# EFFICIENCY OF USING PROBIOTIC ELAC-GROW AND HAN-PROWAY IN LAYING-EGG HENS

## Dinh Thi Tuyet Van<sup>1</sup>, Nguyen Thi Thu<sup>1</sup>, Nguyen Hong Linh<sup>1</sup>, Nguyen Thi Men<sup>1</sup>, Tran Van Khanh<sup>1</sup>, Le Thi Nhi Cong<sup>2,⊠</sup>

<sup>1</sup>Hanvet Pharmaceutical Limited Company, Pho Noi Industrial Park A, Ban-Yen Nhan, My Hao District, Hungyen Province, Vietnam <sup>2</sup>Institute of Biotechnology, Vietnam Academy of Science and Technology, 18 Hoang Quoc Viet Road, Cau Giay District, Hanoi, Vietnam

<sup>III</sup>To whom correspondence should be addressed. E-mail: lenhicong@ibt.ac.vn

Received: 19.5.2021 Accepted: 22.6.2021

## SUMMARY

Industrial chicken farm has to face many challenges such as epidermis and environmental pollution. Antibiotics and synthetic antimicrobial agents are routinely used to alleviate stress as well as to elevate growth and feed efficiency. However, using antibiotic had many negative effects, especially antibiotic resistance; therefore, probiotic products are a bio-secure solution being popularly applied in livestock husbandry. The study was carried out on the two probiotics Elac-grow, a diet supplement, and Han-proway, a waste treatment product, in industrial egg-laying hens. There were two lodgings (test and control) in these experiments; each had 4000 23-week-old hens of Hyline breed. Two probiotics were applied to the test and none to the control. The results showed that there was not significantly difference of egg yield between the two groups, although, the test lodging had higher egg yield growing ratio (67.48%) than the control one (65.73%). Egg quality in the experimental lodging was also significantly improved in the damaged egg ratio, the egg weight and the egg yolk weight after using Elac-grow. The efficiency of using Elac-grow and Han-proway was also shown through the concentration of ammonia emission in animal lodging. The ammonia value of the test lodging was maximum 5 ppm and remained stable at this point, lower than of the control. This efficiency was shown significantly in the microbial fluctuation analysis, when the total bacteria and coliform counts were reduced in the test lodging sequentially above  $1 \times 10^8$  CFU bacteria/g and above  $1 \times 10^7$  CFU coliform/g, the control had uptrend sequentially above  $2 \times 10^9$  CFU bacteria/g and above  $1 \times 10^8$  CFU coliform/g.

Keyword: Ammonia value, Elac-grow, egg yield, egg quality, Han-proway, probiotics

## INTRODUCTION

Probiotics are defined as live microbial feed supplements that benefit host animals by improving their intestinal microbial balance (AFRC, 1989). The addition of probiotics to diets has been demonstrated to improve growth performance and the feed conversion ratio in broilers (Nahashon *et al.*, 1996a), as well as their egg mass, egg weight, egg size in layers, (Nahashon *et al.*, 1996b), feed consumption (Nahashon *et al.*, 1994) and egg-specific gravity (Nahashon *et al.*, 1996b) in layers.

Many researches showed that the mode of action of probiotics in poultry included maintaining normal intestinal micro flora by competitive exclusion and antagonism, altering metabolism by increasing digestive enzyme activity and decreasing bacterial enzyme activity

and ammonia production, and improving feed intake and digestion neutralizing enterotoxins (Jin et al., 1997). Lactobacillus spp. and Enterococcus spp. are the most commonly used species in the production of probiotics. Enterotoxin produced by pathogenic bacteria can be neutralized by the substances produced by these species. Zhang et al (2019) showed that the dietary supplementation with Enterococcus faecium did not affect the average daily egg weight, cracked egg rate, mortality and egg quality. However, Enterococcus faecium supplementation caused a significantly increased laying rate and decreased feed/egg ratio. The differences in caecal microbiota between test group and the control were significant (Zhang et al., 2019).

circumstances, Moreover, under such antibiotics and synthetic antimicrobial agents are often used to alleviate stress and to elevate growth and feed efficiency. However. injudicious and persistent sub-therapeutic antibiotics addition in poultry feed have become undesirable due to their residues in meat products and development of antibiotic-resistant bacterial population (Jin et al., 1997). In Europe, the use of antibiotics as growth promoting agents in poultry has been banned. Due to several negative effects of antibiotics, these have been gradually replaced by probiotics (AFRC, 1989), (Patterson and Burkholder, 2003; Dhama et al., 2011).

On the other hand, breeders were also concerned about the waste treatment, which directly affect hens and people. The use of probiotic additives has lowered the quantity of nitrogen in waste effluent, which potentially represents a gain in feed efficiency and reduced nitrogen requirements in diet formulations, resulting in a reduction of leached nitrogen in the farm and the surrounding environment (Applegate *et al.*, 2010).

With the purpose of providing bio-secure livestock solution, two probiotics were produced by Hanvet Pharmaceutical Limited Company: Elac-grow for hen's diets and Han-proway for waste treatment. The aim of the present study was to investigate the effects of these two commercial probiotics Elac-grow for feed diet and Han-proway for waste treatment on, egg quantity, egg quality, emission evaluation and microbial fluctuation in laying hens.

## MATERIALS AND METHODS

## **Commercial probiotics**

Elac-grow for feeding and Han-proway for waste treatment are produced by Hanvet Pharmaceutical Limited Company. For this experiment, 1 kg of Elac-grow was used for 1000 kg feed, 3 days per week, 1 kg of Han-proway was used for 80-100 m<sup>2</sup> composite, periodically every 7-10 days. Components of 1 kg Elac-grow include *Enterococcus faecium* (7x10<sup>10</sup> CFU), mix of vitamin (A, D2, E, B2, B6, B12, K, C), ZnSO<sub>4</sub>, MgSO<sub>4</sub>, CuSO<sub>4</sub>, calcium, DLmethionine, niacinamide, citric acid, lysine. Components of 1 kg Han-proway include Bacillus licheniformis (2.7x10<sup>9</sup> CFU), Bacillus megaterium  $(2.5 \times 10^9 \text{ CFU}),$ Lactobacillus acidophilus (3x10<sup>9</sup> CFU).

## Laying hens and farm facilities

The experiment was performed at Hoa Phat farm, a laying hen farm in Dong Anh, Hanoi, Vietnam. It is an area of  $500 \text{ m}^2$  with a system of eight exhaust fans in the rear and four rows of double nest. Two animal lodgings were set up for this experiment; each of them had 4000 laying hens. All laying hens belonged to the same Hyline brown variety at the age of 23 weeks.

## **Experimental design**

Experiment lodging: using probiotic Elacgrow for feed and Han-proway for waste treatment; control lodging: did not use probiotic for feed and for waste treatment.

## Effect of using probiotic in diet to egg production

All of the eggs were collected and weighted individually to determine the total number of the egg and the egg weight. Using these values, egg production, birth rate and egg weight were calculated. Egg yield (egg) = The total number of egg

Birth rate (%) =  $\frac{the number of eggs \times 100}{c}$ the number of hens Egg weight  $(g/egg) = \frac{the \ sumery \ of \ egg \ weight \ (gr)}{dr}$ 

the number of eggs (egg)

Egg quality was determined. Thirty eggs were collected randomly from each lodging. Each egg was estimated, the york weighted, york color checked to determine egg quality

## Effect of using probiotic for waste treatment

Gas detector of Bosean BH-4S (China) was used to test the amount of ammonia in the lodging. Ammonia level measurement was carried out during the day from 11 am to 12 am to get the highest ammonia level caused by evaporation that occurred due to heat.

## The fluctuation of microbial in the waste was analyzed in the laboratory

Compost analysis was examined for Coliforms and total bacteria by viable cell numeration method. Coliforms were analyzed on Mackonkey agar (Merck), total bacteria were analyzed on Tryptic Soy Agar (TSA, Merck). Compost samples were collected from the top layer of the middle lodging from 16 pm to 17 pm.

#### **Statistical analysis**

All results were statistically analyzed by Excel 2010.

## **RESULTS AND DICUSSION**

## Effect of using probiotic in diet

## Egg laying rate and egg yield

Nahashon et al. (1994a,b, 1996b) and Tortuero and Fernandez (1995) reported that there were significant increases in egg production of laying hens by using various levels of probiotic supplementation. Our result showed the same result that there was not a significant increase in the egg yield between test and control. However, based on Figure 1, although the birth rate of the control lodging was higher than that of the test lodging, the speed of growing egg yield in the test lodging was higher than that in the control lodging after one week of using probiotics. The egg laying rate in test lodging increased from 19.7 to 87.18%, when this value in control lodging increased more slowly from 23.39 to 89.12%. Therefore, egg yield growing rate of the test was 67.42% higher than 65.73% in the control.

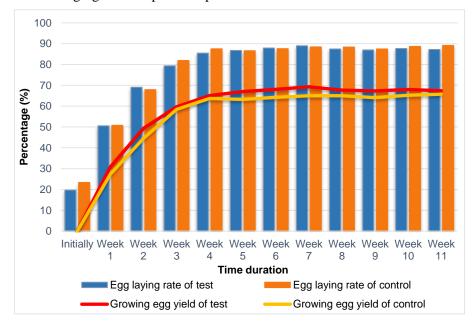


Figure 1. Comparison of the egg laying rate and the growing egg yield between test lodging and control lodging.

## Quality of egg

The damaged egg ratio was based on the hardness of eggshell through calcium supplementation in the diet. In spite of having higher damaged egg rate initially, this rate of the test lodging was improved immediately after adding Elac-grow in the diets for one week and maintained a lower value than the control lodging throughout experiment (Figure 2). After 11 weeks, the damaged egg rate of the test lodging was 0.15% lower than 0.23% of the control (P<0.05). Balevi *et al* (2001) reported that the damaged egg ratio was lower (0.76%) than the control (1.42%) with the 500 mg/kg probiotic supplementation into laying hens' diets (p<0.05) (Balevi *et al.* 2001). Kurtoglu *et al.* (2003) also reported similar results (Kurtoglu *et al.*, 2004).

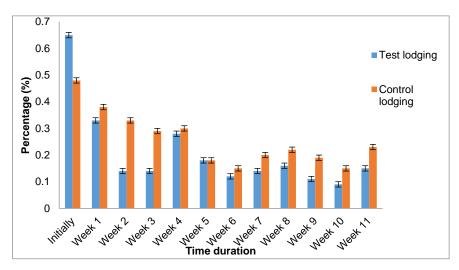


Figure 2. The damaged egg ratio of test lodging and control lodging.

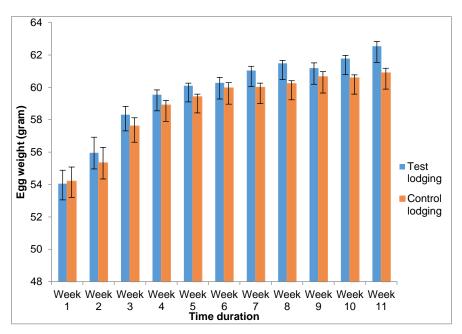


Figure 3. The average egg weight value of test lodging and control lodging.

## Vietnam Journal of Biotechnology 20(1): 143-150, 2022

The egg weight of test lodging was also improved after two weeks of using Elac-grow. This value of test lodging increased 8.25 units from 54.05 to 62.54 g, when the control lodging only increased 7.27 units from 54.2 to 60.89 g (P<0.05). This was similar to the result in the publication of Tortuero and Fernandez (1995) and Nahashon *et al.* (1994b), which observed that significant increases in egg weight were obtained from probiotic addition (Tortuero, Fernández 1995; Nahashon *et al.*, 1994).

The egg yolk weight of the test lodging was higher than that of the control. In the test lodging, the egg yolk weight increased from 11.2 to 14.49 g, this value raised from 11.5 to 14.42 g in the control one (P<0.05) (Figure 4).

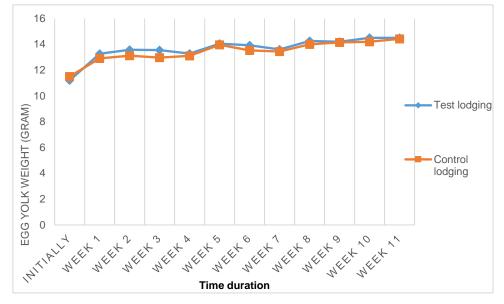


Figure 4. The egg yolk weight of test lodging and control lodging.

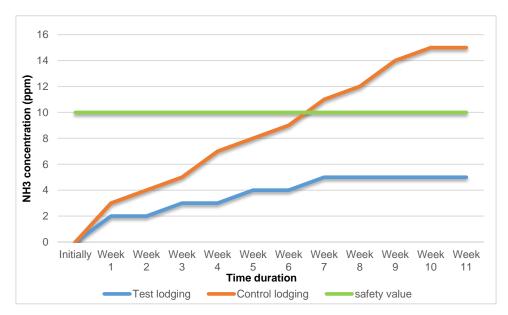


Figure 5. Comparison of NH<sub>3</sub> concentration in the test lodging and in the control lodging.

 $1.68 \pm 0.27 \times 10^8$ 

 $1.74 \pm 0.25 \times 10^{8}$ 

 $1.82 \pm 0.27 \times 10^8$ 

 $1.92 \pm 0.21 \times 10^{8}$ 

#### **Evaluate emissions in the lodging**

Hens' health, egg quantity and quality are not only affected by the diets, but also by the surrounding environment. Ammonia is a gas present in the atmosphere of every poultry house. It results from the chemical decomposition of uric acid in droppings by certain bacteria in the litter (TahseenAziz, 2010). According to National Technical Regulation poultry farms were considered biosecurity when ammonia concentration in the lodging was less than 10 ppm (QCVN01-15: 2010/BNNPTNT). Figure 5 showed that by using the probiotic Han-proway for waste treatment, the test lodging had lower ammonia value of 5 ppm than the control of 15 ppm and the safety value of 10 ppm. Research of Song et al in 2012 showed that poultry manure is one of the major sources of nitrogen pollution (Song et al., 2012), of which ammonia is a major aerial pollutant, with adverse effects on the production of broilers (Miles et al., 2004; Wathes

and Kristensen, 2000). Park et at showed that laying hens fed the probiotic Enterococcus faecium supplementations had lower ammonia emission than those fed the control treatment (Park et al., 2016).

## The fluctuation of microbial in the waste

The fluctuation of microbial in the waste showed significant difference after using probiotic solution, especially in coliform density (Table 1). When the total bacteria and coliform density of test lodging decreased and stabilized at more than  $1 \times 10^8$  CFU bacteria/g and more than  $1 \times 10^7$  CFU coliform/g, these value in the control increased rapidly with more than 2x10<sup>9</sup> CFU bacteria/g and more than  $1 \times 10^8$  CFU coliform/g. This result was the same with the research of Park et al (2016), coliform counts were reduced with the probiotic treatment as compared with the control treatment (P < 0.05) (Park et al., 2016).

Time duration	Total microbial density (CFU/g)		Coliform density (CFU/g)	
Time duration	Test lodging	Control lodging	Test lodging	Control lodging
Initially	2.12 ± 0.13×10 <sup>8</sup>	$3.12 \pm 0.15 \times 10^8$	1.12 ± 0.19×10 <sup>8</sup>	1.67 ± 0.15×10 <sup>8</sup>
Week 1	$2.05 \pm 0.21 \times 10^8$	$5.23 \pm 0.17 \times 10^8$	$8.95 \pm 0.28 \times 10^7$	1.55 ± 0.25×10 <sup>8</sup>
Week 2	1.98 ± 0.19×10 <sup>8</sup>	$5.5 \pm 0.15 \times 10^8$	$6.65 \pm 0.24 \times 10^7$	1.75 ± 0.18×10 <sup>8</sup>
Week 3	1.85 ± 0.22×10 <sup>8</sup>	$1.35 \pm 0.2 \times 10^9$	$4.25 \pm 0.18 \times 10^{7}$	1.23 ± 0.24×10 <sup>8</sup>
Week 4	1.92 ± 0.18×10 <sup>8</sup>	$1.22 \pm 0.14 \times 10^9$	$3.45 \pm 0.12 \times 10^7$	1.35 ± 0.21×10 <sup>8</sup>
Week 5	1.76 ± 0.25×10 <sup>8</sup>	1.45 ± 0.27×10 <sup>9</sup>	$2.25 \pm 0.22 \times 10^7$	1.74 ± 0.19×10 <sup>8</sup>
Week 6	$1.5 \pm 0.17 \times 10^8$	1.55 ± 0.13×10 <sup>9</sup>	$2.12 \pm 0.18 \times 10^7$	1.67 ± 0.21×10 <sup>8</sup>
Week 7	1.56 ± 0.13×10 <sup>8</sup>	2.12 ± 0.17×10 <sup>9</sup>	$1.65 \pm 0.2 \times 10^7$	1.98 ± 0.19×10 <sup>8</sup>

 $2.25 \pm 0.25 \times 10^9$ 

 $2.5 \pm 0.21 \times 10^9$ 

2.45 ± 0.25×10<sup>9</sup>

 $2.55 \pm 0.2 \times 10^9$ 

Table 1. Fluctuation of microbial density and colifor	orm density.	v.
-------------------------------------------------------	--------------	----

 $1.72 \pm 0.21 \times 10^{8}$ 

 $1.32 \pm 0.19 \times 10^{8}$ 

 $1.35 \pm 0.21 \times 10^{8}$ 

 $1.22 \pm 0.15 \times 10^{8}$ 

#### CONCLUSION

Week 8

Week 9

Week 10

Week 11

We could realize a slight improvement of birth rate and growing egg yield in test group, however, further investigation should be carried out to address this effect, the test lodging had higher egg yield growing ratio of 67.48% than the control one of 65.73%. The quality of egg in the experimental lodging was also significantly improved in the damaged egg ratio, the egg weight and the egg yolk weight after using probiotic Elac-grow. The efficiency of using probiotic Elac-grow and Han-proway was also shown through the concentration of ammonia

 $1.22 \pm 0.22 \times 10^7$ 

 $1.32 \pm 0.27 \times 10^7$ 

 $1.25 \pm 0.23 \times 10^7$ 

 $1.05 \pm 0.17 \times 10^7$ 

emission in the animal lodging. The ammonia value of the test lodging was maximum 5 ppm and remained stable at this point, which was lower than the control. This efficiency was shown significantly in the microbial fluctuation analysis.

**Acknowledgements:** *This study was supported by Hanvet fund for research.* 

## REFERENCES

AFRC, R FULLER (1989) Probiotics in Man and Animals. *J Appl Bacteriol* 66(5): 365–78. https://doi.org/10.1111/jam.1989.66.issue-5.

Applegate TJ, V Klose, T Steiner, A Ganner, G Schatzmayr (2010) Probiotics and Phytogenics for Poultry: Myth or Reality? *J Appl Poult Res* 19(2): 194–210. https://doi.org/10.3382/japr.2010-00168.

Balevi T, USU An, B Coskun, V Kurtoglu, IS Etingül (2001) Effect of Dietary Probiotic on Performance and Humoral Immune Response. *Brit Poult Sci* 42 (4): 456–61. https://doi.org/10.1080/ 00071660120073133.

Dhama K, Vinay Verma, P Sawant, Ruchi Tiwari, R Vaid, R Chauhan (2011) Applications of Probiotics in Poultry: Enhancing Immunity and Beneficial Effects on Production Performances and Health - A Review. *J Immunol Immunopathol* 13(1): 1–19.

Jin LZ, YW Ho, N Abdullah, S Jalaludin (1997) Probiotics in Poultry: Modes of Action. *World's Poult Sci J* 53(4): 362–68.

https://doi.org/10.1079/wps19970028.

Kurtoglu V, F Kurtoglu, E Seker, B Coskun, T Balevi, ES Polat (2004) Effect of Probiotic Supplementation on Laying Hen Diets on Yield Performance and Serum and Egg Yolk Cholesterol. *Food Addit Contam* 21(9): 817–23.

https://doi.org/10.1080/02652030310001639530.

Miles DM, SL Branton, BD Lott (2004) Atmospheric Ammonia Is Detrimental to the Performance of Modern Commercial Broilers. *Poult Sci* 83(10): 1650–54. https://doi.org/10.1093/ps/83.10.1650.

Nahashon SN, HS Nakaue, SP Snyder, LW Mirosh (1994) Performance of Single Comb White Leghorn Layers Fed Corn-Soybean Meal and Barley-Corn-Soybean Meal Diets Supplemented with a Direct-Fed Microbial. *Poult Sci* 73(11): 1712–23. https://doi.org/10.3382/ps.0731712.

Nahashon, Samuel N, Harry S Nakaue, Larry W Mirosh (1996a) Nutrient Retention and Production Parameters of Single Comb White Leghorn Layers Fed Diets with Varying Crude Protein Levels and Supplemented with Direct-Fed Microbials. *Anim Feed Sci Technol* 61(1–4): 17–26. https://doi.org/10.1016/0377-8401(96)00956-X.

Nahashon, Samuel N, Harry S Nakaue, Larry W Mirosh (1996b) Performance of Single Comb White Leghorn Fed a Diet Supplemented with a Live Microbial during the Growth and Egg Laying Phases. *Anim Feed Sci Technol* 57(1–2): 25–38. https://doi.org/10.1016/0377-8401(95)00852-7.

Park JW, JS Jeong, SI Lee, IH Kim (2016) Management and Production: Effect of Dietary Supplementation with a Probiotic (Enterococcus Faecium) on Production Performance, Excreta Microflora, Ammonia Emission, and Nutrient Utilization in ISA Brown Laying Hens. *Poult Sci* 95(12): 2829–35. https://doi.org/10.3382/ps/pew241.

Patterson JA, KM Burkholder (2003) Application of Prebiotics and Probiotics in Poultry Production." *Poult Sci* 82(4): 627–31. https://doi.org/10.1093/ps/82.4.627.

QCVN01-15: 2010/BNNPTNT: National Technical Regulation Conditions for biosecurity of poultry farms compiled by the Department of Livestock Production, submitted by the Department of Science, Technology and Environment and issued according to the Circular No. 04/2010/TT-BNNPTNT signed on January 15, 2010 by the Ministry of Agriculture and Rural Development ( in Vietnamese).

Song ZT, XF Dong JM Tong, ZH Wang (2012) Effects of Waste Vinegar Residue on Nutrient Digestibility and Nitrogen Balance in Laying Hens. *Livestock* Sci 150(1–3): 67–73. https://doi.org/10.1016/j.livsci.2012.08.004.

Tahseen Aziz, Rollins Animal Disease Diagnostic Laboratory, North Carolina Department of Agriculture and Consumer Services, and Dr. H. John Barnes, College of Veterinary Medicine, NC State University, Raleigh, NC, USA (2010). https://www.poultryworld.net/Breeders/Health/2010/ 10/Harmful-effects-of-ammonia-on-birds-WP008071W

Tortuero F, E Fernández (1995) Effects of Inclusion

of Microbial Cultures in Barley-Based Diets Fed to Laying Hens. *Anim Feed Sci Technol* 53(3–4): 255– 65. https://doi.org/10.1016/0377-8401(94)00747-W. Wathes CM, HH Kristensen (2000) Ammonia and Poultry Welfare : A Review. *World's Poult Sci J* 56(September): 236–45. Zhang, Yaowen, Wenfeng Ma, Zhidan Zhang, Fangyuan Liu, Jie Wang, Yulong Yin, and Zhanbin Wang (2019) Effects of Enterococcus Faecalis on Egg Production, Egg Quality and Caecal Microbiota of Hens during the Late Laying Period. *Arch Anim Nutr* 73(3): 208–21. https://doi.org/10.1080/1745039X.2019.1591128.