

# Synthesis of hydroxyapatite and strontium-substituted hydroxyapatite for bone replacement and osteoporosis treatment

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**Abstract.** Hydroxyapatite (HAp) is an inorganic component exhibiting bioactivity similar to that of natural bone. However, it is not resorbed by osteoclasts during bone remodeling due to its lack of bio-resorption property. This can be enhanced by the substitution of other element presented in bone mineral. In this research work, hydroxyapatite (HAp) and strontium-substituted hydroxyapatite (Sr-HAp) were synthesized by a precipitation method. Calcium nitrate tetra hydrate [Ca(NO<sub>3</sub>)<sub>2</sub>•4H<sub>2</sub>O], disodium hydrogen phosphate (Na<sub>2</sub>HPO<sub>4</sub>), and Strontium nitrate [Sr(NO<sub>3</sub>)<sub>2</sub>] were used as Ca, PO<sub>4</sub> and Sr sources, respectively. Molar ratio Ca/P=1.67 was used to synthesize HAp, where (Ca+Sr)/P=1.67 was used to synthesize strontium substituted-HAp (Sr-HAp). The reaction was carried out at room temperature. The results show that pure HAp and Sr-HAp were formed with nanometer-sized particles. Sr substitution in the HAp lattice results in an increase in both the lattice disorder and crystal aspect ratio. The results of *in vitro* bioactive testing using simulated bodily fluid also showed that both HAp and Sr-HAp have high bioactive, with the Sr-HAp sample having the greater bioactive. Therefore, HAp and Sr-HAp have great potential for biological applications.

**Keywords:** Biomedical application, Hydroxyapatite, Strontium, Osteoporosis treatment.

**Classification numbers:** 5.2.1.

## 1. INTRODUCTION

Hydroxyapatite [Ca<sub>10</sub>(PO<sub>4</sub>)<sub>6</sub>(OH)<sub>2</sub>, HAp] has been widely studied and applied in orthopedic and dental applications due to its chemical similarity to the inorganic composition of human bone. The synthesized HAp exhibits various physical and biological characteristics, which depend on factors such as shape, microstructure, and crystallinity. However, the biodegradability of HAp is limited, as it remains in the body for years. This situation can lead to issues like infections or alterations in the mechanical properties of the implanted bone [1, 2].

Furthermore, HAp is not resorbed by osteoclasts to participate in bone remodeling process, unlike bone minerals [3, 4]. Consequently, the utility of HAp in biomedical applications is limited. The bioresorption of HAp can be improved by substituting trace elements i.e. Si, CO<sub>3</sub>, Mg, Na, Sr, etc. that are present in the human bone composition [5 - 7]. These ions substitutions reduce the crystallite size of the HAp and facilitate new bone formation [7 - 9].

Additionally, bone defects due to osteoporosis in the elderly as the aging population is increasing are common issues in orthopedic surgery. Research has demonstrated that strontium (Sr) in bone promotes bone regeneration, significantly reduces bone resorption, and can treat osteoporosis [8, 10, 11]. Therefore, the development of biomaterials with chemical compositions resembling those of human bones and containing elements that promote bone regeneration is essential for bone replacement.

In this study, hydroxyapatite (HAp) and strontium-substituted hydroxyapatite (Sr-HAp) was synthesized by a precipitation method using Calcium nitrate tetrahydrate [Ca(NO<sub>3</sub>)<sub>2</sub>•4H<sub>2</sub>O], disodium hydrogen phosphate (Na<sub>2</sub>HPO<sub>4</sub>), and Strontium nitrate [Sr(NO<sub>3</sub>)<sub>2</sub>] as precursors. The reaction was carried out at room temperature. The resulting products were characterized using x-ray diffraction (XRD), scanning electron microscopy (SEM), Energy Dispersive X-Ray Spectroscopy (EDX), Fourier transform infrared attenuated total reflection spectroscopy (ATR-FTIR). Furthermore, the bioactive of the synthesized HAp and Sr-HAp were assessed using the simulated body fluid method.

## **2. MATERIALS AND METHODS**

### **2.1. Materials**

Calcium nitrate tetrahydrate [Ca(NO<sub>3</sub>)<sub>2</sub>•4H<sub>2</sub>O, Xilong, China], Disodium hydrogen phosphate [Na<sub>2</sub>HPO<sub>4</sub>, Xilong, China], Strontium nitrate [Sr(NO<sub>3</sub>)<sub>2</sub>, Xilong, China], Sodium hydroxide [NaOH, Xilong, China] were used to synthesize HAp and Sr-HAp. The chemicals are used directly without purification.

### **2.2. Methods**

In order to synthesize HAp, a solution containing Ca<sup>2+</sup> was prepared by dissolving Ca(NO<sub>3</sub>)<sub>2</sub>•4H<sub>2</sub>O in distilled water at a concentration of 0.5 mol/L (referred to as calcium solution). Similarly, a solution containing PO<sub>4</sub><sup>3-</sup> was also prepared by dissolving Na<sub>2</sub>HPO<sub>4</sub>•12H<sub>2</sub>O in distilled water at a concentration of 0.3 mol/L (referred to as phosphate solution). The Ca/P molar ratio was 1.67, corresponding to stoichiometric HAp [12]. The phosphate solution was slowly added dropwise into the calcium solution, and the reaction was homogenized through continuous stirring using a magnetic stirrer (HS4000-S-E, Benchmark, Korea) at 300 rpm for 2 hours. The solution's pH was controlled at pH = 9-10 by adding 2 mol/L NaOH solution to the reaction mixture.

Similarly, for the synthesis of Sr-HAP, an appropriate amount of calcium and phosphate solutions was also prepared to achieve the molar ration of (Ca+Sr)/P = 1.67. The amount of Sr ion was fixed at 5 mmol, assuming that Sr would replace Ca ions in the HAp lattice. Once the reaction was complete, the precipitates were retained in the mother solution and aged at room temperature for 12 hours. The HAp and Sr-HAP precipitates were washed and filtered with distilled water multiple times. The resulting powder was dried at 70 °C for 12 hours and then ground to obtain a fine powder.

The synthesized powders were characterized using an X-ray diffractometer (XRD, AERIS, Netherland) for phase identifications. The molecular structure was investigated using Total attenuation Fourier Transform Infrared Spectrometer (Thermo Scientific - NICOLET iS50FT-IR). The microstructure of the synthesized powder was observed using a scanning electron microscope (SEM, JEOL JSM-IT200) equipped with an energy dispersive X-ray spectroscopy (EDS) under an accelerating voltage of 10 kV.

The crystallite size ( $d$ ) was calculated using the diffraction peak at approximately  $25.8^\circ$  ( $2\theta$ ) because it was sharper and isolated from others following the Scherrer equation (1) [13]:

$$d = \frac{K\lambda}{\beta \cos \theta} \quad (1)$$

where:  $d$  = crystallite size (Å);  $K$  = a shape factor equal to 0.9;  $\lambda$  = wavelength of Cu  $K\alpha$  radiation equal to 1.5406 Å;  $\theta$  = half of the diffraction angle, degree;  $\beta$  = full width at half maximum (FWHM), rad

The lattice parameters ( $a$  and  $c$ ) of the hexagonal structure for HAp and Sr-HAp of were also calculated using the ( $hkl$ ) plane of apatite from XRD patterns using the equations for hexagonal structure as follows (2) [13]:

$$\frac{1}{d^2} = \frac{4}{3} \left( \frac{h^2 + hk + k^2}{a^2} \right) + \frac{l^2}{c^2} \quad (2)$$

$d$  is the distance between the crystal planes obtained from XRD results at the (002) and (300) Miller plane family of the HAp and Sr-HAp.

The density of the samples was determined using a specific gravity (S.G.). The procedure is as follows: First, an empty 50 ml borosilicate bottle (S.G. bottle) was weighed to measure the dry weigh of the bottle ( $M_1$ ). Subsequently, approximately 1 g of HAp and Sr-HAp powders was placed into the S.G. bottle, and the combined weight of S.G. bottle and HAp/Sr-HAp was recorded ( $M_2$ ). Thirdly, deionized water was added to the S.G. bottle until the powders was fully immersed in the water. The entire S.G. bottle was then placed in a vacuum desiccator for 30 mins to eliminate any trapped air in the powder. The water level was adjusted to fill the S.G. bottle before sealing it with the lid. The S.G. bottle was wiped and weighed ( $M_3$ ). Subsequently, both the water and powder were removed from the S.G. bottle. Finally, the S.G. bottle was filled with deionized water, covered with the lid, and weighed again ( $M_4$ ).

The density of HAp/Sr-HAp powder was then calculated from the formula below:

$$\rho_{\text{powder}} = \frac{M_2 - M_1}{(M_4 - M_1) - (M_3 - M_2)} \times \rho_{\text{H}_2\text{O}}$$

where  $\rho_{\text{H}_2\text{O}}$  is taken as  $1.0 \text{ g/cm}^3$  at room temperature.

The bioactivity of the HAp and Sr-HAp samples was assessed by immersing them in simulated body fluid (SBF) for 7 days, following Kokubo's principle [14]. For this experiment, apatite pellets (10 mm diameter) were prepared by compacting 300 mg of HAp and Sr-HAp powders at room temperature through uniaxial pressing at 20 MPa, followed by heat treatment at  $500^\circ\text{C}$  (Lenton 1500, England) for 2 hours. This low temperature was selected to dense pellets without phase decomposition. The SBF solution was refreshed every 2 days. After immersion, the sample was taken out from the SBF and carefully rinsed with deionized water. Subsequently,

the sample was dried in a desiccator for 48 hours without applying heat. The formation of apatite on the surface of the sample was confirmed by scanning electron microscopy (SEM).

### 3. RESULTS AND DISCUSSION

Figure 1 shows the XRD patterns of various samples. The XRD patterns of the synthesized HAp and Sr-HAp perfectly matched those of the HAp standard. Moreover, the diffraction peaks of the Sr-HAp sample were less intense in comparison to those of the HAp sample. This may be due to the substitution of Sr into the HAp lattice, resulting in an increase in the crystallite size of HAp. The crystallite sizes of the samples were calculated using Scherrer equation [13]. The crystallite size of the HAp sample was 15.08 nm, while that of Sr-HAp was approximately 9.59 nm.

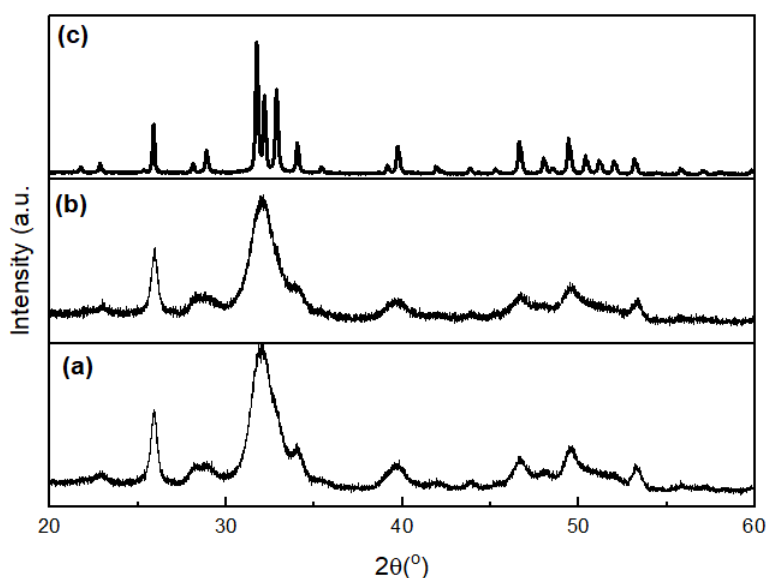


Figure 1. XRD patterns of different samples: (a) Hydroxyapatite (HAp), (b) Sr-substituted Hydroxyapatite (HAp), (c) Hydroxyapatite (HAp) standard (PDF No. 9-432).

The *a* and *c*-axes parameters for the synthesized HAp were 9.639 Å and 6.898 Å, respectively while those of Sr-HAp were 9.662 Å (*a*-axis) and 6.903 Å (*c*-axis). The substitution of Sr in the HAp structure led to an increase in the lattice parameters. This increase was attributed to the larger atomic radius of Sr ion (0.113 nm) in comparison to that of Ca ion (0.099 nm) [15]. These results were consistent with the previous studies indicating that both *a* and *c*-axes increased with Sr substitution [16, 17].

The XRD analysis was supported by the IR spectra. As shown in Fig 2, the IR spectra of HAp and Sr-HAp exhibited the characteristic features of HAp phase, in comparison to the spectrum of the HAp standard. Regardless of Sr substitution, the IR spectra of both samples displayed the typical bands of phosphate groups at approximately 561 cm<sup>-1</sup> ( $\nu_4$ ), 602 cm<sup>-1</sup> ( $\nu_4$ ), 9602 cm<sup>-1</sup> ( $\nu_1$ ) and 1027-1089 cm<sup>-1</sup> ( $\nu_3$ ) [16, 18, 19]. The hydroxide groups in the structure were detected at about 630 cm<sup>-1</sup> ( $\nu_{OH}$ ). It should be noted that, the substitution of Sr in the HAp results in a reduction in the intensity of OH groups, resulting in broader OH absorbance at approximately 630 cm<sup>-1</sup> compared to that of pure HAp. A similar result was observed in a

previous study [20]. Therefore, it can be concluded that Sr had substituted into the HAp lattice structure.

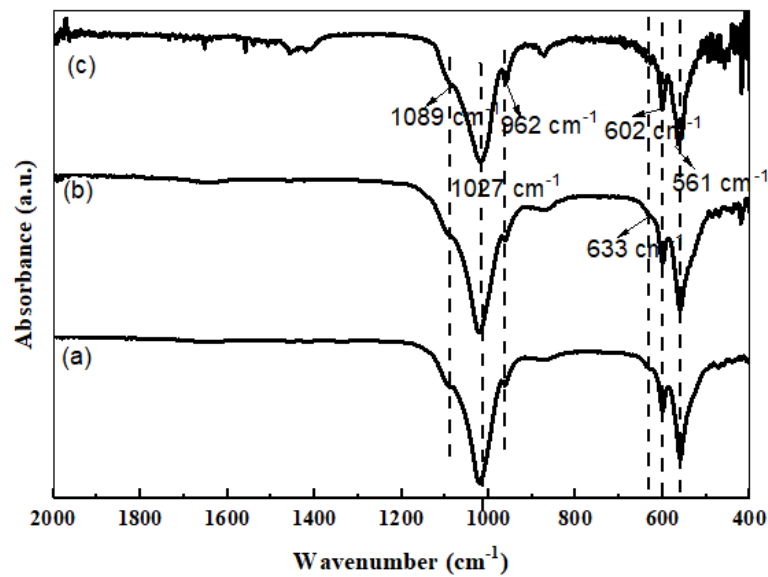


Figure 2. IR patterns of different samples: (a) Hydroxyapatite (HAp), (b) Sr-substituted Hydroxyapatite (HAp), (c) Hydroxyapatite (HAp) standard.

Figure 3 shows the SEM images of the synthesized HAp and Sr-HAp sample at various magnifications.

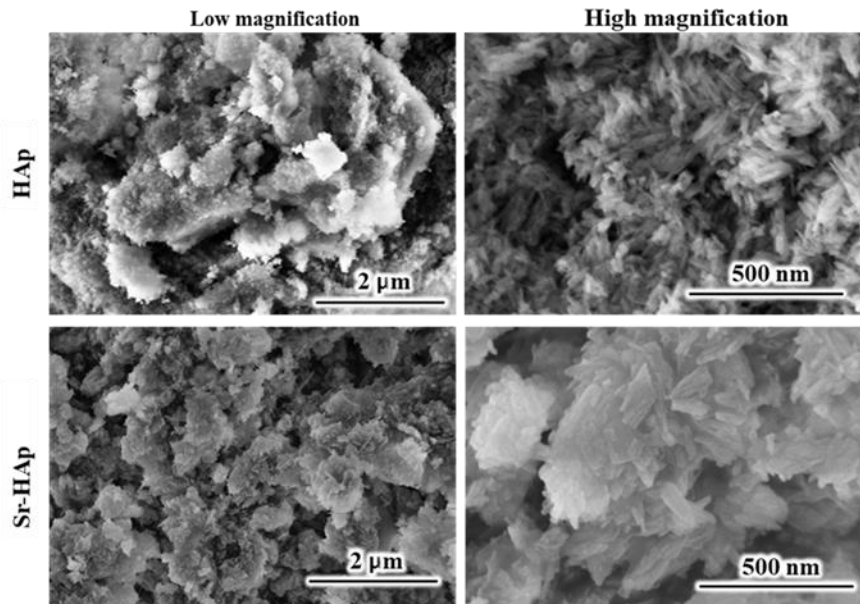


Figure 3. SEM images of different apatite samples at various magnifications

Low magnification SEM scans revealed that the synthesized powders consist of agglomerates or lumps. These agglomerated powders form due to the agglomeration of fine particles with high surface energy that tend to clump together. Both HAp and Sr-HAp crystals

exhibit a needle-like or rod-like morphology, as observed in the high magnification SEM images. The primary particles were observed to be nanosized crystals. The nanocrystals of HAp measured approximately 20 nm in width and about 100 nm in length, while those of Sr-HAp were about 22 nm in width and 126 nm in length. The aspect ratios (length/width) of HAp and Sr-HAp were 4.8 and 5.7, respectively. It has been reported that the higher aspect ratios could lead to improve biological properties of synthesized hydroxyapatite [21].

Figure 4 shows the EDS spectra and the quantities of the elements Ca, P, and Sr in different samples. These results indicate that the powder was composed of Ca, P, and Sr ions in the structure. It is worth noting that NaOH in the reactions and might contribute to sodium contamination in the powder.

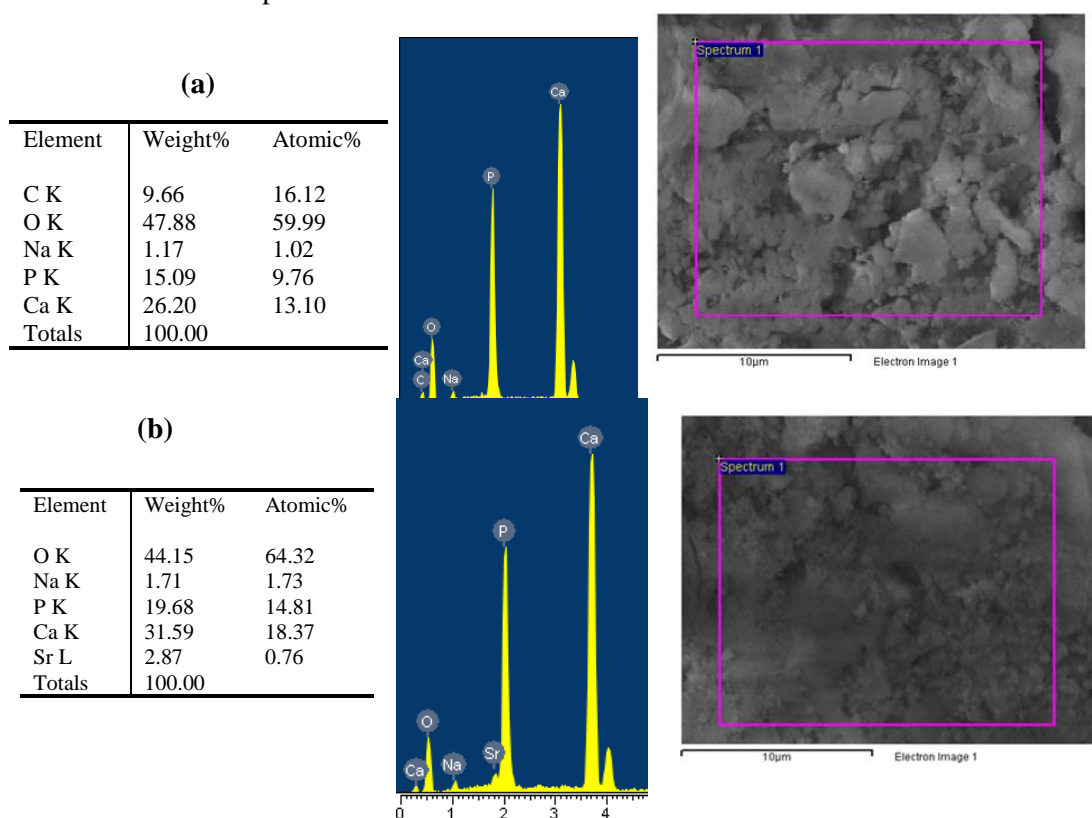


Figure 4. EDX results of different apatite samples: (a) HAp, (b) Sr-HAp.

As detected by the EDS, the amount of Sr substitution in the apatite structure was approximately 2.87 wt%. The substitution of strontium ions for calcium ions in HAp would decrease the calcium concentration in the apatite structure leading to a reduction in the Ca/P ratio [22, 23]. Therefore, the change in Ca/P ratios was attributed to the substitution of Sr in the HAp crystals.

It should be noted that the Sr amount, as well as Ca/P ratios as detected by the EDS are solely for screening purposes to indicate the presence of Sr in the apatite structure. Therefore, for a more accurate investigation, an appropriate characterization method, such as ICP-MS, could be considered.

The densities of HAp and Sr-HAp powders were also determined using the S.G. method. The substitution of Sr led to a reduction in the density of the HAp powder, with densities of HAp and Sr-HAp being  $1.9 \text{ g/cm}^3$  and  $1.6 \text{ g/cm}^3$ , respectively.

The bioactivity of HAp and Sr-HAp were evaluated through the simulated body fluid (SBF) method [24]. Figure 5 shows the surface morphology after immersion for 7 days. It's evident that cauliflower-like apatite layer was observed in both samples. Interestingly, the Sr-HAp sample exhibited a thicker apatite layer under the same immersion period. It is widely acknowledged that a majority of bioactive materials develop an apatite layer on their surfaces when suspended in SBF for a specific period. Consequently, the formation of an apatite layer on these samples confirms their bioactive nature, indicating that the synthetic surface establishes a chemical bond with the host bone after implantation,

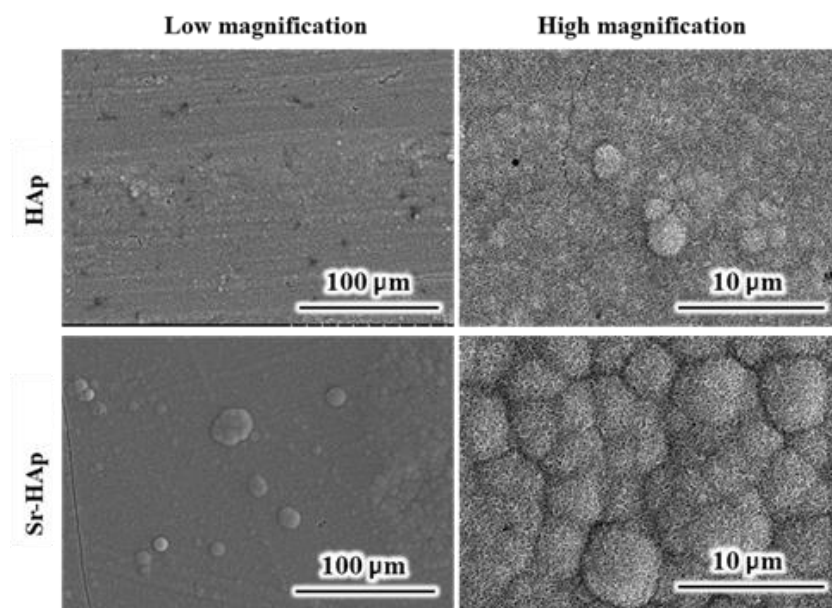


Figure 5. SEM images of the surface of HAp and Sr-HAp after immersion in SBF for 7 days

The formation mechanism of the apatite layer as a consequence of dissolution and precipitation process. Upon immersion, the surface partially dissolves, releasing  $\text{Ca}^{2+}$ ,  $\text{HPO}_4^{2-}$ ,  $\text{PO}_4^{3-}$ , and  $\text{Sr}^{2+}$  to the solution. As the concentration of these ions reaches the supersaturation level with respect to apatite, they precipitate, forming apatite nanocrystals on the surface of the material [24]. The thickening of the apatite layer could arise from accelerated dissolution and precipitation. The substitution of Sr into the HAp lattice is known to enhance the solubility of HAp [25]. The heightened solubility leads to a more rapid dissolution rate. As a result, Sr substitution enhanced the bioactive process, thereby potentially improving osteointegration *in vivo*.

#### 4. CONCLUSIONS

Hydroxyapatite (HAp) and strontium-substituted hydroxyapatite (Sr-HAp) powders were successfully synthesized using a precipitation method at room temperature. It was observed that the substitution of Sr in HAp influenced the structural order and increased the aspect ratios of the HAp crystals. The powders were in the nanosize range, approximately 20 nm in width, with

notable agglomeration. Furthermore, the Sr addition led to a decrease in the Ca/P molar ratio and a reduction in the density of the HAp. The *in vitro* bioactivity assessment using the SBF method demonstrated that the Sr-HAp exhibited superior bioactive property compared to pure HAp.

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**CRedit authorship contribution statement.** Le Thi Bang: Methodology, Investigation, Funding acquisition, First draft manuscript. Nguyen Van Ha: Methodology, Data analysis. Bui Duc Long: Formal analysis, Supervision. Nguyen Thi Hong Nhung: Data analysis.

**Declaration of competing interest.** The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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