Supporting information

Ecdysterols and triterpene glycoside from *Achyranthes aspera* L. and their NO production inhibitory activity

Hoang Thi Tuyet Lan¹, Bui Thi Mai Anh¹, Nguyen Thi Mai^{1,*}, Duong Thi Dung², Bui Huu Tai², Phan Van Kiem^{2,*}

Methods

Equipment

The CD spectra were recorded on a ChiraScan spectrometer. NMR spectra were measured on a Bruker Avance III 600 MHz (150 MHz for ¹³C NMR and 600 MHz for ¹H NMR). Column chromatography (CC) was performed using silica gel (40 - 63 μ m) or reversed phase C-18 (RP18, 150 μ m) as adsorbents. Thin layer chromatography was performed on pre-coated plates. Semi-preparative HPLC was performed on an Agilent 1260 infinity II system including binary pump, autosampler, DAD detector, fraction collector, and equipped with YMC J'sphere ODS-H80 (20 × 250 mm, 4 μ m) column. Mobile phase was an isocratic system of acetonitrile/water at flow rate of 3 mL/min.

Nitric oxide assay

Nitric oxide assay was performed as previously described [1, 2] In brief, RAW 264.7 cells were cultured in DMEM containing L-glutamine (2 mM), HEPES (10 mM), sodium pyruvate (1 mM), and fetal bovine serum (10 %). The cells (2×10^5 cells/well) were incubated in humidified atmosphere (95 % air and 5 % CO₂) at 37 °C. After 24h incubation, each well was added by compounds (0.4 - 100 µM) or vehicle and followed by LPS (1µg/mL) in the next 2h. The cells were then incubated for an additional 24h. After that, cell viability was then measured by MTT assay and amount of NO production in cell medium was determined by Griess reaction. Cultural medium (100 µL) was mixed with equal volume of Griess reagent and incubated in room temperature for 10 minutes. Absorbance was measured at 540 nm on a microplate reader. Nitrite concentration as an indicator of NO production was determined using a standard curve which was built by NaNO₂ serial diluted solutions. Experiments were performed in triplicate. IC₅₀ values were generated by TableCurve 2Dv4 software.

References

- [1] B. H. Tai, P. H. Yen, N. H. Hoang, P. T. T. Huong, N. V. Dung, B. V. Thanh, N. T. Cuong, N. A. Bang, N. X. Nhiem, P. V. Kiem, *RSC Adv.* 2022, *12*, 25433
- [2] S. Cheenpracha, E. J. Park, B. Rostama, J. M. Pezzuto, L. C. Chang, *Mar. Drugs* 2010, 8, 429–437.

Con. (µM)	1		2		3	
	Inhibition	Survival	Inhibition (%)	Survival	Inhibition	Survival
	(%)	cells (%)		cells (%)	(%)	cells (%)
100	13.58	87.04	59.76	84.12	64.79	91.29
20	8.95	88.82	42.31	88.78	47.04	96.40
4	4.94		29.59		32.54	
0.8	0.93		15.98		21.60	
IC ₅₀	>100	-	40.47 ± 4.90	-	27.21±1.23	-
Con. (µM)	4		5		6	
	Inhibition	Survival	Inhibition (%)	Survival	Inhibition	Survival
	(%)	cells (%)		cells (%)	(%)	cells (%)
100	66.10	87.67	60.36	93.16	7.10	95.75
20	43.15	91.44	46.75	95.71	2.78	96.17
4	27.40		35.80		1.23	
0.8	14.38		22.19		-1.85	
IC ₅₀	33.44±1.53	-	27.89±2.11	-	>100	-
Con.			L-NMMA*			
(µg/mL)			Inhibition (%)	Survival		
				cells (%)		
100			100.67	94.09		
20			79.36	98.54		
4			24.71			
0.8			10.03			
			7.99 ± 0.58	-		
IC ₅₀			(µg/mL)			
			$32.24 \pm 2.35 \ (\mu M)$)		

Table S1. NO production inhibition effects of compounds 1-6.

*positive control compound







Figure S5. ¹H-NMR spectrum of compound **2** in CD₃OD.



Figure S7. HSQC spectrum of compound 2.



Figure S9. ¹H-NMR spectrum of compound **3** in CD₃OD.



Figure S11. HSQC spectrum of compound 3.



Figure S12. HMBC spectrum of compound 3.



Figure S13. ¹H NMR spectrum of compound **4** in CD₃OD.



righte 515. TISQC spectrum of compound 4 in CD3





Figure S18. ¹H NMR spectrum of compound **6** in CD₃OD.



Figure S19. Extended ¹H NMR spectrum of compound **6** in CD₃OD.



Figure S21. HSQC spectrum of compound **6** in CD₃OD.

254