doi:10.15625/2525-2518/15442



CHEMICAL CONSTITUENTS FROM THE LEAVES OF ALCHORNEA RUGOSA (LOUR.) MÜLL. ARG. (EUPHORBIACEAE)

Pham Van Huyen, Nguyen Thi Thu Hien, Nguyen Huu Huong Duyen, Nguyen Thi Dieu Thuan, Nguyen Huu Toan Phan^{*}

Tay Nguyen Institute for Scientific Research, VAST, 116 Xo Viet Nghe Tinh, Da Lat, Viet Nam

^{*}Email: *nhtphan@gmail.com*

Received: 31 August 2020; Accepted for publication: 15 January 2022

Abstract. Alchornea is a genus of plants in the family Euphorbiaceae. It is widely distributed in tropical and subtropical regions of Africa, South Asia, Australia, and Latin America. Many species of the genus *Alchornea* have been used in traditional medicine in some countries around the world. Therefore, there have been many studies focusing on the chemical composition and biological activities of some species of the genus *Alchornea*. In the research project on the chemical composition of 4 species of *Alchornea* distributed in Viet Nam, we have reported compounds isolated from 3 species *A. tiliaefolia*, *A. annamatica*, and *A. trewioides*. *Alchornea rugosa* grows wild throughout Viet Nam and has been used in traditional medicine, but little is known about its chemical composition. From the leaves of species *A. rugosa* collected in Me Linh district, Ha Noi city, Viet Nam, five compounds including 3β-friedelanol (1), friedelan-3-one (2), methyl syringate (3), isovitexin (4), and rhoifolin (5) has been isolated from the methanol extract from its leaves by different chromatographic techniques. Their chemical structures were determined based on 1D and 2D NMR spectroscopy and mass spectrometry analysis and comparison with reference material. This is the first report on the isolation of these compounds from *A. rugosa*.

Keywords: Alchornea rugosa, 3β-friedelanol, friedelan-3-one, methyl syringate, isovitexin, rhoifolin

Classification numbers: 1.1.1, 1.1.6.

1. INTRODUCTION

The genus *Alchornea* belonging to the Euphorbiaceae family consists of 55 accepted species of deciduous, evergreen trees, or shrubs, which are distributed in many areas in the world. Some *Alchornea* species have been used in traditional medicine to cure edema, measles, acne boils, swelling, and to stop bleeding [1]. Previously, studies on chemical constituents of the *Alchornea* genus showed the presence of phenolic acids, flavonoids, terpenoids, sterols, and alkaloids [2 -5]. In Viet Nam, the plant *A. rugosa* (syn: *A. javanensis, A. petalostyla, A. hainanensis, Adelia glandulosa, etc.*; called Bo net or Sóc dai in Vietnamese) is widely distributed in nature from the North to the South and is used in folk medicine to treat enema and fever, as well as to enhance vitality [6 - 7]. To the best of our knowledge, little is known about the chemical constituents of this plant. Previous studies show that only a few alkaloids have

been isolated from its stem bark [8 - 9]. In this paper, we report the isolation and structural elucidation of five known compounds 1-5 from the leaves of *A. rugosa*.

2. EXPERIMENTS

2.1. General experimental procedures

NMR spectra were recorded on a Bruker AVANCE III HD spectrometer (Bruker, Billerica, MA, USA) using TMS as an internal standard. ESI-MS were measured on an Agilent 1100 LC/MS system. Column chromatography (CC) was carried out on silica gel (230 - 400 mesh, Merck), C18-reversed phase silica gel (100 Å pore size, Fluka), and Sephadex LH-20 gel (25 - 100 μ m, Pharmacia Fine Chemical Co. Ltd.). Thin-layer chromatography (TLC) was performed using pre-coated silica gel 60 F₂₅₄ (1.05554.0001, Merck) and RP-18 F_{254S} plates (1.15685.0001, Merck), and compounds were visualized by spraying with 10 % H₂SO₄ solution and heating for 3 - 5 min.

2.2. Plant material

The leaves of *Alchornea rugosa* (Lour) Muell. Arg. were collected in Me Linh, Ha Noi, Viet Nam in April 2016 and identified by Dr. Nguyen Quoc Binh from Vietnam National Museum of Nature, VAST. A voucher specimen (No.VTN/1024) is deposited at the Herbarium of Tay Nguyen Institute for Scientific Research, VAST, Viet Nam.

2.3. Extraction and isolation

The dried leaves of *A. rugosa* (4.4 kg) were extracted with MeOH 96 % (3×10 L) at room temperature. The combined methanol extracts were evaporated under reduced pressure to achieve a residue (716 g). The residue was suspended in water (2 L) and then partitioned in turn with *n*-hexane, chloroform, and ethyl acetate to obtain corresponding extracts: *n*-hexane (70 g, H), CHCl₃ (77 g, C), EtOAc (189 g, E), and water layer (2 L, W).

The extract H (70 g) was separated by chromatography column (CC) on a silica gel with stepwise gradient elution of *n*-hexane/EtOAc (1:0-0:1, v/v) to yield six fractions, H1-H6. Fraction H1 (24.6 g) was further separated by silica gel CC eluted with *n*-hexane/CH₂Cl₂ (1:1, v/v) to yield seven subfractions, H1.1-H1.7. Subfraction H1.2 (5.3 g) was separated by Sephadex LH-20 CC with MeOH/H₂O (3:1-1:0, v/v) to give four subfractions, H1.2.1-H1.2.4. Subfraction H1.2.4 (272 mg) was separated by silica gel CC eluted with CHCl₃/MeOH (5:1) and purified by silica gel CC using *n*-hexane/EtOAc (6:0.5, v/v) to afford compound **1** (25 mg). Fraction H3 (2.1 g) was separated by Sephadex LH-20 column eluted with MeOH/H₂O (1:1 - 1:0, v/v) to give five subfractions, H3.1-H3.5. Subfraction H3.5 (888 mg) was further separated on silica gel CC with CH₂Cl₂/acetone (50:1, v/v) to give four subfractions, H3.5.1-H3.5.4. Subfraction H3.5.1 (26 mg) was subjected to silica gel CC eluted with *n*-hexane/CH₂Cl₂ (1:3, v/v) and purified by RP-C18 column eluted with MeOH/H₂O (9:1, v/v) to yield compound **2** (12 mg).

The extract C (77 g) was separated on silica gel CC with stepwise gradient elution using CHCl₃/MeOH (1:0-0:1, v/v) to yield 6 fractions, C1-C6. Fraction C1 (4.8 g) was fractionated by silica gel CC with stepwise gradient elution with CH_2Cl_2 /acetone (1:0-0:1, v/v) to yield six subfractions, C1.1-C1.6. Subfraction C1.3 (706 mg) was purified by the RP-C18 column eluted with MeOH/H₂O (3:1, v/v) to give compound **3** (5 mg).

The water layer was passed through Diaion HP-20 CC and eluted first with water and then with MeOH-H₂O (1:3 - 1:0, v/v) to obtain five fractions, W1-W5. Fraction W4 (37 g) was separated on silica gel CC with stepwise gradient elution using CHCl₃/MeOH (1:0 - 0:1, v/v) to yield seven subfractions, W4.1-W4.7. Subfraction W4.3 (6.2 g) was subjected to chromatography on Sephadex LH-20 with MeOH/H₂O (1:4-1:0, v/v) to give four subfractions, W4.3.1-W4.3.4. Subfraction W4.3.3 (833 mg) was separated by silica gel CC with CHCl₃/MeOH (7:1, v/v) and purified by Sephadex LH-20 CC with MeOH/H₂O (1:4, v/v) to afford compounds **4** (8 mg) and **5** (10 mg).

3β-**Friedelanol** (1): white crystals, mp. 284.7 °C; molecular formula $C_{30}H_{52}O$, ESI-MS *m/z* 429.4 [M+H]⁺. ¹H-NMR (CDCl₃, 500 MHz) $\delta_{\rm H}$ (ppm): 1.55 (1H, m, H-1a), 1.42 (1H, m, H-1b), 1.90 (1H, dt, *J* = 2.5, 10.0 Hz, H-2a), 1.54 (1H, m, H-2b), 3.73 (1H, brs, H-3), 1.25 (1H, m, H-4), 1.74 (1H, dt, *J* = 3.0, 13.0 Hz, H-6a), 0.99 (1H, m, H-6b), 1.38 (2H, m, H-7), 1.27 (1H, m, H-8), 0.89 (1H, m, H-10), 1.38 (1H, m, H-11a), 1.22 (1H, m, H-11b), 1.32 (2H, m, H-12), 1.47 (1H, m, H-15a), 1.29 (1H, m, H-15b), 1.51 (1H, m, H-16a), 1.34 (1H, m, H-16b), 1.54 (1H, m, H-18), 1.44 (1H, m, H-19a), 1.13 (1H, dt, *J* = 4.5, 13.0 Hz, H-19b), 1.51 (1H, m, H-21a), 1.27 (1H, m, H-21b), 1.46 (1H, m, H-22a), 1.27 (1H, m, H-22b), 0.94 (3H, d, *J* = 7.5 Hz, H-23), 0.97 (3H, s, H-24), 0.86 (3H, s, H-25), 1.01 (3H, s, H-26), 0.99 (3H, s, H-27), 1.00 (3H, s, H-28), 0.95 (3H, s, H-29), 1.17 (3H, s, H-30). ¹³C-NMR (CDCl₃, 125 MHz) $\delta_{\rm C}$ (ppm): 15.81 (C-1), 35.23 (C-2), 72.77 (C-3), 49.21 (C-4), 37.13 (C-5), 41.76 (C-6), 17.57 (C-7), 53.22 (C-8), 38.40 (C-9), 61.39 (C-10), 35.36 (C-11), 30.66 (C-12), 37.86 (C-13), 39.70 (C-14), 32.36 (C-15), 36.11 (C-16), 30.04 (C-17), 42.86 (C-18), 35.58 (C-19), 28.19 (C-20), 32.85 (C-21), 39.30 (C-22), 11.62 (C-23), 16.41 (C-24), 18.26 (C-25), 18.65 (C-26), 20.13 (C-27), 31.81 (C-28), 35.03 (C-29), 32.10 (C-30).

Friedelan-3-one (2): white crystals, mp. 262 - 264 °C; molecular formula C₃₀H₅₀O, ESI-MS *m/z* 427.2 [M+H]⁺. ¹H-NMR (CDCl₃, 500 MHz) $\delta_{\rm H}$ (ppm): 1.69 (1H, dd, *J* = 5.0, 13.0 Hz, H-1a), 1.96 (1H, m, H-1b), 2.39 (1H, dd, *J* = 3.5, 13.5 Hz, H-2a), 2.30 (1H, dd, *J* = 6.0, 13.5 Hz, H-2b), 2.25 (1H, q, *J* = 6.5 Hz, H-4), 1.27 (1H, m, H-6a), 1.76 (1H, m, H-6b), 1.39 (1H, m, H-7a), 1.51 (1H, m, H-7b), 1.37 (1H, m, H-8), 1.55 (1H, m, H-10), 1.26 (1H, m, H-11a), 1.45 (1H, m, H-11b), 1.35 (2H, m, H-12), 1.27 (1H, m, H-15a), 1.47 (1H, m, H-15b), 1.58 (1H, m, H-16a), 1.35 (1H, m, H-16b), 1.56 (1H, m, H-18), 1.21 (1H, m, H-19a), 1.38 (1H, m, H-19b), 1.30 (1H, m, H-21a), 1.49 (1H, m, H-21b), 0.93 (1H, m, H-22a), 1.50 (1H, m, H-22b), 0.88 (3H, d, *J* = 6.5 Hz, H-23), 0.73 (3H, s, H-24), 0.87 (3H, s, H-25), 1.00 (3H, s, H-26), 1.01 (3H, s, H-27), 1.18 (3H, s, H-28), 1.05 (3H, s, H-29), 0.95 (3H, s, H-30). ¹³C-NMR (CDCl₃, 125 MHz) $\delta_{\rm C}$ (ppm): 22.3 (C-1), 41.55 (C-2), 213.17 (C-3), 58.26 (C-4), 42.17 (C-5), 41.34 (C-6), 18.27 (C-7), 53.14 (C-8), 37.49 (C-9), 59.53 (C-10), 35.66 (C-11), 30.53 (C-12), 39.73 (C-13), 38.33 (C-14), 32.82 (C-15), 36.05 (C-16), 30.03 (C-17), 42.85 (C-18), 35.38 (C-19), 28.19 (C-20), 32.46 (C-21), 39.28 (C-22), 6.83 (C-23), 14.68 (C-24), 17.96 (C-25), 20.27 (C-26), 18.67 (C-27), 32.12 (C-28), 35.03 (C-29), 31.80 (C-30).

Methyl syringate (3): colourless needles, mp. 103-107 °C, molecular formula $C_{10}H_{12}O_5$, ESI-MS *m/z* 212.9 [M+H]⁺. ¹H-NMR (500 MHz, CDCl₃) $\delta_{\rm H}$ (ppm): 7.32 (2H, s, H-2, H-6), 5.88 (1H, brs, 4-OH), 3.94 (6H, s, 3-OMe; 5-OMe), 3.90 (3H, s, 7-OMe). ¹³C-NMR (CDCl₃, 125 MHz) $\delta_{\rm C}$ (ppm): 121.15 (C-1), 106.72 (C-2, C-6), 146,67 (C-3, C-5), 139.25 (C-4), 166.87 (C-7), 56.46 (3-OMe, 5-OMe), 52.09 (7-OMe).

Isovitexin (4): yellow amorphous powder, mp. 220-221 °C, molecular formula $C_{21}H_{20}O_{10}$. ESI-MS: $m/z = 433.0 \text{ [M+H]}^+$, 431.0 [M-H]^- . ¹H-NMR (500 MHz, CD₃OD) δ_H (ppm): 7.86 (2H, d, J = 8.5 Hz, H-2', H-6'), 6.95 (2H, d, J = 8.5 Hz, H-3', H-5'), 6.61 (1H, s, H-3), 6.51 (1H, s, H-3) 8), 4.93 (1H, d, J = 9.5 Hz, H-1''), 4.20 (1H, dd, J = 9.0, 9.5 Hz, H-2''), 3.50 (1H, m, H-3''), 3.53 (1H, m, H-4''), 3.45 (1H, m, H-5''), 3.89 (1H, dd, J = 2.0, 12.0 Hz, H_a-6''), 3.79 (1H, dd, J = 5.5, 12.0 Hz, H_b-6''). ¹³C-NMR (125 MHz, CD₃OD) $\delta_{\rm C}$ (ppm): 166.11 (C-2), 103.81 (C-3), 183.95 (C-4), 162.84 (C-5), 109.36 (C-6), 166.11 (C-7), 95.55 (C-8), 159.00 (C-9), 105.00 (C-10), 123.17 (C-1'), 129.42 (C-2'), 117.08 (C-3'), 162.84 (C-4'), 117.08 (C-5'), 129.42 (C-6'), 75.45 (C-1''), 72.52 (C-2''), 80.2 (C-3''), 71.64 (C-4''), 82.56 (C-5''), 62.64 (C-6'').

Rhoifolin (5): yellow amorphous powder, mp. 245 °C, molecular formula $C_{27}H_{30}O_{14}$, ESI-MS *m/z* 601.0 [M+Na]⁺. ¹H-NMR (500 MHz, CD₃OD) $\delta_{\rm H}$ (ppm): 7.91 (2H, d, *J* = 8.5 Hz, H-2', H-6'), 6.96 (2H, d, *J* = 8.5 Hz, H-3', H-5'), 6.81 (1H, d, *J* = 2.0 Hz, H-8), 6.68 (1H, s, H-3), 6.49 (1H, d, *J* = 2.0 Hz, H-6), 5.22 (1H, d, *J* = 8.0 Hz, H-1''), 3.72 (1H, m, H-2''), 3.66 (1H, m, H-3''), 3.63 (1H, m, H-4''), 3.57 (1H, m, H-5''), 3.74/ 3.64 (2H, m, H_a-6'', H_b-6'), 5.30 (1H, d, *J* = 1.5 Hz, H-1''), 3.45 (1H, m, H-2'''), 3.98 (1H, m, H-3''), 3.43 (1H, m, H-4'''), 3.96 (1H, m, H-5'''), 1.35 (3H, d, *J* = 6.5 Hz, H-6''). ¹³C-NMR (125 MHz, CD₃OD) $\delta_{\rm C}$ (ppm): 166.85 (C-2), 104.15 (C-3), 184.09 (C-4), 162.92 (C-5), 101.03 (C-6), 164.41 (C7), 95.97 (C-8), 159.01 (C-9), 107.0 (C-10), 123.08 (C-1'), 129.66 (C-2'), 117.09 (C-3'), 162.92 (C-4'), 117.09 (C-5'), 129.66 (C-6'), 99.84 (C-1''), 79.16 (C-2''), 78.97 (C-3''), 72.2 (C-4''), 78.27 (C-5''), 62.41 (C-6''), 102.54 (C-1''), 71.38 (C-2'''), 72.2 (C-3'''), 73.94 (C-4'''), 70.03 (C-5'''), 18.22 (C-6''').

3. RESULTS AND DISCUSSION

Compound **1** was isolated as white crystals. Its molecular formula, $C_{30}H_{52}O$, was determined by the ESI-MS quasi-molecular ion peak at m/z 429.4 [M+H]⁺. The ¹³C-NMR and DEPT spectrum indicated that **1** had 30 carbons including eight methyls, eleven methylenes, four sp³ methines, one oxygenated methine, and six quaternary sp³ carbons. The ¹H-NMR spectra of **1** also showed signals for eight methyl groups at $\delta_H 0.94$ (d, J = 7.5 Hz, H-23), 0.97 (s, H-24), 0.86 (s, H-25), 1.01 (s, H-26), 0.99 (s, H-27), 1.00 (s, H-28), 0.95 (s, H-29), and 1.17 (s, H-30). The above data suggested that **1** was a friedelane-type triterpenoid. The presence of a signal at $\delta_H 3.73$ indicated an oxymethine proton. Furthermore, HMBC correlations observed from H-2 ($\delta_H 1,90/1,54$) to C-1 ($\delta_C 15.81$), C-3 ($\delta_C 72.77$), and C-4 ($\delta_C 49.21$), from H-3 ($\delta_H 3.73$) to C-4 and C-23 ($\delta_C 11.62$), from H-23 to C-3, C-4, and C-5 ($\delta_C 37.13$) allowed to confirm an oxymethine proton at C-3 position. Compound **1** was thus identified as 3 β -friedelanol and the spectroscopic data compared well with those previously reported [10].

Compound **2** was obtained as white crystals. Its molecular formula, $C_{30}H_{50}O$, was determined by the ESI-MS quasi-molecular ion peak at m/z 427.2 [M+H]⁺. Detailed analysis of the ¹³C-NMR and HSQC spectra revealed the presence of 30 carbon signals, including eight methyls, eleven methylenes, four sp³ methines, one carbonyl carbon, and six quaternary sp³ carbons. Comparison of the ¹H- and ¹³C-NMR data of **2** with those of **1** indicated that the structures of both compounds were similar, except for the replacement of the hydroxymethine group (δ_C 72.77) in **1** with a carbonyl group (δ_C 213.17) in **2**. By comparison of the NMR data of **2** with those of the published data [11], **2** was identified as friedelan-3-one.

Compound **3** was isolated as colorless needle. The ¹H-NMR of **3** showed the signals of two protons [$\delta_{\rm H}$ 7.32 (2H, s, H-2 and H-6)] in a tetrasubstituted benzene and three methoxyl groups [$\delta_{\rm H}$ 3.90 (3H, s, 7-OCH₃) and 3.94 (6H, s, 3-OCH₃ and 5-OCH₃)]. The ¹³C-NMR spectrum combined with the HSQC and HMBC spectral showed signals of 10 carbons, including one carbonyl ester ($\delta_{\rm C}$ 166.87, C-7), three methoxy groups [$\delta_{\rm C}$ 52.09 (7-OCH₃), 56.46 (3-OCH₃ and 5-OCH₃)], and six carbons in the aromatic region [$\delta_{\rm C}$ 121.15 (C-1), 106.72 (C-2, C-6), 146.67

(C-3, C-5), and 139.25 (C-4)]. Accordingly, the structure of **3** was elucidated as methyl syringate by comparison with the spectral data in the literature [12].

Compound **4** was isolated as a yellow amorphous powder. The molecular formula was established as $C_{21}H_{20}O_{10}$ by ESI-MS data ([M+H]⁺ m/z 433.0 and [M-H]⁻ m/z 431.0). The ¹H-NMR spectrum of **4** displayed signals of four AA'BB'-type protons [δ_H 7.86 (2H, d, J = 8.5 Hz, H-2', H-6') and 6.95 (2H, d, J = 8.5 Hz, H-3', H-5')], one aromatic proton as a singlet at δ_H 6.51 (H-8), one olefinic proton at δ_H 6.61 (H-3). Therefore, the aglycon of **4** was identified as apigenin. The ¹³C-NMR spectrum of **4** presented signals of 21 carbons, of which 15 carbons were assigned to the flavone aglycone and 6 carbons assigned to the sugar moiety. Its NMR features were also indicative for a β -D-glucopyranose [δ_C 75.45 (CH, C-1"), 72.52 (CH, C-2"), 80.2 (CH, C-3"), 71.64 (CH, C-4"), 82.56 (CH, C-5") and 62.64 (CH₂, C-6")/ δ_H 4.93 (1H, d, J = 9.5 Hz, H-1"), 4.20 (1H, dd, J = 9.0, 9.5 Hz, H-2"), 3.50 (1H, m, H-3"), 3.53 (1H, m, H-4"), 3.45 (1H, m, H-5"), 3.79 (1H, dd, J = 5.5, 12.0 Hz, H_a-6") and 3.89 (1H, dd, J = 2.0, 12.0 Hz, H_b-6 ")]. The HMBC spectrum showed correlations between anomeric proton at δ_H 4.93 (H-1") of Glc and carbon C-6 at δ_C 109.36 of apigenin, which suggested the glycosylation at C-6 of apigenin skeleton. Based on the NMR spectroscopic data and comparison with literature data [13] the structure of **4** was established as isovitexin.

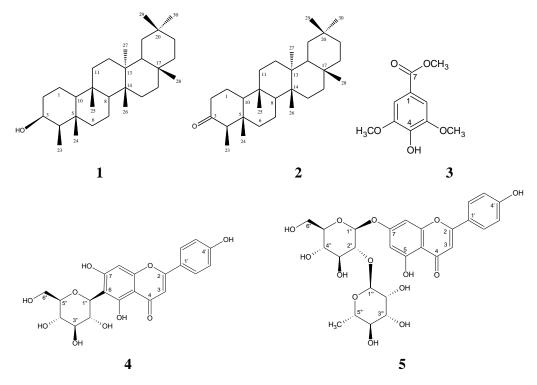


Figure 1. Chemical structures of compounds 1-5.

Compound **5** was isolated as a yellow amorphous powder. The NMR data of **5** revealed the presence of apigenin aglycone and two sugar moieties. The ¹H NMR spectra of **5** showed the signals of four AA'BB'-type protons [$\delta_{\rm H}$ 7.91 (2H, d, J = 8.5 Hz, H-2', H-6') and 6.96 (2H, d, J = 8.5 Hz, H-3', H-5')], two AX-type aromatic protons [$\delta_{\rm H}$ 6.49 (1H, d, J = 2.0 Hz, H-6) and 6.81 (1H, d, J = 2.0 Hz, H-8)], and one olefinic proton at $\delta_{\rm H}$ 6.68 (1H, s, H-3). Moreover, the signals

of two anomeric protons at $\delta_{\rm H}$ 5.22 (1H, d, J = 8.0 Hz, H-1") and 5.30 (1H, d, J = 1.5 Hz, H-1") were assigned to β -D-glucose (Glc) and α -L-rhamnose (Rha) units, respectively. The ¹³C-NMR and DEPT spectra of **5** displayed signals of 27 carbons, including 15 carbons of the aglycone and 12 carbons of the sugar moieties. In the HMBC spectrum, H-1" was shown to correlate with C-2" ($\delta_{\rm C}$ 79.16), indicating that the α -L-rhamnose was located at the C-2" position of the glucose moiety. Besides, the HMBC correlation from H-1" ($\delta_{\rm H}$ 5.22) to C-7 ($\delta_{\rm C}$ 164.41) confirmed the glycosylation at C-7 of aglycon. Based on the spectroscopic evidences and comparison with the reported values in the literature [14], compound **5** was identified as rhoifolin.

4. CONCLUSIONS

From the MeOH extract from the leaves of *Alchornea rugosa*, using various chromatography separations, five compounds 3β -friedelanol (1), friedelan-3-one (2), methyl syringate (3), isovitexin (4), and rhoifolin (5) were isolated. Their structures were elucidated by 1D and 2D NMR spectroscopic interpretation. This is the first report on the chemical constituents of the species collected in Viet Nam.

Acknowledgment. This research is funded by the Vietnam National Foundation for Science and Technology Development (NAFOSTED) under grant number 104.01-2015.39.

CRediT authorship contribution statement. Author 1: Experimental performance, Methodology, Data analysis, Manuscript preparation. Author 2: Experimental performance, Methodology, Formal analysis. Author 3: Experimental performance, Formal analysis. Author 4: Experimental performance, Methodology, Formal analysis. Author 5: Manuscript revision, Supervision.

Declaration of competing interest. The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

REFERENCES

- 1. Ly T. D. 1900 useful plant species in Viet Nam, World Publisher, Ha Noi, 1993, (in Vietnamese).
- 2. Ogungbamila F. O. and Samuelsson G. Smooth muscle relaxing flavonoids from *Alchornea cordifolia*, Acta Pharm. Nord. **2** (6) (1990) 421-422. PMID: 2095800.
- Okoye F. B. and Osadebe O. P. A new anti-inflammatory flavonol glycoside from *Alchornea floribunda* leaves, Nat. Prod. Res. 24 (3) (2010) 266-273. https://doi.org/ 10.1080/14786410902986894.
- Okoye F. B., Osadebe P. O., Proksch P., Edrada-Ebel R. A., Nworu C. S., and Esimone C. O. Anti-inflammatory and membrane-stabilizing stigmastane steroids from *Alchornea floribunda* leaves, Planta Med. **76** (2) (2010) 172-177. https://doi.org/10.1055/s-0029-1186032.
- 5. Cesar A. M., Oscar M. M., Jaime N. Medicinal plants from the genus Alchornea (Euphorbiaceae): A review of their ethnopharmacology uses and phytochemistry, Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas **16** (3) (2017) 162-205.
- 6. Ho P. H. An Illustrated Flora of Viet Nam, Volume 2, Youth publisher, Ho Chi Minh 1999, (in Vietnamese).
- 7. http://theplantlist.org/tpl1.1/record/kew-5835 (accessed 14th November 2020).

- Hart N. K., Johns S. R., Lambert J. A. Hexahydroimidazo-pyrimidines, a new class of alkaloids from Alchomea javanensis, J. Chem. Soc. D (1969) 1484-1485. https://doi.org/ 10.1039/C29690001484.
- Hart N. K., Johns S. R., Lamberton H. A., and Willing R. I. Alkaloids of *Alchornea javanensis* (Euphorbiaceae): The isolation of hexahydroimidazo[1,2-a]pyrimidines and guanidines, Aust. J. Chem. 23 (8) (1970) 1679-1693. https://doi.org/10.1071/CH9701679.
- Kundu J. K., Rouf A. S. S., Nazmul H. M., Hasan C. M., and Rashid M. A. Antitumor activity of epifriedelanol from *Vitis trifolia*, Fitoterapia **71** (2000) 577-579. https://doi.org/ 10.1016/s0367-326x(00)00191-x.
- Vu K. T., Le T. K. A., Dang N. Q., Nguyen V. T., Hoang L. T. A., Nguyen X. N., Dan T. T. H., Chau V. M., Phan V. K. Triterpenes from the leaves of *Glochidion obliquum*. Vietnam J. Chem. **53** (2e) (2015) 103-106. https://doi.org/10.15625/0866-7144.2015-2e-024.
- Xian Y. -X., Zhou H. -L., Wang X., Yu J. -Q., Zheng Z. -J., and Yang B. -T. Chemical constituents of *Gleditsia sinensis* Thorns, Asian J. Chem. 27 (3) (2015) 1063-1065. https://doi.org/10.14233/ajchem.2015.18240.
- Ling T. -J., Ling W. -W., Chen Y. -J., Wan X. -C., Xia T., Du X. -F., and Zhang Z. -Z. -Antiseptic activity and phenolic constituents of the aerial parts of *Vitex negundo var*. *cannabifolia*, Molecules **15** (2010) 8469-8477. https://doi.org/10.3390/molecules 15118469.
- 14. Kiem P. V., Minh C. V., Cuong N. X., Hang D. T. T., Thao N. P., Nam N. H., Ban N. K., and Hai T. N. Study of components flavonoids and megastigmane glucosides of *Ficus callosa*, Vietnam J. Chem. **49** (1) (2011) 55-59.