hypertriglyceridemia is multifaceted due to the intricate interplay of both genetic predispositions and environmental factors. APOE, an apolipoprotein E gene, plays a central role in lipoprotein metabolism. The protein apolipoprotein E binds with lipid particles to form a lipoprotein-mediated complex for lipid transport throughout the body. APOE is also essential for the efficient clearance of chylomicron remnants after enzymatic lipolysis in the circulation (Marais, 2019). Variants of APOE (APOE4 protein) can impair the processing of very low-density lipoproteins (VLDL), leading to an imbalance in cholesterol distribution and causing hypertriglyceridemia potentially (Phillips, 2014). APOA5 encodes for an apolipoprotein A5 that significantly contributes to the regulation of triglyceride levels in the bloodstream. APOA5 acts as a stimulator of lipoprotein lipase, which is a crucial enzyme in the breakdown of triglycerides, thereby amplifying the metabolism of TG-rich particles (Su et al., 2018). APOA5 variants have been implicated by countless previous studies as genetically essential in the mechanism of hypertriglyceridemia and the control of plasma triglyceride concentrations (Martin et al., 2003).

High TG levels ($\geq 20 \text{ mmol/L}$) significantly increase the risk of acute pancreatitis (Valdivielso *et al.*, 2014). It is the third main cause, accounting for up to 10% of acute pancreatitis cases to be reported. A genetic susceptibility to dyslipidemia could potentially lead to elevated triglyceride levels. Such individuals

develop pancreatitis more quickly due to their innate incapacity to control their underscoring triglyceride levels. the hereditary role in hastening this condition (Weiss et al., 2021). Multiple genetic variants positioned in LPL (Lipoprotein lipase), APOC2 (Apolipoprotein C-II), APOA5, APOE, LMF1 (Lipase maturation factor 1), (glycosylphosphatidylinositol-*GPIHBP1* anchored high-density lipoprotein binding protein 1) were reported to be associated with HTG and a predisposition to pancreatitis risk (Dron, Hegele, 2020). The CFTR gene encoding for cystic fibrosis transmembrane regulator has also been reported to be a gene that is associated with pancreatic and respiratory diseases (Wang et al., 2014; Wertheim-Tysarowska et al., 2021). Recently, Fujita et al. (2022) were the first to use whole-exome sequencing (WES) analysis on an HTGP (hypertriglyceridemiainduced pancreatitis) complication and revealed genetic materials in LPL, APOA5, APOE. and CFTR (Cystic fibrosis transmembrane conductance regulator) involved in the condition (Fujita et al., 2022).

This study presented the clinical and genetic characteristics of a young Vietnamese patient with severe hypertriglyceridemia and multiple complications, including pancreatitis and diabetes.

CASE PRESENTATION

Patient's clinical characteristics

A 20-year-old Vietnamese female patient was admitted to the Emergency Department of Hanoi Medical University Hospital. The patient was first diagnosed with acute pancreatitis at the age of 9 and was subsequently diagnosed with diabetes mellitus five years later. She has been managing her diabetes with an insulin injection regimen consisting of 8-8-8-14 units. There is no known history of diabetes, metabolic disorders, premature cardiovascular disease, or allergies in her family. No consanguineous marriages were reported. The patient does not have a history of tobacco or alcohol use and has not been on steroid therapy.

At the Emergency Department of Hanoi Medical University Hospital, the patient presented with symptoms of severe abdominal pain and pronounced fatigue. Upon admission, the patient was diagnosed with diabetic ketoacidosis, as evidenced by an arterial blood gas (ABG) test showing hyperglycemia (glucose = 21.3 mmol/L), acidosis (pH = 7.24), and metabolic acidosis (HCO3 = 5.30 mEq/L) (Table 1). Immediate treatment comprised of fluid administration and insulin therapy was provided. As the patient's blood gas levels improved, her abdominal pain subsided. The patient was transferred the Department to of Endocrinology - Respiratory Medicine for further management.

The patient's electrocardiogram (ECG) revealed a normal sinus rhythm (112 bpm) and an intermediate cardiac axis, consistent with normal heart orientation. The cardioankle vascular index (CAVI) was within the normal range on both sides, indicating no significant risk of arterial stiffness or atherosclerosis. Left ventricular size and systolic function were within normal limits as determined by the echocardiogram. Carotid Doppler ultrasound did not find abnormalities in the carotid and spinal arteries on both sides.

The patient's lipid profile was severe HTG, with a TG level of 26.22 mmol/L, cholesterol of 12.35 mmol/L, HDL-

cholesterol of 0.59 mmol/L, and LDLcholesterol of 3.21 mmol/L. The patient's episode of acute pancreatitis symptoms was expressed in abdominal pain, elevated serum 39.8 g/L, a high total white blood cell (WBC) count of 472, and hyperglycemia (glucose = 21.3 mmol/L) (Table 1). An abdominal ultrasound revealed acute pancreatitis, hepatic steatosis, and gallstones. The pancreatic, intrahepatic, and extrahepatic bile ducts were mildly dilated, possibly indicating obstruction or inflammation due to compression by the enlarged pancreatic head or gallstones. The scan also revealed a fatty liver, mild hepatomegaly, and a left angiomyolipoma. A subsequent renal computed tomography (CT) scan of the abdomen showed an enlarged upper pancreatic head with surrounding fat infiltration. The severity of the condition was assessed using the Balthazar score (C), the computed tomography severity index (CTSI, 4), and the Glasgow score (2), all of which indicated moderate severity.

The patient received glycemic control and fluid infusions and underwent lipidlowering therapy with omega-3 fatty acid treatment during inpatient treatment. After 10 days of hospitalization, her serum TG has declined from a severe to a moderate level (5.04 mmol/L). The levels of aspartate aminotransferase (AST) and alanine aminotransferase (ALT) were within the normal range, indicating no liver damage (13/14 U/L). The serum concentrations of sodium (Na) and potassium (K) were also normal, reflecting adequate electrolyte balance (141/3.8 mmol/L). The activities of the digestive enzyme amylase (83 U/L) and lipase (21.3 U/L) were controlled and below the threshold, suggesting no pancreatic inflammation. The index on the ABG test improved when with condition was stable

and the occurrence of ketoacidosis ceased (pH = 7.411). The metabolic acidosis condition was resolved with paCO₂ raised to 29.2 mmHg and HCO3 enhanced to 18.1 mEq/L (Table 2). Blood glucose levels were

gradually normalized under appropriate management, no indications of pancreatitis were observed, and the patient was discharged after 10 days of hospital treatment.

Biochemic	cal						
	Index	Normal			Index	Normal	
Glucose	21.3	4.0-5.4	mml/L	Са	2.38	2.15 - 2.55	mmol/L
HbA1c	15.00%	≤ 5.7%		Cholesterol	12.35	< 5.5	mmol/L
C-peptid	0.21	0.26-1.27	nmol/L	Triglyceride	26.22	< 1.69	mmol/L
Creatinin	39	52.2-91.9	µmol/l	HDL	0.59	> 1.69	mmol/L
Ure	5.4	2.1-8.5	mmol/L	LDL	3.21	> 2.0	mmol/L
Na	121	135-145	mmol/L	GOT/GPT	10/9	>35	U/L
К	4.7	3.5-5.0	mmol/L	Amlylase	27	30-110	U/L
hs-CRP	2.68	<3.0	mg/dL				
Arterial bl	ood gas						
	Index	Normal			Index	Normal	
pН	7.24	7.35 -7.45		paO2	131	75-100	mmHg
paCO2	12.7	35-45	mmHg	HCO3	5.3	22-26	mEq/L
Endocrine	•						
	Index	Normal			Index	Normal	
Cortisol	633	140-690	nmol/L	TSH	0.567	0.4 - 4.0	µUI/mL
ACTH	142	7.2-63.3	pg/mL	Albumin	39.8	34-45	g/L
FT4	13.3	12.0-22.0	pmol/L				
Hematolog	gical						
	Index	Normal			Index	Normal	
HC	4.56	36-44	T/L	PT	89%	70-140%	
Hb	135	116-150	g/L	APTT	1.03	0.85-1.25	
BC	9.7 g/L	115-165	g/L	INR	1.06	<1.5	
ТС	472	4.5-11	(x 10^9/L)	Fibrinogen	8.78	2.0-4.0	g/L
Urine							
Ketones	(-)			Protein	(-)		

Table	1	Patient's	clinical	indicators	on	admission	dav
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HbA1c: Hemoglobin Ac1; Na: Sodium; K: Potassium; Ca: Calcium; HDL: high-density lipoprotein; LDL: low-density lipoprotein; GOT: Glutamic Oxaloacetic Transaminase; GPT: Glutamic Pyruvic Transaminase; hs-CRP: High-sensitivity C-reactive protein; ACTH: Adrenocorticotropic hormone; FT4: Free Thyroxine; TSH: Thyroid Stimulating Hormone; HC: Hematocrit; HB: Hemoglobin; BC: Blood count; TC: Total count; PT: Prothrombin time; APTT: Activated partial thromboplastin time; INR: International normalized ratio.

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	Admission	Discharge	Normal range	
Biochemical				
Na	121	141	135 - 145	mmol/L
к	4.7	3.8	3.6-5.2	mmol/L
Triglyceride	26.22	5.04	<1.7	mmol/L
GOT	10	13	7 - 56	U/L
GPT	9	14	7 - 35	U/L
Amylase	27	83	30 - 110	U/L
Lipase	47.2	21.3	10 - 140	U/L
hs-CRP	2.68	0.45	≤ 3.0	mg/dL
ABG				
рН	7.24	7.411	7.35 - 7.45	
paCO2	12.7	29.2	35 - 45	mmHg
paO2	131	110.9	85 - 100	mmHg
HCO3	5.3	18.1	21 - 28	mEq/L

Fable 2. Clinica	l progress	of the	patient	at discharge.
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Na: Sodium; K: Potassium; GOT: Glutamic Oxaloacetic Transaminase; GPT: Glutamic Pyruvic Transaminase; hs-CRP: High-sensitivity C-reactive protein; ABG: Arterial blood gas.

Genetic findings

A genetic test was performed after the patient received counseling from medical professionals (Supplements 1). The consent documents were written and signed. The genomic DNA of the patient was extracted from peripheral blood and whole-exome sequencing underwent (WES) analysis. The variants of metabolism-related genes were concentrated on screening causative factors according to criteria described in the

Supplement methods. As a result, one homozygous variant in the APOE, two heterozygous variants in the APOA5, and two heterozygous variants in the CFTR genes were detected (Table 3). Among these variants, only two heterozygous c.553G>T variants, (p.G185C) and c.56C>G (p.S19W), of the APOA5 gene were classified as the risk factors for hypertriglyceridemia condition, according to the ClinVar Database. The presence of these two variants in the patient was validated by Sanger sequencing (Fig. 1).

Gene	Variant			Allele Frequency			<i>In silico</i> prediction
	cDNA	Amino acid	Zygosity	gnomAD	VN GVDB	ClinVar	SIFT/ MutationTaster / PROVEAN
APOE	c.466T>C	p.C156R	hom	0.1485		Conflicting	T/P/N
APOA	c.553G>T	p.G185C	het	0.0062	0.037	Risk factor	D/N/D
5	c.56C>G	p.S19W	het	0.0676	0.003	Risk factor	D/N/N
CFTR	c.1666A>G	p.I556V	het	0.0034	0.047	Conflicting	T/A/N
	c.1251C>A	p.N417K	het	0.0026		Conflicting	T/N/N

Table 3. Variants of metabolism-related genes extracted from WES.

VN GVDB: Vietnamese Genetic Variation Database (NCBI Bioproject ID: 515199); hom: homozygous; het: heterozygous; A: disease-causing automatic; D: Damaging/ Deleterious; P: polymorphism automatic; T: Tolerate; N: Polymorphism - probably harmless (MutationTaster)/ Neutral (Provean); (.): unknown



Figure 1. Sanger sequencing results of the *APOA5* p.G185C and p.S19W variants detected in the patient.

DISCUSSION

In this report, the patient presented with the main symptoms of diabetic ketoacidosis, HTG, and pancreatitis. Besides that, she also harbored the compound heterozygous variants p.G185C and p.S19W of the *APOA5* gene, which are associated with hypertriglyceridemia, and variants of the *APOE* and *CFTR* genes with

conflicting interpretations of pathogenicity. Her clinical expression and genetic findings are quite similar to two those in previous reports. The first one has described two patients, at the age of 12, who suffered acute pancreatitis from diabetic ketoacidosis and HTG. However, this report lacked a comprehensive analysis of the patients' genetic aspects, despite their young age and clean, healthy medical history (Bouchaala et al., 2020). The second one reported the genetic finding of a young HTG patient from Japan with recurrent acute pancreatitis (Fujita et al., 2022). All the patients first sustained high-level TG at a young age, then complications onset later, and they have no smoking or alcohol history. The Japanese patient was found to have two heterozygous variants in the APOA5 gene, including the p.G185C, three heterozygous in the CFTR gene, and one in the APOE gene. They indicated variants in HTG-linked genes (APOA5 and APOE) and CFTR associated with acute pancreatitis could be attributed to the patient severe pathology.

Meta-analysis studies showed that the common APOA5 p.G185C (rs2075291) variant is an HTG risk factor with an impact on TG lipid levels, specifically in Asians (He et al., 2016; Qian et al., 2018). Heterozygous variant p.G185C was also found among subjects with a history of acute pancreatitis (Khovidhunkit et al., 2016). Functional analysis demonstrated that the homozygous variant p.G185C reduces the hydrolysis of VLDL by decreasing the activation of lipoprotein lipase (LPL) (Huang et al., 2012). The accumulation of unprocessed TG-rich lipoproteins suggests the pathogenic impact of p.G185C on patients with severe HTG conditions. The APOA5 p.S19W variant was also reported to be positively correlated with upraised TG and severe HTG in the

Caucasian and Asian populations (Dallongeville et al., 2008; Evans et al., 2011). It has also been considered a biomarker for hyperlipoproteinemia (Wang et al., 2008). With the above evidence, the simultaneous variants. appearance of two APOA5 p.G185C, and p.S19W, could be considered main cause of the patient's the hyperlipoproteinemia condition.

In this study, the appearance of "Variants of Uncertain Significance, VUS" on the APOE and CFTR genes might be followed up. The APOE p.C156R (rs429358) variant appeared widely and was found to be associated with HDL-cholesterol and lipid levels in Europe and southern China (Deng et al., 2021; Richardson et al., 2020). The altered cysteine-arginine interchanges were shown to exert an influence on the configuration of human E apoprotein isoforms ε -4, ε -3, and ε -2 (Weisgraber et al., 1981). Hence, the homozygous variant p.C156R in the APOE gene may synergistically affect lipid metabolism together with variants p.G185C and p.S19W of the APOA5 gene, resulting in serious development of HTG in our patient. In addition, CFTR polymorphisms were believed to contribute to the genetic etiology of acute pancreatitis in young HTG Japanese patients (Fujita et al., 2022). The CFTR p.I566V was established most frequently in idiopathic and acute pancreatitis patients in the Chinese and Japanese populations (Chang et al., 2007; Iso et al., 2019). In the first study of the relationship between CFTR variants and hyperlipidemic pancreatitis, p.I566V was the most common variant present in patients with HTG with the onset of hyperlipidemic pancreatitis (HLP) compared with patients with HTG who do not have HLP (Chang et al., 2008). p.I566V may cause conditions like pancreatitis through lowered open probability channel activity, resulting

in a reduction in current density in the chloride (Cl⁻) current throughout the cell (Lee et al., 2003). p.N417K was predicted as "deleterious" via computational analysis and appeared in cystic fibrosis (CF), and CFTRrelated patients on clinical genetic screening (Trujillano et al., 2015; George Priya Doss et al., 2008). Impaired chloride transport could disturb the pancreas' fluid transport and electrolyte balance, resulting in pancreatitis (Hegyi et al., 2016). In people with CF, the pancreatic transport tube thickens blocked digestive enzymes and produces sticky mucus. This evinces a combination of p.I566V and p.N417K that could defect CFTR protein function and provoke pancreatitis progression. While the existing information on the CFTR variant is inconclusive, our future focus will center on considering and vigilantly monitoring the patient's clinical symptoms. Gallstones detected by abdominal ultrasound could indeed be one of the causes of pancreatitis. However, it was unclear when the gallstone formation occurred since the patient presented symptoms of pancreatitis-induced TG at 9 years old.

Hypertriglyceridemia-induced

pancreatitis presents TG levels exceeding 20 mmol/L; our patient reached the condition threshold at 26.22 mmol/L (Valdivielso et al., 2014). The patient has also been on insulin treatment for diabetes mellitus since 2017. Diabetes is reported as the most commonly seen secondary factor in individuals with HTGP cohorts (Zafrir et al., 2019). Furthermore, diabetes is listed as the primary metabolic driver of HLP due to the association between HTG and diabetes (Shemesh, Zafrir, 2019). Treatment of severe HTG with pancreatitis and diabetes mellitus complications includes both lifestyle innovation with diet and the uptake

of manageable medications. The patient is counseled on a diet rich in dietary fiber and physical activity. She received guidelines for self-administering insulin and recognizing and managing hypoglycemia before discharge. A regular examination is required for her every 2–3 months. Fibrates, Omega-3 fatty acids, and niacin are the cornerstones of HTG therapy, with proficiency in reducing TG levels, hepatic VLDL production, and lipid-regulating agents (Shemesh, Zafrir, 2019). HTGinduced pancreatitis is thought to be treated similarly to various types of inflammation. Interestingly, some studies suggested that anti-diabetes drugs and heparin lower the risk of pancreatitis by promoting LDL activity (Lai et al., 2011).

In this study, we documented a young Vietnamese patient who suffered from complications caused by severe HTG and analyzed the genetic aspect. Applying whole-exome sequencing analysis, we detected two molecular genetic variants in the APOA5 gene, p.G185, and p.S19W, and a homozygous variant, p.C156R, in the APOE gene as the underlying cause of the patient's susceptibility to high serum TG levels. The compound heterozygous variants p.I556V and p.N417K in the CFTR gene could predispose to developing early hyperlipidemic pancreatitis. Further observation of the patient's pancreatic and respiratory conditions is required to draw a definitive conclusion. The identification of variants related to HTG and complications is valuable in confirming clinical diagnoses as well as identifying potential high-risk profiles among HTG cases.

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REFERENCES

Bouchaala K, Bahloul M, Bradii S, Kallel H, Chtara K, Bouaziz M (2020) Acute Pancreatitis Induced by Diabetic Ketoacidosis with Major Hypertriglyceridemia: Report of Four Cases. *Case Rep Crit Care* 2020.

Chang MC, Chang YT, Wei SC, Tien YW, Liang PC, Jan IS, Su YN, Wong JM (2007) Spectrum of mutations and variants/haplotypes of CFTR and genotype-phenotype correlation in idiopathic chronic pancreatitis and controls in Chinese by complete analysis. *Clin Genet* 71(6): 530-539.

Chang YT, Chang MC, Su TC, Liang PC, Su YN, Kuo CH, Wei SC, Wong JM (2008) Association of Cystic Fibrosis Transmembrane Conductance Regulator (CFTR) mutation/variant/haplotype and Tumor Necrosis Factor (TNF) promoter polymorphism in hyperlipidemic pancreatitis. *Clin Chem* 54(1): 131-138.

Dallongeville J, Cottel D, Wagner A, Ducimetière P, Ruidavets JB, Arveiler D, Bingham A, Ferrières J, Amouyel P, Meirhaeghe A (2008) The APOA5 Trp19 allele is associated with metabolic syndrome via its association with plasma triglycerides. *BMC Med Genet* 9: 1-9.

Deng X, Hou J, Deng Q, Zhong Z (2021) Association between the APOE gene polymorphism and lipid profile and the risk of atrial fibrillation. *Lipids Health Dis* 20(1): 1-13.

Dron JS, Hegele RA (2020) Genetics of Hypertriglyceridemia. Front Endocrinol (Lausanne) 11: 455.

Evans D, Aberle J, Beil FU (2011) Resequencing the Apolipoprotein A5 (APOA5) gene in patients with various forms of hypertriglyceridemia. *Atherosclerosis* 219(2): 715-720.

Fujita S, Nishizawa H, Miyashita Y, Imada T, Yamaguchi T, Murano T, Bujo H, Asano Y, Kozawa J, Maeda N, Shimomura I (2022) Genetic assessment using whole-exome sequencing for a young hypertriglyceridemic patient with repeated acute pancreatitis. *Endocr* J 69(9): 1101-1108.

George Priya Doss C, Rajasekaran R, Sudandiradoss C, Ramanathan K, Purohit R, Sethumadhavan R (2008) A novel computational and structural analysis of nsSNPs in CFTR gene. *Genomic Med* 2(1-2): 23-32.

Grundy SM, Stone NJ, Bailey AL, Beam C, Birtcher KK, Blumenthal RS, Braun LT, de Ferranti S, Faiella-Tommasino J, Forman DE, Goldberg R, Heidenreich PA, Hlatky MA, Jones DW, Lloyd-Jones D, Lopez-Pajares N, Ndumele CE, Orringer CE, Peralta CA, Saseen JJ, Smith SC, Sperling L, Virani SS, Yeboah J (2019) 2018 AHA/ACC/AACVPR/AAPA/ABC/ACPM/AD A/AGS/APhA/ASPC/NLA/PCNA Guideline on Management of Blood Cholesterol: the Executive Summary: A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. J Am Coll Cardiol 73(24): 3168-3209.

He H, Lei L, Chen E, Dong J, Zhang K, Yang J (2016) The c.553G>T Genetic Variant of the APOA5 Gene and Altered Triglyceride Levels in the Asian Population: A Meta-Analysis of Case-Control Studies. *Genet Test Mol Biomarkers* 20(12): 758-765.

Hegyi P, Wilschanski M, Muallem S, Lukacs GL, Sahin-Tóth M, Uc A, Gray MA, Rakonczay Z, Maléth J (2016) CFTR: A new horizon in the pathomechanism and treatment of pancreatitis. *Rev Physiol Biochem Pharmacol* 170: 37-66.

Huang YJ, Lin YL, Chiang CI, Yen CT, Lin SW, Kao JT (2012) Functional importance of apolipoprotein A5 185G in the activation of lipoprotein lipase. *Clinica Chimica Acta* 413(1-2): 246-250.

Iso M, Suzuki M, Yanagi K, Minowa K, Sakurai Y, Nakano S, Satou K, Shimizu T, Kaname T (2019) The CFTR gene variants in Japanese children with idiopathic pancreatitis. *Hum Genome Var* 6(1): 17.

Khovidhunkit W, Charoen S, Kiateprungvej A,

Chartyingcharoen P, Muanpetch S, Plengpanich W (2016) Rare and common variants in LPL and APOA5 in Thai subjects with severe hypertriglyceridemia: A resequencing approach. *J Clin Lipidol* 10(3): 505-511.e1.

Lai SW, Muo CH, Liao KF, Sung FC, Chen PC (2011) Risk of acute pancreatitis in type 2 diabetes and risk reduction on anti-diabetic drugs: A population-based cohort study in taiwan. *Am J Gastroenterol* 106(9): 1697-1704.

Lee JH, Choi JH, Namkung W, Hanrahan JW, Chang J, Song SY, Park SW, Kim DS, Yoon JH, Suh Y, Jang IJ, Nam JH, Kim SJ, Cho MO, Lee JE, Kim KH, Lee MG (2003) A haplotype-based molecular analysis of CFTR mutations associated with respiratory and pancreatic diseases. *Hum Mol Genet* 12(18): 2321-2332.

Marais AD (2019) Apolipoprotein E in lipoprotein metabolism, health and cardiovascular disease. *Pathology* 51(2): 165-176.

Martin S, Nicaud V, Humphries SE, Talmud PJ (2003) Contribution of APOA5 gene variants to plasma triglyceride determination and to the response to both fat and glucose tolerance challenges. *Biochim Biophys Acta Mol Basis Dis* 1637(3): 217-225.

Phillips MC (2014) Apolipoprotein E isoforms and lipoprotein metabolism. *IUBMB Life* 66(9): 616-623.

Qian X, Li Y, Liu X, Li L, Yang K, Liu R, Zhang H, Shi Y, Yu F, Mao Z, Bie R, Wang C (2018) The "T" allele of apolipoprotein A5 rs2075291 is significantly associated with higher total cholesterol and triglyceride and lower high-density lipoprotein cholesterol levels in Asians: a meta-analysis. *Nutr Res* 56: 11-22.

Richardson TG, Sanderson E, Palmerid TM, Korpelaid MA, Ference BA, Smith GD, Holmes M V. (2020) Evaluating the relationship between circulating lipoprotein lipids and apolipoproteins with risk of coronary heart disease: A multivariable Mendelian randomisation analysis. *PLoS Med* 17(3): e1003062.

Shemesh E, Zafrir B (2019) Hypertriglyceridemia-related pancreatitis in patients with type 2 diabetes: Links and risks. *Diabetes Metab Syndr Obes* 12: 2041-2052.

Su X, Kong Y, Peng D (2018) New insights into apolipoprotein A5 in controlling lipoprotein metabolism in obesity and the metabolic syndrome patients. *Lipids Health Dis* 17(1): 174.

Trujillano D, Weiss MER, Köster J, Papachristos EB, Werber M, Kandaswamy KK, Marais A, Eichler S, Creed J, Baysal E, Jaber IY, Mehaney DA, Farra C, Rolfs A (2015) Validation of a semiconductor next-generation sequencing assay for the clinical genetic screening of cftr. *Mol Genet Genomic Med* 3(5): 396-403.

Valdivielso P, Ramírez-Bueno A, Ewald N (2014) Current knowledge of hypertriglyceridemic pancreatitis. *Eur J Intern Med* 25(8): 689-694.

Wang J, Ban MR, Kennedy BA, Anand S, Yusuf S, Huff MW, Pollex RL, Hegele RA (2008) APOA5 genetic variants are markers for classic hyperlipoproteinemia phenotypes and hypertriglyceridemia. *Nat Clin Pract Cardiovasc Med* 5(11): 730-737.

Wang Y, Wrennall JA, Cai Z, Li H, Sheppard DN (2014) Understanding how cystic fibrosis mutations disrupt CFTR function: From single molecules to animal models. *Int J Biochem Cell Biol* 52: 47-57.

Weisgraber KH, Rall SC, Mahley RW (1981) Human E apoprotein heterogeneity. Cysteinearginine interchanges in the amino acid sequence of the apo-E isoforms. *J Biol Chem* 256(17): 9077-9083.

Weiss FU, Laemmerhirt F, Lerch MM (2021) Acute Pancreatitis: Genetic Risk and Clinical Implications. *J Clin Med* 10(2): 1-13.

Wertheim-Tysarowska K, Oracz G, Rygiel AM (2021) Genetic risk factors in early-onset nonalcoholic chronic pancreatitis: An update. *Genes (Basel)* 12(5): 785.

Zafrir B, Saliba W, Jubran A, Hijazi R, Shapira C (2019) Severe Hypertriglyceridemia-Related Pancreatitis: characteristics and predictors of recurrence. *Pancreas* 48(2): 182-186.