INSIGHT INTO *PAPHIOPEDILUM X DALATENSE* AVER. (ORCHIDACEAE) ORIGIN BASED ON MORPHOLOGICAL AND MOLECULAR MARKERS

Tran Thai Vinh¹, H' Yon Nie Bing¹, Vu Kim Cong¹, Dang Thi Tham¹, Ngo Phuong Linh², Le Ngoc Trieu², Nong Van Duy^{1, ⊠}

¹*Tay Nguyen Institute of Scientific Research, Vietnam Academy of Science and Technology, 116 Xo Viet Nghe Tinh Road, Da Lat City, Lamdong Province, Vietnam* ²*Da Lat University, 1 Phu Dong Thien Vuong Street, Da Lat City, Lamdong Province, Vietnam*

To whom correspondence should be addressed. E-mail: duynongvan@yahoo.com

Received: 04.11.2021 Accepted: 24.01.2022

SUMMARY

Paphiopedilum callosum, Paphiopedilum villosum and Paphiopedilum \times dalatense are endangered species and are currently listed as Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES, 2019) Appendix I species. Thus, the international trade for these naturally occurring species is forbidden. However, the *in vitro* propagated materials and their inter-species hybrids forced by breeders can be traded for commercial purposes. $P \times dalatense$ is an endemic slipper orchid species of Vietnam with high aesthetic value. Currently, the natural reserves of this species have been greatly reduced by overexploitation. In this study, P. × dalatense was demonstrated to be the reversible hybrid species between P. callosum and P. villosum based on both morphological and molecular markers. P. × dalatense possesses intermediate morphological characteristics between P. callosum and P. villosum which could be recorded in leaf, in flower and its components such as lip, petal, and dorsal sepal. There were 23 sites in ITS1-5.8S rRNA-ITS2 region sequence of P. × dalatense which possess the nucleotide characteristic of both P. callosum and P. villosum. There were two out of three $P_{\cdot} \times dalatense$ investigated samples possessed partial matK gene which was identical to the corresponding sequence in P. villosum and the last sample possessed identical partial matK gene to P. callosum. Proving P. × dalatense to be the reversible hybrid species between P. callosum and P. villosum is the essentially scientific base for commercial breeding of P. x dalatense by artificial hybridization which contributes to avoid overexploiting the natural $P_{\rm e} \times$ dalatense resources.

Keyword: ITS1-5.8S rRNA-ITS2, hybrid species, *Paphiopedilum* \times *dalatense*, partial *mat*K gene, reversible

INTRODUCTION

Paphiopedilum genus includes about 75 species which distribute in tropical Asia, from South India and East Himalaya to Philippines, New Guinea and the Solomon Islands. There are about 22 to 26 naturally original and hybrid Paphiopedilum species distributed in Vietnam. With flower's special structure - the lip's shape looks like the shoe, slipper orchid species and their hybrids have been infatuated, collected and cultivated in many countries around the world.

Natural slipper orchid species usually distribute in the high mountain regions with the altitude above sea level of 800 - 1500 m, however there are several species distribute the lower altitude such as *P. concolor*. Slipper orchids have

been found in cool, shady, high humidity habitats, i.e. temperature from 18 to 24°C, light intensity from 30 to 40% sunlight, humidity about 80% is the suitable condition for growth and development of *Paphiopedilum* species. Almost slipper orchid species naturally grow in the craggy terrains such as upright cliffs in mountainous regions or ledges near waterfalls or under the canopies of tall trees (Averyanov *et al.*, 2003).

Paphiopedilum callosum (Rchb. f.) Stein has Vietnamese name is "Van hai" meaning veined slipper orchid and characterized by clear vein in the dorsal sepal. This is a natural slipper orchid species which has scattered distribution in different areas in Thailand, Cambodia, Lao and Vietnam. In Vietnam P. callosum naturally grows in evergreen, dense primary forest with broadleaf trees, seasonal rain climate. This species usually grows on leaf humus belonging to grass layer in the forest like Cymbidium species. However, occasionally this species can be found on rocks covered with moss (Averyanov et al., 2003). Nowadays, it's extremely rare to meet P. callosum in nature in Vietnam because this species grows in the low altitude places, easy to be exploited resulting in a depletion of the species reserves.

Paphiopedilum villosum (Lindl.) Stein has a Vietnamese name is "Hai vang" meaning yellow slipper orchid and characterized by large and attractive flowers. This species has a wide and varied distribution from Eastern North India, North Myanmar, Western South Yunnan to North Thailand. This species is relatively popular in the locality of Da Lat, especially in Lang Bian mountain and is the only slipper orchid species possesses mainly epiphyte, it is rarely found in the rocks, but at the inaccessible rock tops of steep cliffs in gorges along alpine streams (Averyanov *et al.*, 2003). Currently, *P. villosum* is recorded in the Red list of the plant in Vietnam as EN B1+ 2b, c, e (Ban, 2007).

 $Paphiopedilum \times dalatense$ Aver. was described in 2001 by Averyanov and this author himself considered this species was the hybrid

between *P. villosum* and *P. callosum* due to *P.* \times dalatense possesses the intermediate characteristics in morphology between the two hypothetically parental species. This species is characterized by a relatively large flower with an olive-green vein in dorsal sepal and small blackpurple flat warts along upper margin toward the base of petals, these make P. × *dalatense* its own unique beauty. In nature, $P. \times dalatense$ could be found at evergreen primary forests in Don Duong district, Lamdong province (Averyanov, 2001). Nowadays, this species become rare to meet in nature due to overexploitation and the narrowing of the habitats. Demonstration P. × dalatense was the hybrid species between P. villosum and P. callosum based on morphological and molecular proofs will support to commercial Р. dalatense by artificial breeding х hybridization. However, up to the present time, there has been no in-depth study to determine the genetic relationship between the three species of P. callosum, P. villosum and P. × dalatense found although the artificial hybrids between still possess similar morphological characteristics to $P_{\cdot} \times dalatense$.

In this study, the intermediate characteristics in morphology between *P. callosum* and *P. villosum* expressed in *P. × dalatense* are shown together with the comparison DNA data of ITS1-5.8S rRNA-ITS2 nuclear region and chloroplast partial *mat*K of three investigated species to make an insight into the origin of *P. × dalatense*.

MATERIALS AND METHODS

Materials

Materials for morphological investigation are all of adult individuals of three studied species (*P. callosum*, *P. villosum* and *P.* × *dalatense*) which were collected from their natural habitats in Don Duong district, Lam Dong province, Vietnam from 1999 to 2019 and have been cultivated to keep and to conserve in Tay Nguyen Institute of Scientific Research. These individuals were previously classified and identified by Nong Van Duy. In addition, the dry specimens in VTN botanical museum also used in the research.

In study on molecular characteristics, leaf of three alive above adult individuals per each studied species were sampled for analysis. Samples belong to *P. callosum* were coded as A1-A3, *P. villosum* as B1-B3, and *P. × dalatense* as C1-C3, respectively.

Recording and comparing morphological characteristics

The throughout method in the process of recording morphological characteristics of surveyed species and determining the intermediate characteristics between *P. villosum* and *P. callosum* species expressed in *P.* × *dalatense* is the comparative morphological method in accordance with described by Thin (2007), Klein and Klein (1970).

The differences among three studied species were investigated in vegetative organs, i.e. quantity, shape, size, color of leaves and reproductive organs, i.e. shape, size, distribution of basal color and vein color and warts of flower and its components such as dorsal sepal, petal, lip. Based on that, indicating which morphological characteristics in $P. \times$ dalatense were the intermediate characteristics between P.*villosum* and P. callosum.

Identification hybrid species based on partial DNA sequence of ITS1-5.8S rRNA-ITS2 and *mat*K gene

DNA extraction

Total genomic DNA was extracted using CTAB protocol I (Weising *et al.*, 2005) with a modification of adding 10% SDS to the extraction buffer, which was then dissolved in water for subsequent use.

DNA isolation and amplification by PCR

PCRs were performed in 50 μ L reactions containing 25 μ L My Red HS Taq mix (Bioline), 2.5 μ L 10 pmol/ μ L forward primer, 2.5 μ L 10 pmol/ μ L reverse primer and approximately 50 ng

DNA templates. The reactions were performed in a Mastercycler \mathbb{R} nexus thermocycler (Eppendorf, Germany) with the following thermal programs: (i) For ITS1-5.8S rRNA-ITS2 region: initial denaturation at 94°C for 7 min, 35 cycles of 94°C for 30 s, 51°C for 1 min, 72°C for 1 min 30 s; final extension at 72°C for 10 min; (ii) For partial *mat*K gene: initial denaturation at 94°C for 5 min; 36 cycles of 94°C for 45 s, 51°C for 45 s, 72°C for 1 min 30 s; final extension at 72°C for 15 min.

Forward and reverse primers for ITS1-5.8S rRNA-ITS2 region isolation and amplification were ITS 5P (5'- GGA AGG AGA AGT CGT AAC AAG G -3') and ITS 8P (5'- CAC GCT TCT CCA GAC TAC A -3'), respectively. This primer pair was used in the previous study by Moller and Cronk (1997). Forward and reverse primers for partial *mat*K gene isolation and amplification were *mat*K F (5'- CGA TCT ATT CAT TCA ATA TTT C -3') and *mat*K R (5'- GTT CTA GCA CAA GAA AGT CG -3'), respectively. This primer pair was used in the previous study by Zuo *et al.* (2011)

Electrophoresis, purification and sequencing

The PCR products were detected by electrophoresis on 1.0 % agarose gel electrophoresis and then were purified using AccuPrep ® Gel purification Kits (Bioneer, Korea). DNA sequencing was performed using ABI 3730 sequencers (Phu Sa Biochem Ltd., Vietnam)

DNA sequence data analysis

DNA sequence data were processed by Bioedit 5.6.0 (Hall, 1999) and Chromas 2.6.6 (Goodstadt, Ponting, 2001). Mega 6 software was used to align the corresponding sequences (Tamura *et al.*, 2013).

From the scientific basis of morphological and molecular data, identify P. × *dalatense* is a hybrid between *P. villosum* and *P. callosum*, and indicate which species plays the maternal or paternal role (if any).

RESULTS AND DISCUSSION

The origin of P. × *dalatense* based on morphological characteristics

The morphological data recorded for three investigated species (*P. callosum*, *P. villosum* and *P.* × *dalatense*) in this study were not much different from previous studies (Averyanov, 2001; Averyanov *et al.*, 2003), but also indicated detail characteristics which were not clearly described by previous authors. These characteristics reflected the intermediate nature between *P. callosum* and *P. villosum* in *P.* × *dalatense* (Figure 1.a), as follows:

In leaf morphology, P. callosum were narrowly elliptic, oblong-elliptic or obovate, tridenticulate at the acute apex, ciliate at the base. tessellated pale and dark above, sometimes purple at a base on the lower surface (Figure 1.b1). In P. villosum had leaves linear ligulate, acuminate to acute at unequally bilobate apex mid-green on the undersurface, purple-spotted at the base, basal margins ciliate (Figure 1.b2). Meanwhile, leaves of P. \times dalatense were narrowly or oblong-elliptic acute at apex, green/pale-green finely but indistinctly tessellated above, with fine purple-violet speckled toward the base on the lower surface (Figure 1.b3).

Dorsal sepals of *P. callosum* were white flushed with purple in the lower half, veined with purple and green (Figure 1.d1). *P. villosum* had dorsal sepal green with a white margin and central glossy deep maroon areas (Figure 1.d2). *P.* × *dalatense* had dorsal sepal white at the base with lower half flushed pink, pinkbrown, purple-brown, purple, or olive-green, veined with olive-green, brown or purple (Figure 1.d3).

Petals of *P. callosum* was white to yellowgreen with a purple apical third spotted with maroon on upper margin and sometimes in basal half. Petals sometimes reflexed, sub-sigmoid, ligulate, obtuse or rounded at apex, maroonciliate (Figure 1.e1). *P. villosum* has petals glossy reddish-brown with a central maroon stripe. Petals incurved, obovate-spathulate, retuse-emarginate, rounded at apex, glossy, ciliate, purple-villose at the base (Figure 1.e2). P. \times dalatense petals were white to a dull pink, yellowish-green and pale brownish-green with pink, brownish-purple or purple stripes, sometimes few spotted with small black-purple flat warts along upper margin toward the base. Petals reflexed, sub-sigmoidal, ligulate, obtuse or broad apiculate at apex, with black-purple cilia (Figure 1.e3).

About the lips, in *P. callosum*, the lip had green, heavily flushed deep maroon and warty on incurved side-lobes (Figure 1.f1). In *P. villosum* the lip had lip have ochre, flushed with pink or reddish and lip tapering to apex (Figure 1.f2). *P.* \times *dalatense* had lip pink, yellowish-pink or light yellowish-brown with more deep coloured veins (Figure 1.f3).

Above data showed that $P. \times$ dalatense possessed morphological characteristics which could be considered as intermediaries between P. *villosum* and P. *callosum* in both vegetative and reproductive organs. This is the morphological basis to indicate that $P. \times$ dalatense is a hybrid between the other two species.

The origin of $P. \times$ dalatense based on molecular data

The origin of P. × dalatense based on ITS1-5.8S rRNA-ITS2 region sequence

The length of ITS1-5.8S rRNA-ITS2 region sequence in P. callosum was 764 bp, in P. villosum and P. \times dalatense was 761 bp. Result of careful check the sequences and chromatograms of ITS1-5.8S rRNA-ITS2 region showed that the sequences of same investigated sample induced by sequencing with forward and reverse primers were highly matched; the sequences of the samples belonging to the same species were identical. Comparing ITS1 region among three investigated species indicated that there were 12 nucleotide sites in P. × dalatense having the specific nucleotides of both P. callosum and P. villosum (Table 1).



Vietnam Journal of Biotechnology **20**(2): 279-287, 2022

Figure 1. (a) Morphological; (b) Leaf characteristic; (c) Flower characteristic; (d) Dorsal sepal characteristic; (e) Petal characteristic; (f) Lip characteristic; (1) *P. callosum*; (2) *P. villosum* and (3) *P. × dalatense.*

Site Species	86	117	121	129	165	172	173	184	215	227	236	242	243	248	272
P. callosum	т	G	А	т	т	G	А	т	С	Т	G	С	С	С	т
P. villosum	А	А	С	G	С	-	-	А	А	С	С	G	G	т	-
P. × dalatense	W	R	М	к	Y	-	-	W	М	Y	S	S	S	Y	-

Table1. Variation of ITS1 sequence among three investigated species.

Y: C and T; K: G and T; S: C and G; R: A and G; W: A and T; M: C and A.

 Table 2. Variation of 5.8S rRNA sequence among three investigated species.

Site	453	465	510	536	547	
P. callosum	С	Т	Т	А	А	
P. villosum	т	С	А	G	Т	
P. × dalatense	Y	Y	W	R	W	

Y: C and T; R: A and G; W: A and T.

Table 3. Variation of ITS2 sequence among three investigated species.

Site Species	594	595	603	606	635	637	
P. callosum	Т	С	А	G	С	G	
P. villosum	С	Т	G	Т	G	А	
P. × dalatense	Y	Y	R	К	S	R	

Y: C and T; K: G and T; S: C and G; R: A and G

Comparing 5.8S rRNA region among three investigated species indicated that there were 5 nucleotide sites in P. × *dalatense* having the specific nucleotides of both *P. callosum* and *P. villosum* (Table 2).

Comparing ITS2 region among three investigated species indicated that there were 6 nucleotide sites in P. × *dalatense* having the specific nucleotides of both *P. callosum* and *P. villosum* (Table 3).

Thus, in the ITS1-5.8S rRNA-ITS2 region, P. × dalatense possessed 23 nucleotide sites which were specific for both P. callosum and P. villosum. This phenomena was shown clearly in

the chromatograms and indicated that $P. \times dalatense$ received one sister chromatid from P. villosum and another from P. callosum in certain dyad. Based on sequence data of the ITS1-5.8S rRNA-ITS2, it is recognized that $P. \times dalatense$ is the naturally hybrid species between P. callosum and P. villosum.

Studying on the hybrid orchid species in Guatemala, Szlachetko *et al.* (2017) used to investigate ITS1-5.8S rRNA-ITS2 region and gene *XDH* in several *Cypripedium* species including *C. dickinsonianum, C. irapeanum, C. molle* and *C.* × *fred-mulleri* and indicated *C.* × *fred-mulleri* is the naturally hybrid species

Vietnam Journal of Biotechnology 20(2): 279-287, 2022

between *C. dickinsonianum* và *C. irapeanum* based on *C.* × *fred-mulleri* possesses 4 nucleotide sites in ITS1-5.8S rRNA-ITS2 region and 7 nucleotide sites in *XDH* gene which are specific for both *C. dickinsonianum* and *C. irapeanum*.

Comparing to the research of Szlachetko *et al.* (2017), in the current research the number of nucleotide sites specific for both parent species was significantly higher (23 sites comparing to 5 sites).



Figure 2. Double peaks at 594, 595, 603, 606 sites in the chromatograms of ITS2 sequence in *P. × dalatense* (C) possesses the specific nucleotide of both *P. callosum* (A) and *P. villosum* (B) and result of corresponding alignment.

The origin of P. × dalatense based on partial matK gene sequence

The length of partial *mat*K gene sequence were 854 bp for all of investigated species, from 184 site to 1107 site in whole *mat*K gene. The sequences of same investigated sample induced by sequencing with forward and reverse primers were highly matched; the sequences of the samples belonging to the same species were identical in *P. callosum* and *P. villosum* but the samples belonging to *P.* \times *dalatense* were different.

Table 4. Variation of partial *mat*K gene among three investigated species.

Site on the investigated partial matK gene	184	636	702	805
Site on the matK gene	437	889	955	1058
P. callosum, 3 investigated samples	G	А	G	А
P. villosum, 3 investigated samples	т	С	Т	G
<i>P. × dalatense,</i> sample C1	Т	С	Т	G
P. × dalatense, sample C2	т	С	Т	G
P. × dalatense, sample C3	G	А	G	А

Based on the partial *mat*K gene, the variation among three investigated was shown in Table 4.

Table 4 indicated that C1 and C2 P. × dalatense samples possessed identical partial matK gene to P. villosum samples but C3 P. × dalatense sample possessed identical partial matK gene to P. callosum samples. Accordingly, partial matK gene in P. × dalatense could be identical to any of in P. callosum and P. villosum.

In most flowering plants, the hybrid between two parent species or the next generation of the reproductive species possess the plastid genome inherited from maternal species or maternal individuals respectively (Corriveau, Coleman, 1988). In that study, these authors investigated more than 200 angiosperm species belong to various plant families and indicated that about fourth of total investigated species one possessing plastid genome is biparentally inherited, mainly in families such as Strelitziaceae, Plumbaginaceae, Passifloraceae, Onagraceae. Geraniaceae. In all of representatives of Orchidaceae, including the taxa belong to Paphiopedilum genus in the study of Corriveau and Coleman (1988), the plastid DNA is maternally inherited.

The genus of *Paphiopedilum* also belongs to the most maternally inherited plastid DNA angiosperm species and this is confirmed again in the study of Chochai *et al.* (2012).

Because the *mat*K gene is located on the chloroplast, its sequence in the hybrid species is identical to species playing maternal role in *Paphiopedilum* species. In case of *P. × dalatense* in the current study, partial *mat*K gene in *P. × dalatense* could be identical to any of in two parental species. Thus, *P. × dalatense* can be proved to be the reversible hybrid species between *P. villosum* and *P. callosum*.

Szlachetko *et al.* (2017) also used to investigate 3'trnK-matK sequence of *Cypripedium dickinsonianum, C. irapeanum, C.* molle, C. × fred-mulleri and indicated C. × fredmulleri is the naturally hybrid species between C. dickinsonianum (paternal species) and C. *irapeanum* (maternal species) because $C. \times fred-mulleri$ shares the identical 3'*trnK-matK* sequence with *C. irapeanum* while *C. dickinsonianum* differs to *C. irapeanum* and *C. × fred-mulleri* in four sites. It was different from the study of Szlachetko *et al.* (2017), in the current study, *P. × dalatense* was proved to be the reversible hybrid species.

Together with the information that the samples were collected from Don Duong district, Lam Dong province, where the three investigated species shared the same natural distribution, the morphological and molecular data achieved in this research proved that $P. \times dalatense$ is the reversible hybrid species between P. callosum and P. villosum.

CONCLUSION

morphological characteristics of In vegetative and reproductive organs, P. Х dalatense species possessed the intermediate characteristic between P. callosum and P. villosum. Based on DNA sequence data of nuclear ITS1-5.8S rRNA-ITS2 region, P. × dalatense possessed 23 nucleotide sites which are specific for both P. villosum and P. callosum. Based on DNA sequence data of plastid partial matK gene, two out of three investigated P. \times dalatense samples were identical to P. villosum and the last sample was identical to P. callosum. All of these findings on morphological and molecular characteristics proved that $P_{\text{c}} \times$ dalatense is the reversible hybrid species between P. callosum and P. villosum.

Acknowledgements: The authors gratefully acknowledge Tay Nguyen Institute for Scientific Research, Vietnam Academy of Science and Technology and Tay Nguyen Program 2016 -2020, project TN18/T08 for their support in completing this study.

REFERENCES

Averyanov L, Cribb PJ, Loc PK, Hiep NT (2003) Slipper orchid of Vietnam. With an Introduction to the

Flora of Vietnam. Royal Botanic Gardens, Kew; UK ed. Edition.

Averyanov L (2001) New natural interspecific hybrid - *Paphiopedilum × dalatense* from Vietnam. *Orchid Digest* 65(3): 133-134.

Chochai A, Leitch IJ, Ingrouille MJ, Fay MF (2012) Molecular phylogenetics of *Paphiopedilum* (Cypripedioideae; Orchidaceae) based on nuclear ribosomal ITS and plastid sequences. *Bot J Linn Soc* 170: 176-196.

Corriveau JL, Coleman AW (1988) Rapid screening method to detect potential biparental inheritance of plastid DNA and results for over 200 angiosperm species. *Amer J. Bot.* 75(10): 1443-1458.

Doyle JJ, Doyle JL (1987) A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochem Bull* 19: 11-15.

Goodstadt L, Ponting CP (2001) CHROMA: consensus-based colouring of multiple lignments for publication. *Bioinformatics* 17(9): 845-46.

Hall TA (1999) *BioEdit: a user-friendly biological* sequence alignment editor and analysis program for Windows 95/98/NT. Paper presented at the Nucl. Acids Sympos. Ser.

Klein RM, Klein DT (1970) *Research Methods in Plant science*. Garden City, Newyork. Natural History Press.

Moller M, Cronk QC (1997) Origin and relationships of *Saintpaulia* (Gesneriaceae) based on ribosomal DNA internal transcribed spacer (ITS) sequences. *Amer J. Bot.* 84(7): 956-965.

Thin NN (2007) *Methods for plant research*. Hanoi National University Publisher.

Ban NT (2007) *Vietnam Red Book - Plant*. Natural Science and Technology Publishing.

Sanger F, Nicklen S, Coulson AR (1977) DNA sequencing with chain - terminating inhibitors. *Proc Nalt Acad Sci* 74(12): 5463-5467.

Szlachetko DL, Kolanowska M, Muller F, Vannini J, Rojek J, Górniak M (2017) First Guatemalan record of natural hybridisation between Neotropical species of the Lady's Slipper orchid (Orchidaceae, Cypripedioideae). *PeerJ* 5: e4162.

Tamura K, Stecher G, Peterson D, Filipski A, Kumar S (2013) MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol* 30(12): 2725-2729.

Weising K, Nybom H, Wolff K, Kahl G (2005) DNA fingerprinting in plants: principles, methods, and applications. CRC press.

Zuo Y, Chen Z, Kondo K, Funamoto T, Wen J, Zhou S (2011) DNA barcoding of Panax species. *Planta Med* 77(02): 182-187.